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The Effect of Volatile Anesthetics on the pH Dependence of Calcium Uptake by Cardiac Sarcoplasmic Reticulum

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The effect of volatile anesthetics (VA) on the pH dependence of calcium uptake by cardiac sarcoplasmic reticulum (SR) was studied. SR was incubated at 37° C with 45CaCl2 in the control state (no anesthetic) and in the presence of each of the VA from pH 6.6-7.6. The VA used were: halothane, 1.3%; enflurane, 1.8%; and isoflurane, 1.2%. In the control state, the initial rate of calcium uptake, measured after a 2-min incubation, was maximal at pH 6.8 (mean \pm SEM: 665 \pm 37 nmoles/mg) and markedly inhibited at pH 7.6 (107 \pm 9 nmoles/mg). In the presence of the VA, the calcium uptake rate was mildly depressed (7-32%) at pH 6.6-6.8, unchanged at pH 7.0, and greatly enhanced (52-78%) at pH 7.2-7.6, when compared to control. The maximal uptake of calcium by the SR at a calcium concentration of 10⁻⁶M, measured by a 20-min incubation, had a similar pH dependence in the control state, with a decline first evident at pH 7.2 and a 50% drop in the maximal uptake of calcium from pH 7.0-7.6. The presence of the VA was associated with a uniform depression of the maximal uptake of calcium by the SR at all pH levels measured. In view of these findings, it appears that pH does affect SR function in the presence of VA. This alteration of the pH effect by VA may be a factor responsible for discrepancies in results previously reported by investigators studying the effects of VA on the uptake of calcium by the SR. (Key words: Acid-base equilibrium: acidosis; alkalosis. Anesthestics, volatile: enflurane; halothane; isoflurane. Heart: contractility. Ion, calcium: transport; uptake. Muscle, cardiac: sarcoplasmic reticulum.)

THE SARCOPLASMIC RETICULUM (SR) is one of the organelles involved in the regulation of calcium concentration in the myocardial cell.¹⁻³ The SR is involved in the release and reuptake of myoplasmic calcium during contraction, and is one of the possible sites at which the volatile anesthetics might act to depress cardiac contractility. Previous investigations of the effects of volatile

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anesthetics on the uptake of calcium by the SR have produced varying results, ranging from increased calcium uptake,^{4,5} to decreased calcium uptake,^{4,6} and, in some cases, little to no effect on calcium uptake. This is probably due to the fact that the experimental conditions vary in terms of SR preparations, pH, incubation temperature, and concentrations of adenosine 5'-triphosphate (ATP) and calcium. Since the uptake of calcium by cardiac SR in the control state (absence of anesthetic) is known to be pH dependent, with increased uptake at acidic pH and inhibition at alkaline pH, $^{8-12}$ we felt that different bH conditions may affect SR function in the presence of anesthetics. Knowledge of the effect of different bH conditions on calcium uptake in the presence of the volatile anesthetics may help sort out the discrepancies of previous reports. Therefore, we examined the effect of the volatile anesthetics, halothane, enflurane, and isoflurane, on calcium uptake by cardiac SR from pH 6.6–7.6.

Materials and Methods

Cardiac SR was prepared from adult albino rabbit hearts using a modification of the method of Harigaya and Schwartz.¹³ We utilized rabbit cardiac SR because we wanted a species in which we could perform complimentary biochemical and physiological experiments, and compare them to previous studies.^{6,10,11,14} Protein concentration was determined by the Coomasie binding method using serum albumin as the standard.¹⁵ Water used for buffers was deionized and glass distilled.

Calcium uptake experiments were performed in 31-ml glass vials. The reaction medium consisted of 100 mM KCl, 5 mM MgCl₂, 5 mM sodium azide, 5 mM potassium oxalate, 5 mM ATP, 25 mM piperazine-N,N'-bis-2-ethane-sulfonic acid (PIPES), 25 mM 4-(2-hydroxyethyl)-1-piperazine sulfonic acid (HEPES), 0.1 mM ethylene glycol bis (beta-amino ethyl ether)-N,N,N',N'-tetraacetic acid (EGTA), and 0.1 mM CaCl₂ with ⁴⁵CaCl₂ (150,000 cpm/sample). The temperature of the reaction medium was brought to 37° C, and then the reaction was started with the addition of 25–50 micrograms of SR. The total volume was 1 milliliter.

