CYTOMEGALOVIRUS AL

10 May 31, 1994 and was mittee for the Protection sies were performed on I respiratory failure and/ ventilator-associated pne antemortem bacteriolog imen brush, bronckoalve sampling). Open-lang t a patient's respiratory st teriologic cultures were men brush, bronchoalve sampling) and no expla piratory status could be CMV pneumonia vas di teria were met: (1) mec 7 days; (2) histol@gica This delay virtuall ₹ gua histologically proven CM as such. Immunocompr from the study. Patient compromised if there of immunodeficiency ciency syndrome) sor if munosuppressive agent problem (i.e., lympho tion, etc.). Patients rece sidered to be immunoc data were prospegivel cian: age; sex; Acute Ph Evaluation II scoreson ac blood cell differential lymphocytosis; segum bilirubin, aspartate an aminotransferase; maxin toentgenogram using V

> Lung Biopsy Samplin Histologic assessment specimens obtained by mortem histologic exa In such cases, four or each lobe were taken They were fixed in 109 at room temperature. T in a modified alcohol s

> underlying diagnoses; u

(Cytovene, Syntex Pha

twice a day for at least

ical ventilation; duration

outcome.

1996; 84:280-7 © 1996 American Society of Anesthesiologists, Inc Lippincott–Raven Publishers

Cytomegalovirus

An Unexpected Cause of Ventilator-associated Pneumonia

Laurent Papazian, M.D.,* Alain Fraisse, M.D.,† Louise Garbe, M.D.,‡ Christine Zandotti, M.D.,§ Pascal Thomas, M.D., || Pierre Saux, M.D., * Gilles Perrin, M.D., * François Gouin, M.D.#

Background: Cytomegalovirus (CMV) frequently is observed in immunocompromised hosts. The aim of this study was to report cases of ventilator-associated CMV pneumonia diagnosed by pathologic examination in intensive care patients without acquired immunodeficiency syndrome or hematologic malignancy or who were not receiving immunosuppressive agents.

Methods: From June 1, 1989, to May 31, 1994, 2,785 patients were hospitalized. During the study period, 60 autopsies and 26 open-lung biopsies were performed in nonimmunocompromised patients who were seen with acute respiratory failure and/or symptoms suggestive of ventilator-associated pneumonia. Cytomegalovirus pneumonia was diagnosed using pulmonary samples by the identification of large cells with large nuclei containing a basophilic or eosinophilic inclusion surrounded by a light halo. These typical findings always were associated with a diffuse interstitial pneumonitis.

Results: Cytomegalovirus pneumonia was diagnosed after histologic examination in 25 patients. The reason for admission to the intensive care unit was major surgery in 13 patients and medical problems in 12 patients. Ventilator-associated CMV pneumonia was diagnosed by histologic examination 22.4 \pm 8.8 days after admission to the intensive care unit (median 18 days; range 10-40 days). The clinical description was similar with the 25 patients who were seen with ventilator-associated CMV pneumonia and the 61 patients without ventilator-asso-

ciated CMV pneumonia. However, there was a more severe hypoxemia (72 \pm 16 vs. 95 \pm 41 mmHg, P < 0.05) and a higher Weinberg's radiologic score (9.2 \pm 1.9 vs. 7.4 \pm 2.7, P < 0.05) in the ventilator-associated CMV pneumonia group. Diagnosis of ventilator-associated CMV pneumonia was made for 9 of 17 patients when shell-vial culture technique using fluoresceinlabeled antibody E 13 was performed on bronchoalveolar lavage products. Four of the eight patients treated by ganciclovir therapy died of multiple organ dysfunction syndrome.

Conclusions: The diagnosis of ventilator-associated CMV pneumonia should not be excluded in intensive care patients, even those without acquired immunodeficiency syndrome, hematologic malignancy, or immunosuppressive agents on admission. (Key words: Complications: cytomegalovirus; pneumonia. Lavage: bronchoalveolar. Ventilation: mechani-

MOST adults have been infected by cytomegalovirus (CMV), as shown by the presence of detectable antibodies. Furthermore, this CMV infection remains latent. When cell-mediated immunity is depressed, for example, in renal transplant patients receiving immunosuppressant drugs, reactivation of endogenous CMV is common and may produce clinical infection. Cellmediated immunity also is reduced in other surgical patients, particularly those who are critically ill with serious postoperative complications or those who have multiple injuries.2 Until recently, CMV pneumonia was thought to be a rare but fatal illness, occurring primarily in immunocompromised hosts. Numerous viruses currently are considered nosocomial pathogens,3 but the importance of these agents compared with other pathogens has not been clearly defined, especially in adult patients in whom the lungs are mechanically ventilated. The aim of this study was to report a series of ventilator-associated CMV pneumonia diagnosed by histopathologic examination.

