EDITORIAL VIEWS

Secchi R, Sutton J, Eglen RM: Assessment of the role of alpha2-adrenoceptor subtypes in the antinociceptive, sedative and hypothermic action of dexmedetomidine in transgenic mice. Br J Pharmacol 1997; 122:1339-44

- 5. Lakhlani PP, MacMillan LB, Guo TZ, McCool BA, Lovinger DM, Maze M, Limbird: Substitution of a mutant $\alpha 2a$ -adrenergic receptor via "hit and run" gene targeting reveals the role of this subtype in sedative, analgesic, and anesthetic-sparing responses in vivo. Proc Natl Acad Sci U S A 1997; 94:9950 5
- 6. Stone LS, MacMillan LB, Kitto KF, Limbird LE, Wilcox GL: The $\alpha 2a$ adrenergic receptor subtype mediates spinal analgesia evoked by $\alpha 2$ agonists and is necessary for spinal adrenergic-opioid synergy. J Neurosci 1997; 17:7157–65
- 7. MacMillan LB, Hein L, Smith MS, Piascik MT, Limbird LE: Central hypotensive effects of the $\alpha 2a$ -adrenergic receptor subtype. Science 1996; 273:801–3
- 8. Link RE, Desai K, Hein L, Stevens MS, Chruscinski A, Bernstein D, Barash GS, Kobilka BK: Cardiovascular regulation in mice lacking α 2-adrenergic receptor subtypes b and c. Science 1996; 273:803-5
- 9. Sallinen J, Haapalinna A, Viitamaa T, Kobilka BK, Scheinin M: D-Amphetamine and L-5-hydroxytryptophan-induced behaviours in mice with genetically-altered expression of the α 2C-adrenergic receptor subtype. Neuroscience 1998; 86:959–65

- 10. Aley KO, Levine JD: Multiple receptors involved in peripheral $\alpha 2$, μ , and A_1 antinociception, tolerance, and withdrawal. J Neurosci 1997; 17:735–44
- 11. Kieffer BL: Opioids: First lessons from knockout mice. Trends Pharmacol Sci 1999; 20:19-26
- 12. Mizobe T, Maghsoudi K, Sitwala K, Tianzhi G, Ou J, Maze M: Antisense technology reveals the a2 adrenoceptor to be the subtype mediating the hypnotic response to the highly selective agonist, dexmedetomidine, in the locus coeruleus of the rat. J Clin Invest 1996; 98:1076–80
- 13. Rosin DL, Talley EM, Lee A, Stornetta RL, Gaylinn BD, Guyenet PG, Lynch KR: Distribution of α 2C-adrenergic receptor-like immunoreactivity in the rat central nervous system. J Comp Neurol 1996; 372:135-65
- 14. Winzer-Serhan UH, Leslie FM: α 2B-Adrenoceptor mRNA expression during rat brain development. Brain Res Dev Brain Res 1997; 100:90-100
- 15. Smith MS, Schambra UB, Wilson KH, Page SO, Hulette C, Light AR, Schwinn DA: alpha 2-Adrenergic receptors in human spinal cord: specific localized expression of mRNA encoding alpha 2-adrenergic receptor subtypes at four distinct levels. Brain Res Mol Brain Res 1995; 34:109-17

Anesthesiology 2000; 92:936 - 8 © 2000 American Society of Anesthesiologists, Inc. Lippincott Williams & Wilkins, Inc.

Ion Channels Take Center Stage

Twin Spotlights on Two Anesthetic Targets

ONE of the fundamental ideas behind the science of anesthesiology throughout the majority of the twentieth century was the idea that there exists a "unitary site" of action for all general anesthetics. As our knowledge of the underlying neurobiology has grown, together with the database of potential anesthetic target sites, it has become increasingly obvious that this simplistic notion is incorrect. It now seems unlikely that general anes-

•

This Editorial View accompanies the following article: de Sousa SLM, Dickinson R, Lieb WR, Franks NP: Contrasting synaptic actions of the inhalational general anesthetics isoflurane and xenon. Anesthesiology 2000; 92:1055–66.

Accepted for publication November 19, 1999.

Key words: Binding; GABA; isoflurane; NMDA; Xenon.

