## I EDITORIAL VIEWS

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## Complex Pharmacology of Malignant Hyperthermia

Three articles in this issue of ANESTHESIOLOGY concern malignant hyperthermia (MH), providing new insights into the complexity of this disease. First, in a surprising in vitro lipid bilayer study of skeletal muscle sarcoplasmic reticulum (SR), a very low (nm) concentration of dantrolene was found to open the isolated Ca2+ release channel, whereas a higher (µM) concentration caused the expected blockade of the channel.1 In a second paper, stimulation of the Ca2+ release channel by a common drug preservative (i.e., chlorocresol) caused contractures in intact muscle fibers in vitro.2 Finally, stimulation of the Ca2+ release channel by a serotonin agonist, as measured by biopsied muscle in vitro, resulted in contracture.3 These findings help to expand our view of MH myopathies and enhance our understanding of some of the complex drug effects on the Ca2+ release channel of skeletal muscle and its associated proteins

It is useful to review the subcellular basis and molecular machinery responsible for excitation-contraction coupling in skeletal muscle cells (fig. 1).4 The 10,000-fold inwardly directed Ca<sup>2+</sup> gradient across the sarcolemma of resting skeletal muscle cells is maintained by a large array of Ca2+-ATPases that sequester the ion into the lumen of the SR. Elevation of cytoplasmic Ca<sup>2+</sup> is orchestrated by the activation of voltage-operated Ca<sup>2+</sup> channels of the plasma membrane<sup>5,6</sup> and ryanodine-sensitive Ca2+ channels of SR7,8 that are juxtaposed where the surface membrane transverse tubule meets the junctional SR membrane (the triad). 9,10 The plant alkaloid ryanodine binds to the SR Ca2+ release channel with multiple allosterically coupled affinities. 11-13 Ryanodine has been essential in identifying the junctional "foot" structure that spans the transverse tubule/SR junction as the Ca2+ release channel protein, 10 now commonly referred to as the ryanodine (Ry) receptor.8,14 Each Ry receptor is composed of four identical monomers of approximately 5,000 amino acids. Ry receptor monomers are encoded by at least three distinct genes, 4,15 of which Ry<sub>1</sub> receptor is the isoform expressed by skeletal muscle. 16,17,\* When the transverse tubule depolarizes, charge movement within the skeletal muscle dihydropyridine-sensitive L-type Ca2+ channel (DHP receptor) activates the Ry<sub>1</sub> receptor to open, and Ca<sup>2+</sup> is released into the myoplasm, causing contraction. Although the mechanism coupling the DHP receptor and the Ry<sub>1</sub> receptor has yet to be fully defined, the intracellular loop between repeats II and III of the  $Ca^{2+}$  channel  $\alpha_1$  subunit (fig. 1) is essential for excitation-contraction coupling.6,18

Ry receptor is an enormous protein complex modulated by physiologic ligands including Ca2+, Mg2+, adenine nucleotides, calmodulin, and sphingosine. 18-20 In addition to such intracellular mediators, cellular pathways that result in phosphorylation of Ry, receptors may influence the gating and response of the release channel.21 A wide variety of drugs also can modulate Ry<sub>1</sub> receptor gating and SR Ca<sup>2+</sup> release.<sup>15</sup> The volatile anesthetics are noteworthy among the drugs that may increase Ry<sub>1</sub> receptor activity and perturb Ca<sup>2+</sup> regulation, 22 especially in MH-susceptible individuals. 23,24 Normal skeletal muscle may tolerate a modest perturbation of Ca<sup>2+</sup> homeostasis (e.g., increased Ca<sup>2+</sup> "leak" from the SR) without evidence of a problem. However, various myopathic skeletal muscle disorders that have a "leaky" membrane due to a variety of causes may be compensated normally until perturbed by an agent that further activates Ca2+ release and further strains homeostatic mechanisms. Certain combinations of drug and disease can result in a smoldering reaction that may abate or intensify. The wrong combination of susceptibility and heterogeneous triggers creates a potentially explosive response, so that the sleeping giant of skeletal muscle metabolism overwhelms whole body homeostasis. In addition, triggering of hypermetabolism partly depends on dose of triggering agent, i.e., smaller concentrations trigger more slowly, and larger concentrations are decidedly quicker.<sup>25</sup> Consequently,

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\* According to International Union of Pharmacology Committee on Drug Classification and Receptor Nomenclature (December 1995), the ryanodine receptor isoforms from skeletal muscle, heart, and brain are denoted as Ry<sub>1</sub>, Ry<sub>2</sub>, and Ry<sub>3</sub> receptors, respectively.

