Laboratory Report

Failure of Enflurane and Halothane Anesthesia to Inhibit Lymphocyte Transformation in Volunteers

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Changes in the peripheral blood leukocyte count and in the ability of lymphocytes to transform in response to phytohemagglutinin were studied in healthy volunteers undergoing prolonged enflurane or halothane anesthesia without coincident surgical operation. Anesthesia was associated with a modest leukocytosis that persisted into the first postanesthetic day, primarily due to an influx of neutrophils into the circulation. There was no significant alteration, either during or following anesthesia, in the ability of the volunteers' lymphocytes to transform in response to phytohemagglutinin when compared with either preanesthetic values or unanesthetized controls. Depression of lymphocyte transformation does not appear to follow prolonged enflurane or halothane anesthesia in the absence of a surgical procedure. (Key words: Immune response, volatile anesthetics; Blood, leu-

petency. Since the lymphocyte plays a central Q role in specific immunity to foreign antigens, a deficiency in this phase of the immune response suggests a predisposition to malig- $\frac{Q}{2}$ nancy or infection.

Depression of lymphocyte transformation has been reported to occur in man after anesthesia and operation. It is demonstrable within 9 two hours of induction of anesthesia," maximal 8 in the immediate postoperative period,3 and in the immediate postoperative period," and 3 persists for as long as three weeks. This 80 reduced lymphocyte responsivenses is closely correlated to the extent of surgical trauma. The amount of blood lost, and the presence 40 period. of debilitating disease.^{3,9} It has also been suggested that anesthetic agents and techniques do not affect lymphocyte transformation 5.10; N however, the influence of general anesthesia on the absence of coincidental operation has ont been determined. In this report we describe the effects of general anesthesia with of enflurane or halothane, administered to healthy volunteers, on lymphocyte transformation and the peripheral blood leukocyte count.

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Methods and Materials

Enflurane or halothane anesthesia was ad-® ministered to unpremedicated human volunteers in San Diego. The protocol for these studies was approved by the Human Re-82 search Committees of the Veterans Adminis-4

enflurance; Anesthetics, volatile, balothane.) THE IN-VITRO DETERMINATION of the ability of lymphocytes to respond to specific antigens or nonspecific mitogens is commonly accepted as 2

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TABLE 1. Effects of Euflurane in Human Volunteers (Mean ± SE)

	Before Amesthesia	Mid-anesthesia	End of Anesthesia	One Day after Anexthesia	Five Days after Anosthesia
Lymphocyte transforma- tion (per cent of pre- anesthetic value) Anesthesia (Group I) No anesthesia (Group II)	100 100	184 ± 34.6 116 ± 6.5	162 ± 54.1 124 ± 9.7	236 ± 19.8 162 ± 56.41	205 ± 51.0 140 ± 76.84
Mitogenic index Anesthesia (Group I) No anesthesia (Group II)	4.4 ± 0.6 4.5 ± 1.0	6.8 ± 0.9 4.2 ± 0.5	8.4 ± 1.4 4.6 ± 0.9	11.2 ± 1.7 5.9 ± 2.91	10.1 ± 2.9 11.7 ± 7.51
Leukocyte count (× 1,000 mm ³ Anesthesia (Group 1) No anesthesia (Group II)	7.3 ± 0.4 8.3 ± 0.7	10.0 ± 0.4 8.7 ± 0.9	12.0 ± 0.7° 8.1 ± 0.9	9.4 ± 0.5 7.3 ± 0.0†	8.3 ± 0.9 9.5 ± 1.61
Lymphocytes (× 1,000 mm ³) Anesthesia (Group I) No anesthesia (Group II)	2.8 ± 0.2 2.4 ± 0.1	3.2 ± 0.3 2.4 ± 0.2	3.3 ± 0.2° 2.4 ± 0.2	3.1 ± 0.2 2.5 ± 0.5†	3.4 ± 0.4 3.1 ± 0.4
Hours until into culture Anesthesia (Group I) No anesthesia (Group II)	51.5 ± 0.6° 42.6 ± 4.3	50.1 ± 3.0° 38.3 ± 4.3	43.4 ± 0.5 44.2 ± 0.3	37.0 ± 4.6 39.0 ± 8.91	43.5 ± 4.5 41.0 ± 11.9†

Significantly different from corresponding unanesthetized control value (P ≤ .05).

tration Hospital and University of California, San Diego. A detailed informed consent was obtained from each subject.