The author is not supported by, nor maintains any financial interest in, any commercial activity that may be associated with the topic of this article. thetics interact with a single common target site because the function of a variety of membrane proteins has been shown to be altered within the clinically relevant range of anesthetic concentrations.³ Not only do multiple potential anesthetic targets exist, but the array of susceptible targets varies among different classes of anesthetic (review of Krasowski and Harrison⁴). For example, clinical concentrations of pentobarbital inhibit depolarization mediated via AMPA- and kainate-type glutamate receptors, enhance and prolong γ-aminobutyric acid (GABA)-mediated inhibition via an action at GABAA receptors, and inhibit the function of neuronal nicotinic acetylcholine receptors (n-nAChRs), whereas ketamine has no effect at GABAA receptors, but inhibits the function of the N-methyl-D-aspartate (NMDA) subtype of glutamate receptors (reviews of Franks and Lieb3 and Krasowski and Harrison⁴).

As the century draws to a close, this "multiple alternate target" hypothesis gains further support from a study published in this issue of Anesthesiology.⁵ In this study,

Sara de Sousa et al.,5 from Nick Franks' laboratory in London, compare the synaptic actions of the everyday inhaled anesthetic isoflurane with those of the more exotic noble gas xenon. Although isoflurane is easy to obtain and study, the expense and lack of potency of xenon have long been obstacles to the study of its mechanism of action. Yet, in evaluating the various hypotheses of anesthetic mechanism, it is desirable to study a variety of anesthetic structures to test the general applicability of a potential mechanism. Xenon is a monoatomic inert gas and therefore seems like an unpromising substance for which to seek selective actions. Remarkably. Franks et al. has shown that xenon, applied at approximately 80% atm to cultures of rat hippocampal neurons, inhibits currents through the NMDA receptor, but fails to alter the function of GABAA receptors.

In the de Sousa *et al.* article,⁵ the authors point out the stark contrast between the actions of isoflurane and xenon. Isoflurane, at 1 minimum alveolar concentration (MAC), was found to increase the duration of inhibitory postsynaptic currents, while causing a small decrease in the amplitude of excitatory postsynaptic currents. The increase in inhibitory postsynaptic currents duration is consistent with the actions of isoflurane on postsynaptic GABA_A receptors, whereas the decrease in amplitude of excitatory postsynaptic currents probably reflects presynaptic actions of the volatile anesthetics.^{6,7} Conversely, xenon had no effect on the inhibitory postsynaptic current, but selectively reduced the slow component of the excitatory postsynaptic current that is mediated by NMDA receptors.⁵

These differences between two simple anesthetic gases at the level of molecular and cellular targets may seem surprising at first and are certainly at odds with the unitary models that dominated thinking in this field for so long. In fact, the discrepancy is less surprising when one considers the pharmacologic profile of the two anesthetics. Isoflurane produces hypnosis and unconsciousness and depresses spinal reflexes, yet confers little analgesia. Xenon, however, is an excellent analgesic and has the ability to produce hypnosis and amnesia. The pharmacologic profile of xenon anesthesia is very similar to that of ketamine, another known antagonist of glutamate at NMDA receptors. The analgesic and amnestic actions of xenon and ketamine are shared by other NMDA receptor antagonists and fit well with what is known about the anatomic distribution and physiologic functions of this ligand-gated ion channel.⁵

So much for the cellular and synaptic pharmacology of isoflurane and xenon. But what about the molecular

level? How can such selectivity between ligand-gated ion channels be exhibited by simple gaseous anesthetics? The answer may lie in the differences of molecular structure between the four transmembrane domain GABA, receptor subunits and the three transmembrane domain NMDA receptor subunits (which appear to be members of a distinct gene superfamily among the receptor molecules) and in the existence of anesthetic-binding pockets or cavities within these target molecules. Recent work using site-directed mutagenesis has shown that specific mutations at serine 270 in the GABA_A receptor α subunit can alter the sensitivity of the receptor to enflurane and isoflurane.^{8,9} A new article suggests that serine 270 might be part of a hypothetical binding pocket of defined volume for anesthetic ethers, ¹⁰ located between adjacent transmembrane domains within each receptor subunit polypeptide.