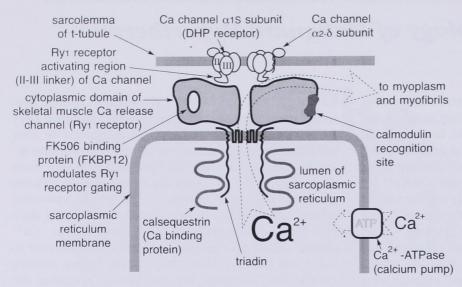


Fig. 1. The major components of the excitation-contraction pathway of skeletal muscle. The major feature is the Ca2+ release channel (Ry1 receptor) of the junctional sarcoplasmic reticulum (JSR); two of the four monomers that compose the Ca2+ release channel are shown in crosssection. A voltage-dependent conformational change in the skeletal muscle Ca2+ channel (DHP receptor,  $\alpha_{18}$  subunit) is transmitted to the Ry1 receptor through the protein segment linking motifs II and III, which activates Ry<sub>1</sub> receptor opening. The large cytoplasmic domain ("foot process") has multiple binding sites for ATP, Ca2+, Mg2+, and a variety of drugs, including volatile anesthetics. The FK binding protein (FKBP12) binds to the Ry<sub>1</sub> receptor and modulates its gating. Triadin is a protein closely associated with JSR that appears to regulate Ca2+ release from the SR lumen, possibly by transmitting the release signal to calsequestrin, an acidic protein with many low affinity Ca2+ binding sites.

the reports that chlorocresol and serotonin agonists can activate Ca2+ release are important reminders that pharmacologic or even physiologic regulatory mechanisms can perturb muscle homeostasis. In normal muscle, such additional "leak" caused by drugs or by normally adaptive physiologic regulatory pathways may cause no appreciable change in muscle metabolism. However, in myopathic muscle, either physiologic or pharmacologic mediators of increased release may cause the muscle to become hypermetabolic. In this setting, the MH episode represents the "final common pathway" resulting from the convergence of a number of small abnormalities. Such interventions may be insufficient triggers by themselves, but in combination with other agents, such as volatile anesthetics, their effect may result in excessive Ca<sup>2+</sup> leak, which triggers an MH episode. It is possible that, in rare instances, sufficient pharmacologic effects and physiologic modulations may lead to triggering of muscle that bears no inherited defect, i.e., acquired MH.

The critical physiologic role of the Ry<sub>1</sub> receptor in controlling Ca<sup>2+</sup> release is emphasized by a mutation in this protein (Arg<sup>615</sup>→Cys) that is associated with and almost certainly accounts for porcine MH.<sup>26</sup> Anesthetics or the neuroendocrine changes induced by stress cause loss of Ca<sup>2+</sup> homeostasis, hypermetabolism, and the catastrophic consequences. A homologous mutation (Arg<sup>614</sup>→Cys) to that in the pig accounts for perhaps 5% of human MH.<sup>27,28</sup> Central core disease is a

rare myopathy characterized by hypotonia, weakness in the lower extremities, and MH susceptibility. This defect, which is associated with Ry<sub>1</sub> receptor mutations, <sup>29,30</sup> probably accounts for a small fraction of MH. Other mutations in Ry<sub>1</sub> receptor (Gly<sup>248</sup>—Arg; Gly<sup>341</sup>—Arg; Gly<sup>2433</sup>—Arg), <sup>31–33</sup> or at least located on chromosome 19q13.1, have been defined and may underlie the etiology of additional forms of MH. However, the gene locus responsible for MH in more than 50% of European MH families appears not to be located within the Ry<sub>1</sub> receptor gene.<sup>34</sup>