Experiments were performed in the control state (no

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anesthetic) and in the presence of each anesthetic (1.3% halothane, 1.8% enflurane, and 1.2% isoflurane), separately. Only one concentration was used for each anesthetic because the focus was on the changes with pH, rather than changes with different anesthetic concentrations. Volatile anesthetic concentrations were determined by infrared spectroscopy, as previously described.16 Experiments done in the presence of anesthetics had the volatile anesthetic added using a Hamilton microliter syringe at the same time the SR was added via a micropipette. Immediately after the VA and SR were added, the vials were sealed with teflon caps and incubated at 37° C. Experiments were done in triplicate at each pH ranging from 6.6-7.6 in increments of 0.2 pH units. The pH level was checked prior to and after each experiment, and varied less than 0.04 pH units during the course of the incubation.

In order to determine the initial rate of calcium uptake and the maximal uptake of calcium by the SR, timed experiments, using ⁴⁵CaCl₂, were done at each pH with samplings of the reaction medium taken at 0, 2, 4, 6, 8, 10, 20, and 30 min. In subsequent experiments, the initial rate of calcium uptake was estimated by a 2-min incubation, and the maximal uptake of calcium by the SR was estimated by a 20-min incubation. The incubation was terminated by filtration of an aliquot of the reaction medium onto a glass fiber filter on a Millipore filtration apparatus. The filter was immediately rinsed with 15 ml of 10 mM CaCl₂, and counted in a scintillation vial with 5 ml of scintillation counting cocktail.

Estimations of free ions and ligands in our reaction medium at each pH were calculated using the computer program of Fabiato and Fabiato. 17 The buffering of the calcium concentration by EGTA is known to vary with pH, with greater binding of calcium by EGTA at more alkaline pH. 18 To account for the differences in calcium ion concentration at different pH levels, we employed a calcium electrode,19 and added calcium chloride to maintain a constant free calcium concentration at 5 $\times 10^{-6}$ M for each pH in the control state. Calcium uptake by cardiac SR, as measured by the calcium electrode, was performed at pH 6.6-7.6 to verify that anesthetic effects were not due to the small change in free calcium in the reaction mixture. Additional CaCl2 was added at pH 7.4 and 7.6 to keep the free calcium concentration constant, since there is greater binding of calcium to the ligands (EGTA and ATP) at the higher pH. The pH dependence of calcium uptake, determined by this method, was identical to that found using ⁴⁵CaCl₂. In the ⁴⁵CaCl₂ experiments, the free calcium concentration ranged from 0.8×10^{-6} M to 8×10^{-6} M.

All reagents used were analytical grade. HEPES and PIPES, nonionic buffers, and ATP were purchased

from Calbiochem-Behring Corp., American Hoechst Corp. 45CaCl₂, 10 mCi/ml, was purchased from New England Nuclear Co. GFF glass fiber filters were purchased from Whatman, Inc. Liquid scintillation counting was performed in a Beckman LS 2800 scintillation counter using Complete Counting Cocktail 3a70 purchased from Research Products International, Corp. Thymol-free halothane was a gift from Halocarbon Laboratories (Hackensack, NJ), enflurane was purchased from Ohio Laboratories (Madison, WI), and isoflurane from Anaquest (Madison, WI). The calcium electrode was purchased from Ionetics (Costa Mesa, CA). The pH meter, Model 4500, was purchased from Beckman Instruments, Inc. (Fullerton, CA), and the pH electrode was purchased from Markson Science, Inc. (Del Mar, CA).

Statistical analysis of the data was done using three-way analysis of variance, the factors being pH, SR preparation, and experimental condition (control vs. anesthetic). This was done to determine the significance of the difference in calcium uptake in the control state and in the presence of anesthetics and the effect of pH on calcium uptake in the presence and absence of anesthetic. Testing of both between and within groups was done using paired t tests to determine which of the group means differed. Significance was determined at P < .05.

Results

Figure 1 is an example of the time dependence of calcium uptake at pH 6.6, 6.8, and 7.4. As is shown in figure 1, the uptake of calcium is linear up to about 3 min. The reaction reaches a plateau at 20 min. With this information, we approximated the rate of calcium uptake from a 2-min incubation, and maximal calcium uptake (at a calcium concentration of 10^{-6} M) from a 20-min incubation.

TWO-MINUTE INCUBATION

Figure 2 shows the effect of volatile anesthetics on the rate of calcium uptake by cardiac SR after a 2-min incubation as a function of pH. A three-way analysis of variance of these data shows a highly significant (P < .0001) difference in the rate of calcium uptake in the control state when compared to calcium uptake in the presence of anesthetics. It also shows that the changes in calcium uptake as a function of pH are highly significant (P < .0001). The interaction of experimental condition (control vs. anesthetic) and pH is also highly significant (P < .0001), indicating that the control SR responds differently to pH than the SR in the presence of anesthetics.