Received from the Department of Anesthesia and Intensive Care, Laboratory of Virology, Department of Pathology, and Department of Thoracic Surgery, Hôpital Sainte-Marguerite, Marseille, France. Submitted for publication December 9, 1994. Accepted for publication September 13, 1995. Presented in part at the 33rd Interscience Conference on Antimicrobial Agents and Chemotherapy, New Orleans, Louisiana, October 17-20, 1993.

Address reprint requests to Dr. Papazian: Staff Anesthesiologist, Département d'Anesthésie-Réanimation, Hôpital Sainte-Marguerite, 13274 Marseille Cedex 9, France.

Materials and Methods

This study was conducted in our 15-bed medicosurgical intensive care unit (ICU) from June 1, 1989

Anesthesiology, V 84, No 2, Feb 1996

Anesthesiology, V 84, No 2, Fo

^{*} Staff Anesthesiologist, Département d'Anesthésie-Réanimation.

[†] Resident, Département d'Anesthésie-Réanimation.

[‡] Staff Pathologist, Service d'Anatomo-Pathologie.

[&]amp; Staff Virologist, Laboratoire de Virologie

Staff Surgeon, Service de Chirurgie Thoracique.

[#] Professor, Département d'Anesthésie-Réanimation

to May 31, 1994 and was approved by the local Committee for the Protection of Human Subjects. Autopsies were performed on patients who died with acute respiratory failure and/or symptoms suggestive of ventilator-associated pneumonia (VAP) with negative antemortem bacteriologic cultures (protected specimen brush, bronchoalveolar lavage, blind bronchial sampling). Open-lung biopsy was performed when a patient's respiratory status was worsening but bacteriologic cultures were negative (protected specimen brush, bronchoalveolar lavage, blind bronchial sampling) and no explanation for the impaired respiratory status could be found. Ventilator-associated CMV pneumonia was diagnosed when these two criteria were met: (1) mechanical ventilation for at least 7 days; (2) histologically proven CMV pneumonia. This delay virtually guaranteed that all patients with histologically proven CMV pneumonia were classified as such. Immunocompromised hosts were excluded from the study. Patients were classified as immunocompromised if there was an established diagnosis of immunodeficiency (i.e., acquired immunodeficiency syndrome), or if the patient was receiving immunosuppressive agents for an underlying medical problem (i.e., lymphoma, prior organ transplantation, etc.). Patients receiving steroids were also considered to be immunocompromised.4 The following data were prospectively recorded by a single physician: age; sex; Acute Physiology and Chronic Health Evaluation II score on admission⁵; hemoglobin; white blood cell differential count; presence of atypical lymphocytosis; serum concentrations of creatinine, bilirubin, aspartate aminotransferase, and alanine aminotransferase; maximal abnormalities on the chest roentgenogram using Weinberg's radiologic score6; underlying diagnoses; use of 5.0 mg · kg⁻¹ ganciclovir (Cytovene, Syntex Pharmaceuticals, Palo Alto, CA) twice a day for at least 2 weeks; duration of mechanical ventilation; duration of hospitalization; and final outcome.

Lung Biopsy Sampling and Processing

Histologic assessment was performed on pulmonary specimens obtained by open-lung biopsy or post-mortem histologic examination of the whole lung. In such cases, four or more biopsy specimens from each lobe were taken for histologic examination. They were fixed in 10% buffered formalin for 24 h at room temperature. Then samples were dehydrated in a modified alcohol series: 95% for 15 min, 100%

for 15 min, and xylene for 15 min. After dehydration, samples were embedded in a single paraffin block and serially cut to 4-µm thick with standard microtomes with disposable blades. Slides were stained with hematoxylin-eosin-safran. Cytomegalovirus pneumonia was diagnosed on pulmonary samples by the identification of large cells with large nuclei containing a basophilic or eosinophilic inclusion surrounded by a light halo.^{7,8} Multiple cytoplasmic granular inclusions were often present. These typical findings were always associated with a diffuse interstitial pneumonitis characterized by the presence of inflammatory cells (predominantly lymphocytes), thickened alveolar septi, and an interstitial inflammation. A fibrinous alveolitis and a mild or moderate fibrosis were often associated. Bacterial pneumonia was defined by the presence of scattered neutrophilic infiltrates localized to terminal bronchioles and surrounding alveoli with evident confluence of infiltrates between adjacent lobules. 9,10 Bacteriologic investigation was performed on open-lung biopsy or autopsy and included Gram and Ziehl-Neelsen staining and culture for bacteria, mycobacteria, and fungi.