Although the Ry<sub>1</sub> receptor is the major component of the Ca2+ release pathway, important interactions between Ry<sub>1</sub> receptors and at least five other triadic proteins (DHP receptor  $\alpha_{18}$  subunit, FKBP12, triadin, calsequestrin, and calmodulin) appear to contribute to the function and modulation of the Ca2+ release mechanism. An exogenous peptide fragment (KC7) that copurifies with Ry<sub>1</sub> receptor<sup>35</sup> was shown to be identical to the major cytoplasmic immunophilin of human Tcells, FKBP12 (FK-506 binding protein-12kDa).36 FKBP12 is a 12 kDa protein that belongs to a growing class of cytosolic proteins that regulate signal transduction pathways essential to immune function. Recent evidence suggests that the FKBP12/Ry<sub>1</sub> receptor heterocomplex appears to stabilize the closed conformation of the Ca<sup>2+</sup> release channels.<sup>37</sup> Mack et al. identified a class of macrocyclic alkaloids known as bastadins, which mediate their actions by markedly altering

the interaction between FKBP12 and Rv, receptors, resulting in a marked (~50-fold) increase in the time the channel is open but with no change in unitary conductance. 38 Through their actions on FKBP12, bastadins modulate the ratio of ryanodine-insensitive "leak" states to ryanodine-sensitive channel states in the SR membranet and may define a new site of drug action. Triadin is a 95-kDa protein that closely associates with Ry<sub>1</sub> receptors and appears to play a functional role in excitation-contraction coupling. 39,40 A small number of highly reactive sulfhydryl moieties on both Ry1 receptors and triadin proteins have been identified and appear necessary for normal Ca2+ release channel function.41 Calsequestrin is a highly acidic protein that resides in the lumen of the SR and has a high capacity for binding Ca2+. Ikemoto et al. 42 showed that activation of Ry<sub>1</sub> receptors by ligands elicits a signal in the junctional face membrane which is transmitted to calsequestrin. This signal appears to be important to release of bound Ca<sup>2+</sup> within the SR lumen. In support, intraluminal SR Ca2+ has been shown to be an important factor in Ry<sub>1</sub> receptor function; furthermore, triadin may functionally relate calsequestrin to Ry<sub>1</sub> receptor. 42

These Ry<sub>1</sub> receptor-associated proteins may be of considerable importance in a number of ways in addition to their physiologic role. First, these proteins represent sites at which mutations might occur that may alter skeletal muscle function, leading to altered Ca<sup>2+</sup> homeostasis, myopathies, and possibly some of the as yet genetically undefined forms of MH. It is interesting that a mutation in the largely extracellular  $\alpha_2$ - $\delta$  subunit of the surface Ca channel, which can modulate the  $\alpha_1$  DHP receptor, is associated with MH.<sup>43</sup>

Second, these proteins may be additional sites of drug action. Although limited by the solubility of dantrolene, Parness and Palnitkar<sup>44</sup> identified a specific binding site for [ $^3$ H]-dantrolene in preparations of porcine skeletal muscle SR having a K<sub>d</sub> of 277 nm. In addition, the binding characteristics were highly distinct from ryanodine, an agent that also stimulates release at low concentrations ( $<10~\mu$ M) and blocks at higher concentrations ( $>100~\mu$ M). More recent data from that laboratory suggests that the specific dantrolene binding site resides on a smaller (10-120~kDa) protein,  $^{45}$  compatible with either triadin or FKBP12. Multiple sites may exist for dantrolene binding to explain its biphasic effects. The observation that Ry<sub>1</sub> receptor may not be a site of action

for dantrolene is consistent with the fact that dantrolene can be used to treat human MH, which presumably arises from a variety of defects.

Finally, the presence of these proteins in "native" channels isolated from SR may differ from the behavior of "purified" Ry1 receptor protein reconstituted into bilayer lipid membranes.8 The isolation process may alter responses such that the preparation does not accurately reflect in vivo responses. When pure Ry1 receptor channels are expressed from cDNA, their gating is considerably different from channels obtained with fused SR vesicles. The incidence of subconductance states is reduced when purified FKBP12 protein is added to pure Ry<sub>1</sub> receptor (expressed from cDNA), but normal (rapid) gating behavior is not completely restored. 46 Consequently, when examining studies such as that by Nelson et al.1 in this issue, it is important to recognize that the presence and role of the additional regulatory segments are uncertain.