Are anesthetic binding pockets merely fanciful inventions of molecular pharmacologists? Apparently not, because the existence and precise location and dimensions of an anesthetic binding cavity has been demonstrated using X-ray crystallography in firefly luciferase. The binding of anesthetics within such pockets, although necessarily of low affinity, would be driven by a combination of enthalpic and entropic free-energy changes and hence be governed by the customary laws of thermodynamics. The such pockets are considered by the customary laws of thermodynamics.

If one accepts for the moment the premise that such binding pockets exist within these ion channels, it follows that xenon does not bind well within the anesthetic ether pocket associated with the GABA_A receptor. This might reflect the inappropriate size or shape of the xenon atom, or perhaps an inability to participate in hydrogen-bonding interactions. Apparently the NMDA receptor is also selective, admitting xenon but excluding isoflurane. The search surely will now be on for the molecular determinants of the actions of xenon on the NMDA receptor, and for further clues concerning the lack of interaction of the noble gas with the GABA_A receptor.

The study by de Sousa *et al.*⁵ therefore provides a satisfying conclusion to the discussions concerning unitary mechanisms of anesthesia. The unitary hypothesis has clearly outlived its usefulness; but all is not lost in terms of understanding. The illumination provided by this monochromatic concept has indeed been diffracted across a rainbow of molecular targets in recent years, but may now be refocussed to throw the spotlight onto two molecular stars of the synaptic stage: the GABA_A and NMDA receptors.

Neil L. Harrison, Ph.D.

Professor and Director C. V. Starr Laboratory of Molecular Neuropharmacology Department of Anesthesiology Weill Medical College of Cornell University New York, New York 10021 neh2001@mail.med.cornell.edu

References

- 1. Meyer HH: Contribution to the theory of narcosis. Trans Faraday Soc 1937; 33:1062-8
- 2. Collins JF, Kendig JJ, Mason P: Anesthetic actions within the spinal cord: Contributions to the state of general anesthesia. Trends Neurosci 1995;18:549-53
- 3. Franks NP, Lieb WR: Molecular and cellular mechanisms of general anesthesia. Nature 1994; 367:607-14
- 4. Krasowski MD, Harrison NL: General anaesthetic actions on ligand-gated ion channels. Cell Molec Life Sci 1999; 55:1278-303
- 5. De Sousa SLM, Dickinson R, Lieb WR, Franks NP: Contrasting synaptic actions of the inhalational general anesthetics isoflurane and xenon. Anesthesiology 2000; 92:1055-66
 - 6. MacIver MB, Mikulec AA, Amagasu SM, Monroe FA: Volatile an-

esthetics depress glutamate transmission via presynaptic actions. ANESTHESIOLOGY 1995; 85:823-4

- 7. Schlame M, Hemmings HC Jr: Inhibition by volatile anesthetics ofendogenous glutamate release from synaptosomes by a presynaptic mechanism. Anesthesiology 1995; 82:1406-16
- 8. Mihic SJ, Ye Q, Wick M, Koltchine VV, Finn SE, Krasowski MD, Hanson KK, Mascia MP, Valenzuela CF, Greenblatt EP, Harris RA, Harrison NL: Sites of alcohol and volatile anaesthetic action on GABA_A and glycine receptors. Nature 1997; 389:385-9
- 9. Krasowski MD, Koltchine VV, Rick CEM, Ye Q, Finn SE, Harrison NL: Propofol and other intravenous anesthetics have sites of action on the γ -aminobutyric acid-A receptor distinct from that for isoflurane. Mol Pharmacol 1998; 53:530 8
- 10. Koltchine VV, Finn SE, Jenkins A, Nikolaeva N, Lin A, Harrison NL: Agonist gating and isoflurane potentiation in the human $GABA_A$ receptor determined by the volume of a TM2 residue. Mol Pharmacol 1999; 56:1087-93
- 11. Franks NP, Conti E, Jenkins A, Lieb WR, Brick P: Structural basis for the inhibition of firefly luciferase by a general anaesthetic. Biophys J 1998; 75:2205-11
- 12. Jenkins A, Franks NP, Lieb WR: Effects of temperature and volatile anesthetics on ${\rm GABA_A}$ receptors. Anisthesiology 1999; 90:484-91