Serologic Status: Viral Cultures

The CMV antibody status of patients was determined on admission to the ICU by an enzyme-linked immunosorbent assay (Behring, Marburg, Germany). Bronchoalveolar lavage (BAL), blood, or urine cultures were performed when the diagnosis of CMV pneumonia was suspected clinically. Bronchoalveolar lavage was performed by wedging the bronchoscope into a subsegment of the area with greatest radiologic abnormality, or when disease was diffuse, into the lingula or right middle lobe. Normal saline was sequentially instilled in 20-ml aliquots (total, to 100 ml) and suctioned into sterile traps. The first aliquot was discarded. Bronchoalveolar lavage specimens were sent for conventional microbiologic processing. In addition, BAL, blood, and urine samples were sent for shell-vial culture technique. Specimens for these cultures were inoculated onto MRC-5 cells in tissue culture.11 Monoclonal antibodies directed against immediate early antigen (E 13, Biosoft, Clonatec, Paris) were applied 48 h after inoculation for detection of viral antigen expression.¹² Only BAL, blood, and urine cultures performed within a 1-week period preceding histologic assessment were consid-

edico-, 1989

severe

nigher

(0.05)

gnosis of 17

scein-

lar la-

clovir

1 CMV

tients.

rome.

nts on

virus;

chani-

virus

anti-

atent.

or ex-

mmu-

CMV

Cell-

rgical

l with

have

ia was

marily

es cur-

ut the

other

ally in

nically

series

sed by

Histologic Diagnosis of Ventilator-associated Cytomegalovirus Pneumonia

There were 2,785 admissions to the ICU during the study period. During this period, 60 autopsies and 26 open-lung biopsies were performed in nonimmunocompromised patients. In all, we observed 25 cases of CMV pneumonia diagnosed by histologic examination (fig. 1). In the remaining 61 cases, no histologic sign of CMV pneumonia was observed. Ventilator-associated CMV pneumonia was diagnosed by histologic examination 22.4 ± 8.8 days after ICU admission (median, 18 days; range, 10-40 days). Seventeen of these cases of CMV pneumonia were diagnosed on autopsy and eight cases by open-lung biopsy. Histologic examination showed that CMV was the sole respiratory pathogen in 88% of the cases. In three patients, lung cultures were positive with one microorganism (Staphylococcus aureus, Serratia marcescens, and Streptococcus pneumoniae) and histologic results were consistent with a bacterial pneumonia associated with the CMV pneumonia.

Diagnostic Performance of the Various Testing Methods Used to Diagnose Ventilator-associated Cytomegalovirus Pneumonia

Patient serologic status was determined in 18 cases. Thirteen of these 18 patients were seropositive for CMV on admission to the ICU. During the same study period, 58% of the 2,785 patients admitted to the ICU were seropositive for CMV. Bronchoalveolar lavage was performed in 17 of the 25 study patients whereas blood and urine cultures were performed in 15 of the 25 study patients. Bronchoalveolar lavage, blood, and urine cultures were performed during the week preceding histologic assessment in 43 of the 61 patients without histologic signs of CMV pneumonia. The sensitivity of BAL, blood, and urine cultures for the diagnosis of histologically proven ventilator-associated CMV pneumonia was 53%, 20%, and 13%, respectively. The specificity of these tests for the diagnosis of histologically proven ventilator-associated CMV pneumonia was 92%, 83%, and 62%, respectively.

Comparison of Clinical Presentation of Ventilator-associated Cytomegalovirus Pneumonia with Patients without Cytomegalovirus Pneumonia

Characteristics of the 25 patients with ventilatorassociated CMV pneumonia are outlined in table 1. The onset of the clinical deterioration of respiratory status occurred at 17.4 ± 8.8 days after ICU admission (median, 14 days; range, 8-38 days). The presence of fever >38°C was observed in all but 6 patients (mean, 38.5°C; range, 37.0-40.1°C). Chest roentgenograms did not show any specific signs; Weinberg's score was 9.2 ± 1.9 (median, 9; range, 4-12). Evidence of CMV infection of the retina was never found. Involvement of the gastrointestinal tract was noted in one of the 25 patients with CMV pneumonia. At the moment of diagnosis of CMV pneumonia, the minute ventilation used was $13.2 \pm 3.0 \, l \cdot min^{-1}$ (range, 9.0-20.0), and the highest FIO2 used was 0.68 ± 0.17 (range, 0.45-1.0). All patients were ventilated with positive end-expiratory pressure. Laboratory data are summarized in table 2. Hypoxemia and moderate hypercapnia were observed on arterial blood gases at the moment of diagnosis of VAP. The CD4+:CD8+ ratio was assessed in 17 patients and was found reversed in 5 of them.

Comparison of the 25 patients with VAP and the 61 patients without histologic sign of CMV pneumonia showed a more severe hypoxemia in the CMV group $(72 \pm 16 \text{ mmHg } vs. 95 \pm 41 \text{ mmHg, } P < 0.05)$ and a higher Weinberg's score $(9.2 \pm 1.9 \text{ } vs. 7.4 \pm 2.7, P < 0.05)$ whereas age, Acute Physiology and Chronic Health Evaluation II score on admission, duration of mechanical ventilation, PaCO₂, white blood cell count, and temperature were not statistically different.

Evolution of the Respiratory Status of the Treated Patients

Only the eight patients who underwent open-lung biopsy were treated with ganciclovir for their ventilator-associated CMV pneumonia (table 3). The pulmonary status of four of these eight patients was improved within the first 5 days after the onset of ganciclovir therapy permitting an increase of more than 75% of their PaO₂/FIO2 ratio. Of the four patients who died, we observed an initial good response to antiviral therapy in two cases, with an increase of the PaO₂/FIO2 ratio of 25–30%.