What should we conclude regarding dantrolene? The effect of dantrolene in enhancing Ca2+ release might be only a biochemical curiosity, were it not for the MHAUS data suggesting that it may result clinically in mild hypermetabolism after treatment. Unfortunately, we cannot be certain whether the situations between the dantrolene-treated and untreated groups are truly comparable, nor can we specifically determine the basis for the MH episode. Nevertheless, as drug metabolism results in declining blood dantrolene concentrations, recrudescence of hypermetabolism appears to be a risk, although its severity is unclear. It is interesting that abnormal Ry<sub>1</sub> receptor channels were less profoundly activated by dantrolene than were the normal Ry, receptor channels. Perhaps individuals with normal Ry<sub>1</sub> receptor but with myopathies and MH susceptibility due to other gene defects are the most susceptible to activation by low concentrations of dantrolene, particularly if combined with other agents (e.g., chlorocresol) or particular clinical settings. Ongoing investigation into the various genetic causes of MH and the pharmacology of dantrolene should provide answers. In any case, dantrolene remains an as yet irreplaceable drug for MH crises and is the lifeline to survival once MH is triggered. No other drug permits dependable reversal of skeletal hypermetabolism. However, careful informed observation is essential during recovery from dantrolene-treated MH episodes.

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## References

- 1. Nelson TE, Lin M, Zapata-Sudo G, Sudo RT: Dantrolene sodium can increase or attenuate activity of skeletal muscle ryanodine receptor calcium release channel: Clinical implications. Anesthesiology 1996; 84:1368–79
- 2. Tegazzin V, Scutari E, Treves S, Zorzato F: Chlorocresol, an additive to commercial succinylcholine, induces contracture of malignant hyperthermia-susceptible muscles *via* activation of the ryanodine receptor Ca<sup>2+</sup> channel. Anesthesiology 1996; 84:1380–5
- 3. Wappler F, Roewer N, Köchling A, Scholz J, Löscher W, Steinfath M, Schulte am Esch J: Effects of serotonin<sub>2</sub> receptor agonist DOI on skeletal muscle specimens from malignant hyperthermia-susceptible patients. Anesthesiology 1996; 84:1280–7
- 4. McPherson PS, Campbell KP: The ryanodine receptor/Ca<sup>2+</sup> release channel. J Biol Chem 1993; 268:13765–8
- 5. Tanabe T, Beam KG, Powell JA, Numa S: Restoration of excitation-contraction coupling and slow calcium current in dysgenic mice by dihydropyridine receptor complementary DNA. Nature 1988; 336:134–9
- 6. Tanabe T, Beam KG, Adams BA, Niidome T, Numa S: Regions of the skeletal muscle dihydropyridine receptor critical for excitation-contraction coupling. Nature 1990; 346:567–9
- 7. Pessah IN, Waterhouse AL, Casida JE: The calcium-ryanodine receptor complex of skeletal and cardiac muscle. Biochem Biophys Res Commun 1985; 128:449–56
- 8. Pessah IN, Francini AO, Scales DJ, Waterhouse AL, Casida JE: Calcium-ryanodine receptor complex: Solubilization and partial characterization from skeletal muscle junctional sarcoplasmic reticulum vesicles. J Biol Chem 1986; 261:8643–8
- 9. Fleischer S, Ogunbunmi EM, Dixon MC, Fleer EAM: Localization of Ca2+ release channels with ryanodine in junctional terminal cisternae of sarcoplasmic reticulum of fast skeletal muscle. Proc Natl Acad Sci USA 1985; 82:7256–9
- 10. Block BA, Imagawa T, Campbell KP, Franzini-Armstrong C: Structural evidence for direct interaction between the molecular components of the transverse tubule/sarcoplasmic reticulum junction in skeletal muscle. J Cell Biol 1988; 107:2587–600
- 11. Pessah IN, Zimanyi I: Characterization of multiple [<sup>3</sup>H]-ryanodine binding sites on the Ca<sup>2+</sup> release channel of the sarco-plasmic reticulum from skeletal and cardiac muscle: Evidence for a sequential mechanism in ryanodine action. Mol Pharmacol 1991; 39:679–89
- 12. Buck E, Zimanyi I, Abramson JJ, Pessah IN: Ryanodine stabilized multiple conformational states of the skeletal muscle calcium release channel. J Biol Chem 1992; 267:23560–7

