Silent Cholinesterase Gene—Report of a Family

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The enzyme, pseudocholinesterase (serum cholinesterase, acylcholine acylhydrolase, I.U.B. Commission on Enzymes 3. 1. 1. 8.), is found in many human tissues. Although the enzyme is genetically controlled, with no known biologic function, pharmacologically it hydrolyzes succinylcholine, shortening the action of this muscle relaxant. Genetically, the type and amount of pseudocholinesterase are determined by genes at two loci (E1 and E2). At locus E1, with at least four alleles, there can be the usual or normal (E1"), the atypical (E1"),1 the fluoride (E1f),2 and the silent (E1s).3 At locus E_2 , the C_5^+ variant results in a 30 per cent increase in activity of the enzyme.4

The silent gene (E18) was described in 1962.3 Patients homozygous for the silent gene (E1*E1*) have no pseudocholinesterase activity.5,6 About 64 such cases have been reported, 48 of them in Eskimos.7 Succinylcholine causes prolonged paralysis in these patients. The following case adds to those previously reported and demonstrates the importance of genetic typing of those patients suspected of having prolonged apnea resulting from succinylcholine.

REPORT OF A CASE

A 27-year-old Caucasian woman of northern European extraction was admitted to the hospital for anterior cervical fusion. Following two previous anesthesias, for lumbar laminectomy and hysterectomy, she had been very slow to awaken, and she was concerned that she might have received too much anesthesia. Although specifically questioned, the patient gave no history of problems with muscle relaxants.

Physical examination The patient was thin. and laboratory studies disclosed no abnormalities. Anesthesia was induced with thiopental, 200 mg iv, followed by succinylcholine, 40 mg, to facilitate

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endotracheal intubation. Anesthesia was main-tained with oxygen, 2l, nitrous oxide, 2l, and halothane, 0.5-0.75 per cent, through an in-line vaporizer (Fluotec). Vital signs were stable throughout the 90-minute procedure. However, the pa-tient was apneic throughout the procedure, with ventilation being controlled for a total of three hours before adequate spontaneous ventilation was maintained. Because a decrease in succinylcholine hydrolysis was suspected, a blood sample was assayed for pseudocholinesterase activity, using the method of Garry.8.9 This method distinguished atypical (Ei*) and fluoride (Eif) activity by buffer inhibition.

Review of the patient's previous anesthetic records disclosed that she had been apneic for 95 minutes (succinylcholine, 60 mg) at the lumbar laminectomy and for 75 minutes (succinylcholine, 40 mg) at the hysterectomy. A diagnosis of succinylcholine apnea had been made each time, once with laboratory confirmation, but the patient had not been informed.

No pseudocholinesterase activity was found in the patient's serum. Therefore, she is homozygous for the silent gene (E₁*E₁*).

The patient's family was tested for pseudo-cholinesterase activity. The results of the determinations and deduced genotypes are shown in table 1. A family tree is shown in figure 1. Subject III-2 is presumed to be E1"E1". Subject IV-1 could be either E,"E," or E,"E,". The patient and

TABLE I

| Generation | Subject | Pseudocholin- esterase Activity, L.U. | Deduced Genotype |
|------------|----------------------------|--|---|
| I | 1 | 5,62 | EtaEta |
| 11 | 1 2 | 2,29 2,95 | EıºEıº EıºEıº |
| III | 1 2 3 5 | 4.73 2.40 0.00 0.00 | E1ªE1ª E1ªE1ª E1ªE1ª E1ªE1ª |
| IV | 1 3 4 5 6 7 | 3.84 2.60 3.35 2.86 2.55 2.82 | E ₁ ^u E ₁ ^u or E ₁ ^u E ₁ * E ₁ ^u E ₁ * |

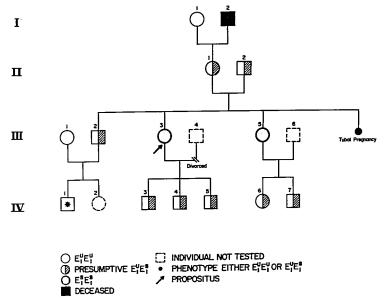


Fig. 1. Family tree of the patient (III-3), showing deduced genotypes for pseudocholinesterase in four generations.

her sister are $E_1^*E_1^*$. No atypical (E_1^*) or fluoride (E_1^*) activity was found in any sample.

Family testing of all patients with genetic deficiencies of pseudocholinesterase should be done, with counseling of the family. Some record for the deficient member, such as a letter or, preferably a medical warning bracelet, should prevent exposure to succinyleholine should the individual need anesthesia.

REFERENCES

- Kalow W, Staron N: On distribution and inheritance of atypical forms of human serum cholinesterase as indicated by dibucaine numbers. Can J Biochem 35:1305-1317, 1957
- Harris H, Whittaker M: Differential inhibition of human serum cholinesterase with fluoride: Recognition of two new phenotypes. Nature 191:496-498, 1961
- Lidell J, Lehman H, Silk E: A "silent" pseudocholinesterase gene. Nature 193:561-562, 1962

- Harris H, Robson E. B, Glenn-Bott A M: et al: Evidence for non-allelism between genes affecting serum cholinesterase. Nature 200: 1185–1187, 1963
- Rubinstein H M, Dietz L. K, Hodges L K, et al: Silent cholinesterase gene: Variations in the properties of serum enzyme in apparent homozygotes. J Clin Invest 49:479–486, 1970
- Hodgkin W E, Giblett E R, Levine H, et al: Complete pseudocholinesterase deficiency: Genetic and immunologic characterization. J Clin Invest 44:486–493, 1965
- Gutsche B B, Scott E M, Wright R C: Hereditary deficiency of pseudocholinesterase in Eskimos. Nature 215:322–323, 1967
- Garry P J: A manual and automated procedure for measuring serum cholinesterase activity and identifying enzyme variant, Differentiation by means of Tris and phosphate buffers. Clin Chem 17:183-191, 1971
- Garry P J: Serum cholinesterase variants: Examination of several differential inhibitors, salts, and buffers used to measure enzyme activity. Clin Chem 17:192–198, 1971