Discussion

Cytomegalovirus Pneumonia and Mechanical Ventilation

The purpose of this study was to describe and elaborate on the previously unreported problem of venti-

Fig. 1. Patient 18. (A Surgi men from a right upper lo Lobular adenocarcinoma a surrounded by healthy par (magnification ×250 hen eosin-safran stain). (B) Ope opsy on day 37 afteg surg megalovirus pneumonitis cells with large nuclei con inclusion surrounded by a (arrows) associated with ence of inflammators cells cation ×400, hematexylin ran stain). (C) Postmorte nation on day 51 after Cytomegalovirus prieumo large cells with largenucle ing an inclusion surroun light halo (arrow) associate presence of inflammato thickened alveolar septi w fibrosis, and hyaline m (magnification ×400 her eosin-safran stain).

lator-associated CMV printsologic signs, the "go of CMV pneumonia. Act tologic examination has tool for the diagnosis of CMV induces highly spidismic inclusions) that produced by other Herythe presence of pathogn

00005.pdf by guest on 05 April 2024

Fig. 1. Patient 18. (A) Surgical specimen from a right upper lobectomy. Lobular adenocarcinoma associated surrounded by healthy parenchyma (magnification ×250, hematoxylineosin-safran stain). (B) Open-lung biopsy on day 37 after surgery. Cytomegalovirus pneumonitis with large cells with large nuclei containing an inclusion surrounded by a light halo (arrows) associated with the presence of inflammatory cells (magnification ×400, hematoxylin-eosin-safran stain). (C) Postmortem examination on day 51 after surgery. Cytomegalovirus pneumonitis with large cells with large nuclei containing an inclusion surrounded by a light halo (arrow) associated with the presence of inflammatory cells, thickened alveolar septi with septal fibrosis, and hyaline membranes (magnification ×400, hematoxylineosin-safran stain).

ssion sence tients oent-Wein-12). never t was onia.

i, the

nin⁻¹

l was

were

sure.

pox-

d on

sis of

7 pa-

ne 61

ionia

roup

and and

2.7,

ronic

on of

cell liffer-

eated

ventie pulas imgancin 75% o died,

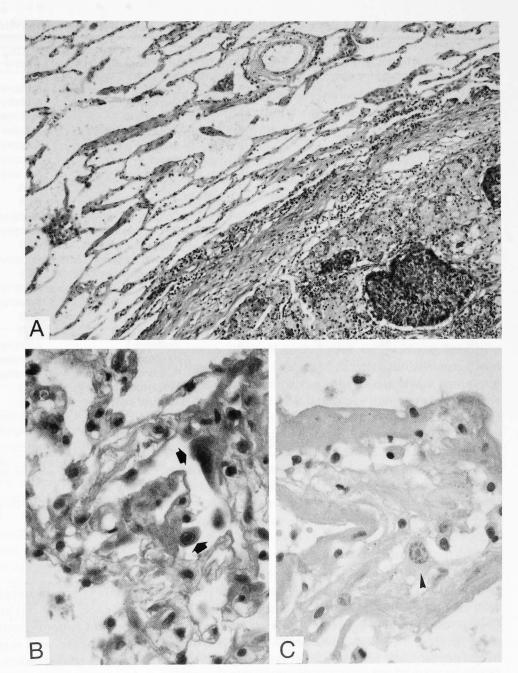
l ther-

/FIO2

al

d elab-

venti-



lator-associated CMV pneumonia in the ICU. We used histologic signs, the "gold standard," for the diagnosis of CMV pneumonia. According to the literature, histologic examination has proved to be a highly specific tool for the diagnosis of CMV pneumonia. 11,13–15 In fact, CMV induces highly specific cytologic modifications (*i.e.* cytomegaly, intranuclear inclusions, intracytoplasmic inclusions) that are totally different from those produced by other Herpesviridae. 16,17 We considered the presence of pathognomonic cells with intranuclear

inclusions to be a necessary criterion for the diagnosis of CMV pneumonia. From a practical point of view, it would appear necessary to examine several sections because cytologically altered cells are present only in some sections. Distribution of infected cells within a tissue may be highly variable, thus reducing the chance of finding them in a given section.