- 13. Zimányi I, Buck E, Abramson JJ, Mack MM, Pessah IN: Ryanodine induces persistent inactivation of the Ca<sup>2+</sup> release channel from skeletal muscle sarcoplasmic reticulum. Mol Pharmacol 1992; 42: 1049–57
- 14. Smith JS, Imagawa T, Ma J, Fill M, Campbell KP, Coronado R: Purified ryanodine receptor from rabbit skeletal muscle is the calcium-release channel of sarcoplasmic reticulum. J Gen Physiol 1988; 92: 1–26.
- 15. Coronado R, Morrissette M, Sukhareva M, Vaughn DM: Structure and function of ryanodine receptors. Am J Physiol 1994; 266: C1485–504
- 16. Takeshima H, Nishimura S, Matsumoto T, Ishida H, Kangawa K, Minamino N, Matsuo H, Ueda M, Hanaoka M, Hirose T, Numa S: Primary structure and expression from complementary DNA of skeletal muscle ryanodine receptor. Nature 1989; 339:439–45
- 17. Zorzato F, Fujii J, Otsu K, Phillips M, Green NM, Lai FA, Meissner G, MacLennan DH: Molecular cloning of cDNA encoding human and rabbit forms of the Ca<sup>2+</sup> release channel (ryanodine receptor) of skeletal muscle sarcoplasmic reticulum. J Biol Chem 1990; 265: 2244–56
- 18. Meissner G, Henderson JS: Rapid calcium release from cardiac sarcoplasmic reticulum vesicles is dependent on Ca<sup>2+</sup> and is modulated by Mg<sup>2+</sup>, adenine nucleotide, and calmodulin. J Biol Chem 1987; 262:3065–73
- 19. Pessah IN, Stambuk RA, Casida JE:  $Ca^{2+}$ -activated ryanodine binding: mechanisms of sensitivity and intensity modulation by  $Mg^{2+}$ , caffeine and adenine nucleotides. Mol Pharmacol 1987; 31:232-8
- 20. Sabbadini R, McNutt W, Jenkins G, Betto R, Salviati G: Sphingosine is endogenous to cardiac and skeletal muscle. Biochem Biophys Res Commun 1993; 193:752–8
- 21. Wang J, Best P: Inactivation of the sarcoplasmic reticulum calcium channel by protein kinase. Nature 1992; 359:739–
- 22. Connelly TJ, Hayek R-E, Rusy BF, Coronado R: Volatile anesthetics selectively alter [³H]ryanodine binding to skeletal and cardiac ryanodine receptors. Biochem Biophys Res Commun 1992; 186:595–600
- 23. Nelson TE: Halothane effects on human malignant hyperthermia skeletal muscle single calcium-release channels in planar lipid bilayers. Anesthesiology 1992; 76:588–95
- 24. Nelson TE, Lin M: Abnormal function of porcine malignant hyperthermia calcium release channel in the absence and presence of halothane. Cell Physiol Biochem 1995; 5:10–22
- 25. McGrath CJ, Lee JC, Rempel WE: Halothane testing for malignant hyperthermia in swine: Dose-response effects. Am J Vet Res 1984; 45:1734–6
- 26. Fujii J, Otsu K, Zorzato F, De Leon S, Khanna VK, Weiler JE, O'Brien PJ, MacLennan DH: Identification of a mutation in porcine ryanodine receptor associated with malignant hyperthermia. Science 1991; 253:448–51
- 27. Gillard EF, Otsu K, Fujii J, Khanna VK, De Leon S, Derdemezi J, Britt BA, Duff CL, Worton RG, MacLennan DH: A substitution of cysteine for arginine 614 in the ryanodine receptor is potentially causative of human malignant hyperthermia. Genomics 1991; 11: 751–5
- 28. Hogan K, Couch F, Powers PA, Gregg RG: A cysteine-for-arginine substitution (R614C) in the human skeletal muscle calcium release channel cosegregates with malignant hyperthermia. Anesth Analg 1992; 75:441–8