FIG. 1. Time dependence

of calcium uptake using

decreases the plateau. The plateau represents the maximal uptake of calcium by the SR. Each data point is the mean of three experimental determinations. Error bars represent standard error of the mean. Significant differences are as noted in "Results."

The analysis of variance supports the results of paired t tests (table 1), which show that all three anesthetics significantly (P < .001) attenuate the inhibition of calcium uptake seen in the control state at pH 7.2–7.6. The anesthetics produce a 52–78% increase in the rate of calcium uptake, when compared to the control state (fig. 2).

The effect of the anesthetics at pH 6.6–6.8, in contrast to the higher pH, is to decrease the rate of calcium uptake, when compared to control. At pH 6.6, all three anesthetics significantly decrease the rate of calcium uptake, when compared to the control state. At pH 6.8, both halothane and isoflurane significantly decrease the rate of calcium uptake, while enflurane has no significant effect when compared to control. There is little

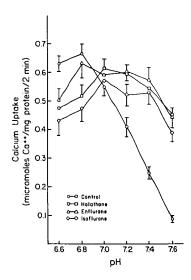


Fig. 2. The pH dependence of the rate of calcium uptake (2 min incubation) in the control state (circles) and in the presence of the volatile anesthetics, 1.3% halothane (squares), 1.7% enflurane (triangles), and 1.2% isoflurane (diamonds). Calcium uptake was measured using 45CaCl2. Experimental conditions are as stated in "Materials and Methods." Each point is the mean of nine experimental determinations performed in duplicate (n = 18). Error bars represent standard error of the mean. Significant differences are as noted in "Results" and in table 1.

change in calcium uptake at pH 7.0 in the presence of any of the three volatile anesthetics, although the small change in the presence of halothane is statistically significant (P < .02).

When the control state is removed from the analysis of variance, the response of the SR to all three anesthetics is similar with regard to changes in pH. This is supported by the results of paired t tests in table 1, which show that the differences between the anesthetics themselves are not significant, except at pH 6.8 (enflurane), pH 7.2 (isoflurane), and pH 7.4 (isoflurane vs. enflurane).

A separate set of paired t tests were done to compare the differences in calcium uptake between each pH under the different experimental conditions (control, halothane, enflurane, isoflurane). In the control state, the difference in calcium uptake with the various hydrogen ion concentrations is highly significant (P < .001), except between pH 6.6 and 6.8, where the difference is not statistically significant. In the presence

TABLE 1. The Values that P is "Less Than," Determined From Paired t tests Comparing Differences between Calcium Uptake in the Control State with Each of the Volatile Anesthetics, and Comparing Differences between the Volatile Anesthetics Themselves, After a 2-Min Incubation.

рН	C Vs. H	C Vs. E	C Vs. 1	H Vs. E	H Vs. 1	E Vs. I
6.6 6.8	.001 .01	.01 n.s.	.001 .001	n.s .01	n.s. n.s.	n.s. .001
7.0	.02	n.s.	n.s.	n.s.	n.s.	n.s.
7.2 7.4	.001	.001	.01 .001	n.s. n.s.	.01 n.s.	.05
7.6	.001	.001	.001	n.s.	n.s.	n.s.

C = control; H = halothane; E = enflurane; I = isoflurane; n.s. = not significant.

of each of the volatile anesthetics, there is no significant difference between pH 6.6 and 7.6. However, the differences between the pH 7.0–7.2 range and the two extreme hydrogen ion concentrations (pH 6.6 and 7.6) is significant (P < .001) for all three anesthetics.

TWENTY-MINUTE INCUBATION

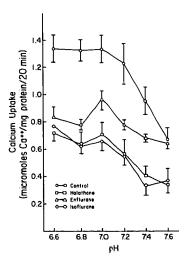
Figure 3 shows the effect of volatile anesthetics on the maximal calcium uptake by the SR after a 20-min incubation as a function of pH. A three-way analysis of variance shows a highly significant (P < .0001) difference in calcium uptake in the control state when compared to calcium uptake in the presence of anesthetic. The changes in calcium uptake as a function of pH are also highly significant (P < .0001). However, the interaction of experimental condition and pH was not significant, indicating that the SR responded to hydrogen ion concentration in a similar fashion under all conditions. This is seen in figure 3, where calcium uptake decreases with decreasing hydrogen ion concentration (increasing pH) in the control state and in the presence of each anesthetic. Therefore, although the amount of calcium uptake by the SR after a 20-min incubation was quantitatively different from calcium uptake in the presence of anesthetics, the SR responded qualitatively in a similar fashion to changes in hydrogen ion concentration, both in the control state and in the presence of anesthetic. This finding was supported by paired t tests, done to compare changes in calcium uptake as a function of pH under each condition, which showed a significant (P < .05) decrease in calcium uptake occurring at pH 7.2, both in the control state and in the presence of anesthetic.