Oda *et al.*¹⁸ have noted that CMV pneumonia could be observed in patients without hematologic malignancies or acquired immunodeficiency syndrome. They

Patient No.	Age (yr)	Sex	APACHE II on Admission	Diagnosis	OLB/ Autopsy	Mechnical Ventilation (days)*	Blood Transfusion
	66	F	25	COLD‡	Autopsy	14	Yes
1	76	м	17	Peritonitis§	Autopsy	42	Yes
2	72	M	16	Peritonitis§	Autopsy	22	Yes
	82	M	18	Cardiac surgery§	Autopsy	34	Yes
4	69	M	12	COLD§	Autopsy	15	Yes
5 6	77	M	22	Coma	Autopsy	22	Yes
7	67	M	16	COLD‡	Autopsy	29	Yes
	72	F	27	COLD‡	OLB	8	Yes
8	78	М	20	COLD‡	OLB	7	Yes
9	76	F	17	Coma	Autopsy	16	Yes
10	76	F	29	Gastric cancer§	Autopsy	14	Yes
11	67	M	13	Cardiac surgery§	OLB	17	Yes
12	54	M	14	Cardiac surgery§	Autopsy	7	Yes
13		M	12	Esophageal cancer§	OLB	26	No
14	53 75	F	29	Coma	Autopsy	17	No
15	75 71	F	31	Meningitis	Autopsy	15	No
16		F	21	Myocarditis	OLB	31	No
17	49	M	15	Lung cancer§	OLB	37	Yes
18	72	F	18	Peritonitis§	Autopsy	13	Yes
19	84	F	23	Hepatocarcinoma§	Autopsy	10	Yes
20	68		35	Pulmonary embolism	Autopsy	36	Yes
21	65	М	15	Esophageal cancer§	OLB	20	No
22	63	F			OLB	31	Yes
23	65	М	20	Vascular surgery§ Coma		10	Yes
24	54	F	30		Autopsy	7	No
25	62	М	17	Lung cancer§	Autopsy	20.0 ± 10.5	140
Mean \pm SD	68.3 ± 9.2		20.5 ± 6.5			20.0 ± 10.5	

OLB = open lung biopsy; APACHE II = Acute Physiology and Chronic Health Evaluation score; COLD = chronic obstructive lung disease Duration of mechanical ventilation before OLB or autopsy

also observed histologic CMV pneumonia in patients with cancer. In most cases, documented infection with CMV in nonimmunosuppressed hosts produces little, if any, disease. 19 Although one could consider that the frequency of CMV infection fluctuates depending on immunologic disturbances observed in ICU patients, we were unable to find any published data on the incidence of CMV pneumonia in mechanically ventilated patients. It is noteworthy that ICU patients are considered hosts susceptible to bacterial or fungal infections (linked to impaired host defenses) but not viral infections. The absence of previous publications on the existence of ventilator-associated CMV pneumonia may reflect the lack of a standardized approach to the documentation of CMV pneumonia. Our diagnostic approach, which uses frequent virologic assessment of blood, urine and BAL fluid, suggests that CMV pneumonia is not an exceptional cause of VAP. In our experience, CMV pneumonia could develop in all categories of patients (multiple trauma, chronic obstructive lung disease, and surgical patients). It must be noted that the true incidence of CMV pneumonia could not be drawn from our study. Indeed, open-lung biopsy and autopsies were performed essentially to locate the cause of a respiratory failure when all bacteriologic cultures were negative. Indirect findings suggest that CMV infection could play an important role in the morbidity and mortality of ICU patients. For example, Domart et al.20 have shown that, in patients with mediastinitis after cardiac surgery, mortality was higher for patients with viral shedding than for patients without

The mechanisms of acquisition of CMV pneumonia during ICU stays remain unknown. Human CMV is CYTOMEGALOVIRUS AND

Table 2. Laboratory Abnormal Ventilator-Associated Cytomes

Leukocyte count (g · L-1) Neutrophils (%) Lymphocytes (%) Mononucleosis picture of the peri blood smear Hemoglobin (g/dl) Trombocyte count (g Pão₂ (mmHg) Pa₀/Fi₀₂ (mmHg) Paco₂ (mmHg) ASAT > 2× normal ALAT > 2x normal Serum bilirubin (µmol/I) >15 x normal Serum creatinine (µmol/I) >120 µmol/1 Values are mean ± SD (range

aymptomatic and silent, al and produce lesions at any host. There is also a growi ogenous transmission of C tients, with the risk of acq to the number of transfuse evidence for nosocondial tr demonstrated, 22 Faix sho jects may retain live wirus fectious ability for several This possibility stresses the fore and after contact wit environmental objects po

fected secretions.

thought to infect 70% of a

Clinical and Radiologic associated Cytomegalor We found that clinical differentiating CMV from b be noted, however, that th were diagnosed after a lo Ventilation (median, 18 d VAP as reported in the lite played only a small role in nonia. All ICU patients had abnormality at baseline, pro

Inesthesiology, V 84, No 2, Feb 7

[†] Blood transfusions from unrelated volunteers unscreened for CMV received before the development of CMV pneumonia.

[‡] Acute exacerbation of COLD

[§] Major surgery.