Anesthesia was induced by inhalation, with end-tidal anesthetic gas concentrations maintained between 1.0 and 2.0 MAC for each anesthetic agent. Total anesthesia time was 5 to 7 hours. Seven volunteers were anesthetized with halothane in oxygen. They received no additional medication and were maintained encapnic by controlled ventilation. Nine volunteers were anesthetized with enflurane in oxygen except for two periods when the carrier gas was changed to 70 per cent nitrous oxide in oxygen. Three of the nine enflurane volunteers each received a single dose of sucemylcholine to facilitate endotracheal intubation. While controlled ventilation was used throughout most of the study to maintain Pacos at normal levels, each enflurane volunteer was allowed to breathe spontaneously for two periods, early and late during the anesthesia, to assess the effects of the anesthetic on respiratory function.

Heparinized blood was obtained from the volunteers and simultaneously from a comparable group of unanesthetized control subjects immediately before, midway through, at the gend of, one day after, and five days after fat room temperature to Seattle, where they were put immediately into culture.

Lymphocyte transformation in response to phytohemagelutinin (PHA) was assessed by the whole-blood technique of Pauly et al., 10 as outlined in detail previously. Briefly, 48 the leukocyte count was determined electronithe leukocyte count was determined electroni- & cally, the percentage lymphocytes counted, \$\frac{1}{2}\$ and a sample of whole blood diluted in unsupplemented RPMI-1640 culture medium (Grand Island Biological Co.) to a concentration of 10° lymphocytes per ml. Three-ml 80° volumes of the suspension were then pipetted of into polypropylene tubes (Falcon plastics 8 #2063) and 10 µg of PHA (Burroughs Well-9 come) were added to each of seven of the ten© tubes prepared for each blood sample. Lymphocytes were incubated for five days of at 37 C in an atmosphere of 5 per cent CO_{200}^{-1} in air. Tritiated thymidine (1 μ Ci) was added 10 to each culture vessel 24 hours prior to the end of the incubation period. The extent of lymphocyte tranformation was quantitated № in a liquid scintillation counter and expressed

 $i_{n} = 2.$

either as a percentage of the preanesthetic value for each individual or as a mitogenic index where:

Mitogenic index

 $= \frac{\text{DPM of stimulated cultures (PHA added)}}{\text{DPM of unstimulated cultures (no PHA)}}$

The lymphocytic response of halothane volunteers was also assessed independently by one of us (R.B.) in San Diego, using the method of Park and Good. These specimens were cultured immediately, avoiding any influence of delays associated with transport of cells to Scattle.

Data were analyzed statistically by Student's t test for paired data, comparing the responses during and after anesthesia with premeither values, and by Student's t test for unpaired data, comparing the response of volunteers with that of simultaneously-sampled unanesthetized controls. The level of significance was accepted as P < 0.05.

Results

Due to mailing difficulties, there was variation in the intervals from drawing the specimens to establishing the cultures. In general, delays were similar for enfluranc-anesthetized volunteers and awake controls, but they differed at the initial and mid-anesthesia sampling periods (table 1). In addition, only two specimens were received from unanesthetized controls on days 1 and 5 of the enflurance study, making these results of little values Similar time-related variability was not found in the halothane study (table 2).

General anesthesia was associated with an slight leukocytosis that persisted into the first postanesthetic day. Although this was not sign inficant for the halothane volunteers, it was valid for those receiving enflurane compared with both preanesthetic values and values for manesthetized controls. The increased leukophic cyte count was mainly due to an influx of neutrophils, although lymphocytosis was also apparent in the enflurane volunteers.

Lymphocyte transformation, expressed as percentage of preanesthetic levels, increased during enfluranc anesthesia, but was not significant. (table 1) Because awake subjected demonstrated a similar change in transformation, a normal diurnal variation is suggested. When expressed as a mitogenic index, there was a similar increased transformation during enfluranc anesthesia that was not as obvious in unanesthetized controls. Indeed, the mitogenic index was significantly greater in anesses.

Table 2. Effects of Halothane in Human Volunteers (Mean ± SE)