Table 2 lists the results of t tests done to determine specific differences between the control and each of the anesthetics, and between the anesthetics themselves, at each pH. These comparisons indicate that the amount of calcium uptake by the control SR is significantly decreased by the presence of anesthetic, except for enflurane at pH 7.6. In the presence of halothane and isoflurane, the SR responds identically to changes in hydrogen ion concentration. However, the presence of enflurane produces significantly less depression of calcium uptake than halothane or isoflurane, even though the pattern of response to pH is similar. This difference in the SR response to enflurane is seen in figure 3.

Discussion

The rate of oxalate-supported, ATP-dependent calcium uptake by cardiac SR of several species in the absence of volatile anesthetics is known to be pH dependent. The highest initial calcium uptake rate occurs at pH 6.2–6.8, and inhibition of uptake occurs at

FIG. 3. The pH dependence of the maximal uptake of calcium by the SR (20 min incubation) in the control state (circles) and in the presence of the volatile anesthetics, 1.3% halothane (squares), 1.7% enflurane (triangles), and 1.2% isoflurane (diamonds). Calcium uptake is measured using 45 CaCl2. Experimental conditions are as stated in "Materials and Methods." Each point is the mean of nine experimental determinations performed in duplicate (n = 18). Error bars represent standard error of the mean. Significant differences are as noted in "Results" and table 2.



pH 7.2-7.6. Our results for the rate of calcium uptake by cardiac SR in the control state concur with these findings.

Tate et al. have shown that preincubation of cardiac SR with calcium prevents the inhibition of calcium uptake at pH 7.2–7.6. The results of our experiments show that the volatile anesthetics can also prevent the inhibition of the initial rate of calcium uptake at these pH levels. These data suggest that the volatile anesthetics, similar to calcium preincubation, remove the requirement of hydrogen ion for the uptake of calcium. It is possible that the anesthetics alter the environment of the calcium pump protein, perhaps by changing its catalytic properties.

The results of our experiments concur with previous results measuring calcium uptake by canine cardiac SR at 5mM ATP and 37° C, which showed increased calcium uptake at pH 7.3 in the presence of 1–1.13% halothane, 4 as well as in the presence of .64–2.82% enflur-

TABLE 2. The Values P is "Less Than" Determined from t tests Comparing Differences between Calcium Uptake in the Control State with Each of the Volatile Anesthetics and Comparing Differences between the Volatile Anesthetics Themselves,

After a 20-Min Incubation.

ρН	C Vs. H	C Vs. E	C Vs. 1	H Vs. E	HVs. I	E Vs. I
6.6 6.8 7.0 7.2 7.4 7.6	.001 .001 .001 .001 .001	.001 .001 .001 .01	.001 .001 .001 .001 .001	.05 .05 .05 n.s. .01	.02 n.s. n.s. n.s. .01	n.s. .01 .01 .05 .001

C = control; H = halothane; E = enflurane; I = isoflurane; n.s. = not significant.

ane and .82–1.99% isoflurane.⁵ The findings of a decrease in the maximal uptake of calcium by the SR in the presence of enflurane at pH 7.0 and halothane at pH 6.9 are also supported by previous studies, even though the concentrations used in those studies (2.5–7.5% enflurane and halothane 1.75%) are higher than those used in this study.^{4,6}

The intramyocardial pH of the perfused rabbit heart has been shown to be 7.18 by two different methods. The appears that, at the normal intramyocardial pH, the volatile anesthetics cause an increase in the velocity of calcium uptake and a decrease in the maximal calcium uptake when compared to control. These findings at the anesthetic concentrations studied do not necessarily pertain to lower or higher concentrations. It is interesting that all three anesthetics altered calcium uptake in a similar fashion, despite the fact that approximately 1 MAC halothane was compared to approximately .6 MAC isoflurane and enflurane. The similar fashion and enflurane.

In comparing the effects of the volatile anesthetics on the SR, the results show that halothane and isoflurane effect the SR in an almost identical fashion. Enflurane, on the other hand, effects the SR somewhat differently in that it has less of a depressant effect on the maximal calcium uptake by the SR at all hydrogen ion concentrations. Enflurane also has less of a depressant effect on the rate of calcium uptake by the SR, most notably at pH 6.8.