Table 2. Laboratory Abnormalities of the 25 Patients with Ventilator-Associated Cytomegalovirus Pneumonia

	ALTERNATION AND DESIGNATION OF
Leukocyte count (g·L ⁻¹)	$15.1 \pm 6.7 (7.6 - 38.0)$
Neutrophils (%)	$83 \pm 7 (68-97)$
Lymphocytes (%)	$9 \pm 4 (2-17)$
Mononucleosis picture of the peripheral blood smear	3 patients
Hemoglobin (g/dl)	$9.4 \pm 1.8 (7.2 - 13.8)$
Thrombocyte count (g ⋅ L ⁻¹)	$220 \pm 144 (30-552)$
Pa _{O2} (mmHg)	72 ± 16
Pa _{O2} /Fi _{O2} (mmHg)	115 ± 35
Pa _{CO₂} (mmHg)	47 ± 12
ASAT	$31 \pm 24 (9-89)$
> 2× normal	4 patients
ALAT	48 ± 54 (7-194)
> 2× normal	5 patients
Serum bilirubin (µmol/l)	$54 \pm 73 (5-339)$
$> 1.5 \times normal$	10 patients
Serum creatinine (µmol/l)	$121 \pm 91 (41-483)$
>120 μmol/l	10 patients

Values are mean ± SD (range).

ood fusiont

'es

'es

'es 'es

es es

es

es

es

es es

'es

'es

10

ю

10

lo 'es

'es

'es

'es

10

'es

'es

ur ex-

cate-

uctive

noted

ld not

piopsy

te the

ologic

st that

e mor-

e, Do-

edias-

er for

ithout

monia

CMV is

ASAT = aspartate aminotransferase; ALAT = alanine aminotransferase.

thought to infect 70% of adults but generally remains asymptomatic and silent, although it may be reactivated and produce lesions at any time during the life of the host. There is also a growing body of evidence for exogenous transmission of CMV even in seropositive patients, with the risk of acquiring viral infection linked to the number of transfused units. ²¹ Although a lack of evidence for nosocomial transmission of CMV has been demonstrated, ²² Faix ²³ showed that contaminated objects may retain live virus and therefore potential infectious ability for several hours after contamination. This possibility stresses the need for hand washing before and after contact with CMV-infected subjects or environmental objects possibly contaminated by infected secretions.

Clinical and Radiologic Aspects of Ventilatorassociated Cytomegalovirus Pneumonia

We found that clinical features were not useful in differentiating CMV from bacterial infection. It should be noted, however, that the cases of CMV pneumonia were diagnosed after a longer period of mechanical ventilation (median, 18 days) than cases of bacterial VAP as reported in the literature.²⁴ Chest radiograph played only a small role in the diagnosis of CMV pneumonia. All ICU patients had some type of radiographic abnormality at baseline, probably reflecting interstitial

edema, atelectasis, scarring, or other postoperative changes. Episodes of CMV pneumonia were superimposed upon baseline radiographic abnormalities, making it difficult to identify subtle changes. However, we found that radiographic infiltrates were generally bilateral (median Weinberg's score, 9) associating interstitial and alveolar infiltrates.

Nonhistologic Diagnostic Procedures in the Diagnosis of Ventilator-associated Cytomegalovirus Pneumonia

As clinical and radiologic signs lack specificity and even sensitivity, it would seem necessary to use virologic methods to diagnose CMV pneumonia. Rapid diagnostic techniques (e.g., centrifugation cultures, which are read within 16-48 h) could be performed on BAL, blood, and urine products. These techniques are more useful in the treatment of ICU patients than conventional cultures, which require 1-4 weeks to diagnose CMV. The sensitivity of these techniques varies in the literature and has not been evaluated in mechanically ventilated patients. Thus, comparing a shell vial culture technique with conventional viral culture of lung tissue in marrow transplant patients, Crawford et al.25 found a sensitivity of 96% with this rapid culture technique. Other authors have found that between 60% and 70% of the specimens positive by shell vial assay yield CMV by conventional culture method. 26-28 It has even been reported that shell vial assay could be negative in 59% of the blood specimens that are positive with conventional tube cultures.²⁸ Our results showed that sensitivity of shell vial assay was low, 53% for BAL culture and 20% or less for blood and urine cultures. Conversely, and as published previously, 25 we found a good specificity for the shell vial assay (92%). Some recent diagnostic procedures are available for clinicians. For example, Erice et al.29 reported that CMV antigenemia assay was significantly more sensitive than shell vial cultures of CMV in the polymorphonuclear leukocyte fraction of blood leukocytes. The CMV antigenemia assay is relatively simple to perform and may be completed in 5-6 h.