	Before Anesthesia	Mid-anesthesia	End of Anesthesia	One Day after Anesthesia	Five Days after Anesthesia
Lymphocyte transformation (per cent of preanesthetic value) Anesthesia (Group 1) No anesthesia (Group II)	100	111.9 ± 18.0 114.0 ± 18.6	87.9 ± 10.1 126.2 ± 19.6	87.3 ± 14.5 99.6 ± 10.9	98.1 ± 23.66 97.8 ± 18.17
Mitogenic index Auesthesia (Group I) No anesthesia (Group II)	3.1 ± 0.5 3.8 ± 0.9	3.5 ± 0.7 4.5 ± 1.0	2.8 ± 0.4 4.8 ± 0.9	2.7 ± 0.4 3.8 ± 0.8	2.2 ± 0.48 3.0 ± 0.88
Leukocyte count (×1,000/mm³) Anesthesia (Group I) No anesthesia (Group II)	6.8 ± 1.0 7.5 ± 0.4	S.1 ± 0.8 7.5 ± 0.3	9.0 ± 0.6 8.1 ± 0.4	8.9 ± 0.9 7.5 ± 0.4	7.7 ± 2.05 8.2 ± 1.49
Lymphocytes (×1,000/mm³) Anesthesia (Group I) No anesthesia (Group II)	2.1 ± 0.2 2.4 ± 0.2	2.4 ± 0.1 2.5 ± 0.1	$\frac{2.2 \pm 0.1}{2.9 \pm 0.3}$	$\begin{array}{ccc} 2.7 \pm & 0.2 \\ 2.6 \pm & 0.2 \end{array}$	2.0 ± 0.5 2.5 ± 0.3
Hours until into culture Anesthesia (Group I) No anesthesia (Group II)	38.6 ± 4.5 38.4 ± 4.5	43.7 ± 3.7 46.9 ± 5.3	42.6 ± 5.2 39.3 ± 3.6	39.4 ± 9.5 39.4 ± 9.5	44.7 ± 7.66 44.7 ± 7.66

No value was significantly different from the corresponding unanesthetized control value $(P \le 0.05)$.

thetized than in control subjects at the midanesthesia sampling period. This single statistically valid difference may be due, in part, to the large variation in times between sampling and establishing the cultures for the midanesthesia specimens.

Halothane anesthesia was associated with no significant change in lymphocyte transformation, expressed either as percentage of preamesthetic value or as a mitogenic index (table 2). At the end of anesthesia there was a tendency toward slight depression of transformation, which returned to control values by day 5. A normal diurnal variation was again seen in control subjects but was not demonstrable in volunteers given halothane. There was, however, no statistically significant difference between the responses of an exthetized and unanesthetized individuals at any time.

Similar results for halothane volunteers were obtained in our San Diego laboratories using the method of Park and Good. There was no significant alteration in lymphocyte transformation during or after halothane anesthesia when cells were cultured without the delay associated with mailing to Seattle.

Discussion

Anesthetic agents are capable of many celhilar effects, and it is reasonable to implicate them in postoperative immune deficiency states. While lymphocyte transformation is inhibited by halothane in vitro, the duration of exposure to elinically relevant concentrations must exceed 24 hours. (5.4) The present study demonstrates that halothane or enfluranc anesthesia in man is not associated with alterations in lymphocyte transformation in response to PHA. These results corroborate similar observations by Kanto¹⁵ and Cullen,⁵ who observed no abnormality of lymphocyte function when patients underwent minor operations with brief anesthesias.

Although the response of volunteers anesthetized with either agent was not associated with any significant difference from the response of unanesthetized controls, the data suggest that enflurance tends to augment transformation and halothane to reduce transformation. Assuming that inhibition of lymphocyte transformation is harmful, these differences suggest that enflurance may be more beneficial than halothane. However, the data do not allow valid comparisons of this type □ since the volunteers in the two studies were ₹ not the same and the experimental protocols ਰੁੱ differed slightly. First, total anesthetic exposure was greater for the halothane volunteers (mean ± SEM | 13.7 ± 0.8 | MAC-hours) than for the enfluranc volunteers (9.6 \pm 0.4 MAC-hours). Second, the enflurance volunteers were subjected to periods of inhalation of nitrons oxide with a reduced inspired oxygen & tension. Nitrous oxide has been associated human volunteers but not in vitro. 17 However, the reduction in vivo correlated with a the adrenal cortical response to light anesthe-8 sia rather than a direct effect of nitrous \$\bar{\bar{\geq}}\$ oxide. Third, volunteers receiving enflurane were subjected to two periods during which \mathbb{g} their Pacor's rose to more than 70 torr. This d stress was not given to the subjects receiving halothane. Finally, the time from blood⊗ collection to placement of cells into culture was more varied in the enflurane study. ₹ Although lymphocytes remain viable for at $\overline{\underline{\Phi}}$ least 57 hours at room temperature rats with little effect on T-cell responsiveness,19 subtle & differences in lymphocyte reactivity at the extremes of acceptable time intervals may, in part, account for the different trends with different drugs. However, the similarity of results when cells cultured in Seattle were compared with those cultured in San Diegood suggests that the delay in mailing samples was not a significant factor.

In conclusion, we feel that no significant immunosuppression, as assessed by leukocyte count and by PHA-induced lymphocyte transformation, has been demonstrated in volunteers anesthetized with either halothane or enflurane in the absence of operation. The impaired lymphocytic responsiveness associated with operation must therefore be seconded any to other aspects of the procedure, most probably neurohumoral responses to surgical stress. The anesthetic drugs appear to affect immune competence more by modifying the response to this surgical stress than by any direct effect on immunocompetent lymphocytes.

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