In summary, the volatile anesthetics decrease the rate of calcium uptake by cardiac SR at pH 6.6–6.8, have little effect at pH 7.0, and significantly increase the rate of calcium uptake at pH 7.2–7.6. Overall, the volatile anesthetics depress the maximal amount of calcium uptake by the SR from pH 6.6–7.6, when compared to the control state.

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References

- Rasmussen H: The calcium messenger system. N Engl J Med 314:1094-1170, 1986
- Jones LR, Cala SE: Biochemical evidence for functional heterogeneity of cardiac sarcoplasmic reticulum vesicles. J Biol Chem 256:11809–11818, 1981
- Chapman RA: Excitation-contraction coupling in cardiac muscle. Prog Biophys Mol Biol 35:1–52, 1979
- Blanck TJJ, Thompson M: Calcium transport by cardiac sarcoplasmic reticulum: Modulation of halothane action by substrate concentration and pH. Anesth Analg 60:390–394, 1981
- 5. Blanck TJJ, Thompson M: Enflurane and isoflurane stimulate

- calcium transport by cardiac sarcoplasmic reticulum. Anesth Analg 61:142-145, 1982
- Su JY, Kerrick GL: Effects of enflurane on functionally skinned myocardial fibers from rabbits. ANESTHESIOLOGY 52:385– 389, 1980
- Lain RF, Hess ML, Gertz EW, Briggs FN: Calcium uptake activity
 of canine myocardial sarcoplasmic reticulum in the presence of
 anesthetic agents. Circ Res 23:597-604, 1968
- Nakamaru Y, Schwartz A: Possible control of intracellular calcium metabolism by [H⁺]: Sarcoplasmic reticulum of skeletal and cardiac muscle. Biochem Biophys Res Comm 41:830, 1970
- Dunnett J, Nayler WG: Effect of pH on calcium accumulation and release by isolated fragments of cardiac and skeletal muscle sarcoplasmic reticulum. Arch Biochem Biophys 198:434–438, 1979
- Tate CA, Chu A, McMillin-Wood J, Van Winkle WB, Entman ML: Evidence for a calcium-sensitive factor which alters the alkaline pH sensitivity of sarcoplasmic reticulum calcium transport. J Biol Chem 256:2934–2939, 1981
- Tate CA, Van Winkle WB, Entman ML: Time-dependent resistance to alkaline pH of oxalate-supported calcium uptake by sarcoplasmic reticulum. Life Sci 27:1453-1464, 1980
- Fabiato A, Fabiato F: Effects of pH on the myofilaments and the sarcoplasmic reticulum of skinned cells from cardiac and skeletal muscles. J Physiol (Lond) 276:233-253, 1978
- Harigaya S, Schwartz A: Rate of calcium binding and uptake in normal animal and failing human cardiac muscle. Circ Res 25:781-794, 1969
- Su JY, Kerrick GL: Effects of halothane on Ca²⁺-activated tension development in mechanically disrupted rabbit myocardial fibers. Pflugers Arch 375:111-117, 1978
- Bradford MM: A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein dye binding. Anal Biochem 72:248-254, 1976
- Blanck TJJ: A simple closed system for performing biochemical experiments at clinical concentration of volatile anethetics. Anesth Analg 60:435-332, 1981
- Fabiato A, Fabiato F: Calculator programs for computing the composition of the solutions containing multiple metals and ligands used for experiments in skinned muscle cells. J Physiol (Paris) 75:463-505, 1979
- Bartfai T: Preparation of metal-chelate complexes and the design of steady-state kinetic experiments involving metal nucleotide complexes, Advances in Cyclic Nucleotide Research, Vol 10. Edited by Brooker G, Greengard P, Robison GA. New York, Raven Press, 1979, pp 219-242
- Madiera VMC: A rapid and ultrasensitive method to measure Ca⁺⁺ movements across biological membranes. Biochem Biophys Res Comm 64:870-876, 1975
- Gonzalez NC, Clancy RL: Inotropic and intracellular acid-base changes during metabolic acidosis. Am J Physiol 228:1060– 1064, 1975
- Jacobus WE, Pores 1H, Taylor GE, Nunnally RL, Hollis DP, Weisfeldt ML: Tight coupling of intracellular pH and ventricular performance. J Mol Cell Cardiol 10:39, 1978
- Flaherty JT, Weisfeldt ML, Bulkley BH, Gardner TJ, Gott VL, Jacobus WE: Mechanisms of ischemic myocardial cell damage assessed by phosphorus-31 nuclear magnetic resonance. Circulation 65:561–571, 1982
- Drummond JC: MAC for halothane, enflurane, and isoflurane in the New Zealand white rabbit: And a test for the validity of MAC determinations. ANESTHESIOLOGY 62:336-338, 1985