Table 3. Evolution of the 25 Patients with Ventilatorassociated Cytomegalovirus Pneumonia

Ganciclovir treatment	8 patients (mortality 50%)
Duration of gancyclovir therapy (8	
patients) (days)	18 ± 8 (6–31)
Duration of mechanical ventilation (days)	31 ± 22 (7-90)
Duration of hospitalization (days)	45 ± 34 (15-150)

97:18-22

1984; 154:169-71

patients with mediastinitis follo

21. Wilhem JA, Matter L, Sho

tomegalovirus to patients recei

22. Adler SP, Baggett J, Wilson

epidemiology of cytomegalovir

nosocomial transmission [9] Ped

23. Faix RG: Survival of cyt

faces. J Pediatr 1985; 1085:649

24. Langer M, Moscoffi P, C

Long-term respiratory support

patients. Am Rev Respir Dis 19

25. Crawford SW, Bowden F

D, Clark JG: Rapid detection of

by bronchoalveolar lavage and

26. Paya CV, Wold AD, Smi

infections in specimens other

conventional tube cell aulture

odf/84/2/280/648823/0000542-199602000-00005.pdf by guest on 05 April 2024

Med 1988; 108:180-5

5. Knaus WA, Draper EA, Wagner DP, Zimmerman JE: APACHE II: A severity of disease classification system. Crit Care Med 1985; 13:

6. Weinberg PF, Matthay MA, Webster RO, Roskos KV, Goldstein IM, Murray JF: Biologically active products of complement and acute lung injury in patients with the sepsis syndrome. Am Rev Respir Dis

7. Smyth RL, Scott JP, Borysiewicz LK, Sharples LD, Stewart S, Wreghitt TG, Gray JJ, Higenbottam TW, Wallwork J: Cytomegalovirus infection in heart-lung transplant recipients: Risk factors, clinical associations, and response to treatment. J Infect Dis 1991; 164:1045-

8. Miller RR: Viral infections of the respiratory tract, Pathology

Jarlier V, Le Charpentier Y, Grosset J, Viars P: Nosocomial bronchopneumonia in the critically ill. Am Rev Respir Dis 1992; 46:1059-

J, Ramirez J, Xaubet A, Ferrer M, Rodriguez-Roisin R: Validation of different techniques for the diagnosis of ventilator-associated pneumonia. Comparison with immediate postmortem pulmonary biopsy.

11. Martin II WI. Smith TF: Rapid detection of cytomegalovirus in bronchoalveolar lavage specimens by a monoclonal antibody

12. Mazeron MC, Jahn G, Plachter B: Monoclonal antibody E-13 (M-810) to human cytomegalovirus recognizes an epitope encoded by exon 2 of the major immediate early gene. J Gen Virol 1992; 73:

13. Jiwa M, Steenbergen RDM, Zwaan FE, Kluin PM, Raap AK, van der Ploeg M: Three methods for the detection of cytomegalovirus in lung tissue of patients with interstitial pneumonitis. Am J Clin Pathol

14. Wallace JM, Hannah J: Cytomegalovirus pneumonitis in patients with AIDS. Findings in an autopsy series. Chest 1987; 92:198-

15. Theise ND, Haber MM, Grimes MM: Detection of cytomegalovirus in lung allografts: Comparison of histologic and immunohistochemical findings. Am J Clin Pathol 1991: 96:762-6

16. Schulman LL, Reison DS, Austin JHM, Rose EA: Cytomegalovirus pneumonitis after cardiac transplantation. Arch Intern Med 1991; 151:1118-24

17. Cordonnier C, Escudier E, Nicolas JC, Fleury J, Deforges L, Ingrand D, Bricout F, Bernaudin JF: Evaluation of three assays on alveolar lavage fluid in the diagnosis of cytomegalovirus pneumonitis

18. Oda Y, Katsuda S, Okada Y, Kawahara El, Ooi A, Kawashima A, Nakanishi I: Detection of human cytomegalovirus, Epstein-Barr virus, and herpes simplex virus in diffuse interstitial pneumonia by polymerase chain reaction and immunohistochemistry. Am J Clin Pathol 1994; 102:495-502

19. Pass R: Epidemiology and transmission of cytomegalovirus. J

20. Domart Y, Trouillet JL, Fagon JY, Chastre J, Brun-Vézinet F, Gibert C: Incidence and morbidity of cytomegaloviral infection in

One of the most important features of the antigenemia assay is that it can be quantified and that high levels of antigenemia appear to correlate with CMV disease.30 Polymerase chain reaction has found an ever-increasing number of clinical applications, including CMV diagnosis. The method has some major pitfalls. Owing to its high sensitivity, even a small trace of contaminating DNA can cause false-positive results. False-negative results may be caused by the genetic variability of clinical strains of CMV, because altered nucleotide sequence may prevent annealing of the primers. Development of a rapid and sensitive double polymerase chain reaction to detect conserved sequences from the immediate early gene of human CMV could help to differentiate latency from active infection.31 We are currently evaluating antigenemia and double polymerase chain reaction for the diagnosis of CMV pneumonia in ICU pa-

Is CMV infection of the lung a clinically significant process justifying specific antiviral therapy? If so, are conventional pulmonary diagnostic techniques sufficiently sensitive and do they provide for early detection of CMV pneumonia to initiate an optimal treatment program? Discrepancies between our results and those of other investigators concerning the existence of CMV as a causative agent of VAP indicate that additional studies are needed to determine the incidence and mortality of CMV pneumonia in patients whose lungs are mechanically ventilated. These studies will no doubt use shell vial culture techniques, antigenemia, and polymerase chain reaction assay. The main problem is that none of these techniques has been validated in the diagnosis of ventilator-associated CMV pneumonia. Nevertheless, these new techniques may not necessarily increase specificity in diagnosing CMV pneumonia. Under these circumstances, precise diagnosis of CMV pneumonia continues to rely on documenting CMV inclusions on tissue specimens obtained by open-lung biopsy or transbronchial lung biopsy and improved methods for diagnosing CMV pneumonia are required.