- 29. Zhang Y, Chen HS, Khanna VK, De Leon S, Phillips MS, Schappert K, Britt BA, Brownell AKW, MacLennan DH: A mutation in the human ryanodine receptor gene associated with central core disease. Nature Genetics 1993; 5:46–50
- 30. Quane KA, Healy JMS, Keating KE, Manning BM, Couch FJ, Palmucci LM, Douguzzi C, Fagerlund TH, Berg K, Ording H, Bendixen D, Mortier W, Linz U, Muller CR, McCarthy TV: Mutations in the ryanodine receptor gene in central core disease and malignant hyperthermia. Nature Genetics 1993; 5:51–5
- 31. Gillard EF, Otsu K, Fujii J, Duff C, De Leon S, Khanna VK, Britt BA, Worton RG, MacLennan DH: Polymorphisms and deduced amino acid substitutions in the coding sequence of the ryanodine receptor (RYR1) gene in individuals with malignant hyperthermia. Genomics 1992; 13:1247–54
- 32. Quane KA, Keating KE, Manning BM, Healy JM, Monsieurs K, Heffron JJ. Lehane M, Heytens L, Krisovic-Horber R, Adnet P, Ellis FR, Monnier N, Lunardi J, McCarthy TV: Detection of a novel common mutation in the ryanodine receptor gene in malignant hyperthermia: Implications for diagnosis and heterogeneity studies. Hum Mol Genet 1994; 3:471–6
- 33. Phillips MS, Khanna VK, De Leon S, Frodis W, Britt BA, MacLennan DH: The substitution of Arg for Gly2433 in the human skeletal muscle ryanodine receptor is associated with malignant hyperthermia. Hum Mol Genet 1994; 3:2181–6
- 34. Ball SP, Johnson KJ: The genetics of malignant hyperthermia. J Med Genet 1993; 30:89–93
- 35. Marks A, Fleischer S, Tempst P: Surface topography analysis of the ryanodine receptor/junctional channel complex based on proteolysis sensitivity mapping. J Biol Chem 1990; 265:13143–9
- 36. Jayaraman T, Brillantes A, Timerman A, Fleischer S, Erdjument-Bromage H, Tempst P, Marks A: FK506 binding protein associated with the calcium release channel (ryanodine receptor). J Biol Chem 1992; 267:9474–7
- 37. Timerman AP, Ogunbumni E, Freund E, Wiederrecht G, Marks AR, Fleischer S: The calcium release channel of sarcoplasmic reticulum is modulated by FK506-binding protein. J Biol Chem 1993; 268:22992–9

- 38. Mack MM, Molinski TF, Buck ED, Pessah IN: Novel modulators of skeletal muscle FKBP12/calcium channel complex from *lanthella basta*. J Biol Chem 1994; 269:23236–49
- 39. Caswell AH, Brandt NR, Brunschwig J-P, Purkerson S: Localization and partial characterization of the oligomeric disulfide-linked molecular weight 95 000 protein (triadin) which binds the ryanodine and dihydropyridine receptors in skeletal muscle triadic vesicles. Biochemistry 1991; 30:7507–13
- 40. Knudson CM, Stang KK, Jorgensen AO, Campbell KP: Biochemical characterization and ultrastructural localization of a major junctional sarcoplasmic reticulum glycoprotein (triadin). J Biol Chem 1993; 268:12637–45
- 41. Liu G, Pessah IN: Molecular interaction between ryanodine receptor and glycoprotein triadin involves redox cycling of functionally important hyperreactive sulfhydryls. J Biol Chem 1994; 269: 33028–34
- 42. Ikemoto N, Antoniu B, Kang J-J, Mészáros LG, Ronjat M: Intravesicular calcium transient during calcium release from sarcoplasmic reticulum. Biochemistry 1991; 30:5230–7
- 43. Iles DE, Lehmann-Horn F, Scherer SW, Tsui L-C, Weghuis DO, Suijkerbuijk RF, Heytens L, Mikala G, Schwartz A, Ellis FR, Stewart AD, Deufel T, Wieringa B: Localization of the gene encoding the  $\alpha_2/\delta$ -subunits of the L-type voltage-dependent calcium channel to chromosome 7q and analysis of the segregation of flanking markers in malignant hyperthermia susceptible families. Hum Mol Genet 1994; 3:969-75
- 44. Parness J, Palnitkar SS: Identification of dantrolene binding sites in porcine skeletal muscle sarcoplasmic reticulum. J Biol Chem 1995; 270:18465–72
- 45. Palnitkar SS, Parness J: Partial purification of the dantrolene receptor from skeletal muscle: non-identity with the ryanodine receptor (abstract). Anesthesiology 1995; 83:A729
- 46. Brillantes A-MB, Ondrias K, Scott A, Kobrinsky E, Ondriasová E, Moschella MC, Jayarama T, Landers M, Ehrlich BE, Marks AR: Stabilization of calcium release channel (ryanodine receptor) function by FK506-binding protein. Cell 1994; 77:513–23