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In Reply:—We appreciate the comments by Dr. Sheng-Nan Wu. He raises three important points; (1) the ability of isoproterenol to induce Ca²⁺ influx into endothelium, (2) inhibitory effect of halothane on Ca²⁺ ionophore (A23187)-induced endothelium dependent relaxation, and (3) the possibility of modification of the relaxation response to isoproterenol by Ca²⁺ channel blocking action of halothane. We discuss these points, respectively.

From our findings and other reports (please see the article), it is likely that isoproterenol can not initiate nitric oxide (NO) production by vascular endothelium of other species except for rat and that adenylyl cyclase-mediated signal transduction within endothelium of other species differs from that of rat. From experiments using various inhibitors, we propose that cAMP-PKA play an important role in the activation of Ca²⁺/calmodulin dependent NO synthase (cNOS) elicited by isoproterenol. The calcium ionophore, A23187, can increase intracellular Ca²⁺ in the endothelium not only via Ca²⁺ influx but also via release from intracellular store. The latter mechanism has been demonstrated to be essential for A23187-induced NO production by endothelium.^{1,2} Thus, it is likely that halothane exerts the inhibitory action on NO-cGMP relaxation mechanism only when cNOS is activated by Ca²⁺ releasing mechanism.

Halothane has been shown to modulate Ca2+ dynamics in vascular smooth muscle cells including the inhibition of Ca2+ influx and the facilitation of Ca2+ releasing from sarcoplasmic reticulum (caffeinelike action).3 These dual actions of halothane on Ca2+ dynamics are likely to induce the net effects on vascular tone within a few minutes. Our experimental protocols basically comprised an initial equilibration period followed by elevation of vascular tone with noradrenaline and subsequent test of relaxation response of rings with sustained tone to isoproterenol in the absence or presence of various inhibitors or halothane. Aortic rings were exposed to halothane for more that 40 min, during which the modulation of halothane to Ca2+ dynamics might reach plateau. Therefore, isoproterenol was applied to the rings with sustained tone and Ca2+ dynamics in the presence of halothane. Further, our study demonstrated that isoproterenol produced endothelium-dependent and NO-mediated relaxation accompanying with the increases in cGMP contents of rat aortic rings and that halothane failed to affect not only the relaxation response but also the cGMP increase elicited by isoproterenol. It has been known that the post-cGMP cascade mediating relaxation of vascular smooth

muscle cells includes the modulations of contractile proteins and of various ion currents (Na⁺, K⁺, Cl⁻ and Ca²⁺), however, the contribution of L-type Ca²⁺ inward current to it has not been well known. Even if halothane has an ability to inhibit the L-type of Ca²⁺ inward current through the membrane of vascular smooth muscle cells as shown in the short report by Dr. Wu *et al.*, ⁴ in our opinion, it seems unlikely that this inhibitory action accounts for the failure of halothane to inhibit the isoproterenol-induced relaxation.

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