References

- 1. Slade MS, Simmons RL, Yunis E, Greenberg LI: Immunodepression after major surgery in normal patients. Surgery 1975; 78:363-72
- 2. Zimmerli W: Impaired host defence mechanisms in intensive care unit patients. Intensive Care Med 1985; 11:174-8
- 3. Anderson LJ: Major trends in nosocomial viral infections. Am J Med 1991; 91(suppl 3B):108S-11S
- 4. Schuller D, Spessert C, Fraser VJ, Goodenberger M: Herpes simplex virus from respiratory tract secretions: Epidemiology, clinical

characteristics, and outcome in immunocompromised and nonim-

munocompromised hosts. Am J Med 1993; 94:29-33

818-29

1984; 130:791-6

of the Lung. Edited by Thurlbeck WM, Churg AM. New York, Thieme Medical, 1995, pp 195-222

9. Rouby JJ, Martin De Lassale E, Poette P, Nicolas MH, Bodin L,

10. Torres A, El-Ebiary M, Padro L, Gonzalez J, Puig De La Bellacasa Am J Respir Crit Care Med 1994; 149:324-31

method. J Clin Microbiol 1986; 23:1006-8

1990; 93:491-4

after bone marrow transplantation. J Infect Dis 1987; 155:495-500

Infect Dis 1985; 152:243-8

Anesthesiology, V 84, No 2, Feb 1996

CYTOMEGALOVIRUS AND VENTILATOR-ASSOCIATED PNEUMONIA

patients with mediastinitis following cardiac surgery. Chest 1990; 97:18-22

- 21. Wilhem JA, Matter L, Shopfer K: The risk of transmitting cytomegalovirus to patients receiving blood transfusions. J Infect Dis 1984: 154:169–71
- 22. Adler SP, Baggett J, Wilson M, Lawrence L, McVoy M: Molecular epidemiology of cytomegalovirus in a nursery: Lack of evidence for nosocomial transmission. J Pediatr 1986; 108:117–23
- 23. Faix RG: Survival of cytomegalovirus on environmental surfaces. J Pediatr 1985; 106:649–52
- 24. Langer M, Mosconi P, Cigada M, Mandelli M, and ICUGIC: Long-term respiratory support and risk of pneumonia in critally ill patients. Am Rev Respir Dis 1989; 140:302–5
- 25. Crawford SW, Bowden RA, Hackman RC, Gleaves CA, Meyers JD, Clark JG: Rapid detection of cytomegalovirus pulmonary infection by bronchoalveolar lavage and centrifugation culture. Ann Intern Med 1988; 108:180–5
- 26. Paya CV, Wold AD, Smith TF: Detection of cytomegalovirus infections in specimens other than urine by the shell vial assay and conventional tube cell cultures. J Clin Microbiol 1987; 25:755–7

- 27. Manez R, St George K, Linden P, Martin M, Kusne S, Grossi P, Ho M, Rinaldo C: Diagnosis of cytomegalovirus infections by shell vial assay and conventional cell culture during antiviral prophylaxis. J Clin Microbiol 1994; 32:2655–9
- 28. Landry ML, Ferguson D: Comparison of quantitative cytomegalovirus antigenemia assay with culture methods and correlation with clinical disease. J Clin Microbiol 1993; 31:2851–6
- 29. Erice A, Holm MA, Gill PC, Henry S, Dirksen CL, Dunn DL, Hillam RP, Balfour HH Jr: Cytomegalovirus antigenemia assay is more sensitive than shell vial cultures for rapid detection of cytomegalovirus in polymorphonuclear blood leukocytes. J Clin Microbiol 1992; 30:2822–5
- 30. van der Bij W, Schirm J, Torensma R, van Son WJ, Tegzess AM, The TH: Comparison between viremia and antigenemia for detection of cytomegalovirus in blood. J Clin Microbiol 1988; 26:2531-5
- 31. Brytting M, Sundqvist VA, Stalhandske P, Linde A, Wahren B: Cytomegalovirus DNA detection of an immediate early protein gene with nested primer oligonucleotides. J Virol Methods 1991; 32:127–38

ovirus. J

ezinet F, ection in

nonim-

CHE II:

35; 13:

ldstein

dacute

pir Dis

wart S.

lovirus

linical

1045-

hology

hieme

odin L,

oncho-

1059-

ellacasa tion of l pneupiopsy.

lovirus itibody

ly E-13 ncoded 92; 73:

AK, van virus in Pathol

in pa-2:198–

omegaunohis-

alovirus 1991;

orges L, says on monitis 95–500 washima ein-Barr nonia by