

Up-regulation of MicroRNA-21 Mediates Isoflurane-induced Protection of Cardiomyocytes

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ABSTRACT

Background: Anesthetic cardioprotection reduces myocardial infarct size after ischemia–reperfusion injury. Currently, the role of microRNA in this process remains unknown. MicroRNAs are short, noncoding nucleotide sequences that negatively regulate gene expression through degradation or suppression of messenger RNA. In this study, the authors uncovered the functional role of microRNA-21 (miR-21) up-regulation after anesthetic exposure.

Methods: MicroRNA and messenger RNA expression changes were analyzed by quantitative real-time polymerase chain reaction in cardiomyocytes after exposure to isoflurane. Lactate dehydrogenase release assay and propidium iodide staining were conducted after inhibition of miR-21. miR-21 target expression was analyzed by Western blot. The functional role of miR-21 was confirmed *in vivo* in both wild-type and miR-21 knockout mice.

Results: Isoflurane induces an acute up-regulation of miR-21 in both *in vivo* and *in vitro* rat models ($n = 6$, $247.8 \pm 27.5\%$ and $258.5 \pm 9.0\%$), which mediates protection to cardiomyocytes through down-regulation of programmed cell death protein 4 messenger RNA ($n = 3$, $82.0 \pm 4.9\%$ of control group). This protective effect was confirmed by knockdown of miR-21 and programmed cell death protein 4 *in vitro*. In addition, the protective effect of isoflurane was abolished in miR-21 knockout mice *in vivo*, with no significant decrease in infarct size compared with nonexposed controls ($n = 8$, $62.3 \pm 4.6\%$ and $56.2 \pm 3.2\%$).

Conclusions: The authors demonstrate for the first time that isoflurane mediates protection of cardiomyocytes against oxidative stress *via* an miR-21/programmed cell death protein 4 pathway. These results reveal a novel mechanism by which the damage done by ischemia/reperfusion injury may be decreased. (ANESTHESIOLOGY 2015; 122:795-805)

CARDIOVASCULAR disease is one of the leading causes of morbidity and mortality worldwide. Myocardial ischemia–reperfusion (I/R) injury alone accounts for approximately 450,000 deaths per year in the United States.¹ Therapies to reduce infarct size are predicted to increase patient survival rates, as final infarct size after an I/R event has been directly correlated to patient prognosis for future cardiac events.²

One way to reduce the size of infarct before an I/R injury is through the use of preconditioning. Exposure to volatile anesthetics, such as isoflurane, has been shown to decrease myocardial infarct size *in vivo* and increase cell viability after oxidative stress *in vitro*.^{3–7} Previously, we have shown that anesthetic preconditioning with isoflurane affected expression levels of nitric oxide synthase and heat shock proteins, as well as decreased levels of reactive oxygen species, resulting in increased cell viability.^{3,8} However, knowledge of the underlying mechanisms remains incomplete. In this study, we investigate whether

What We Already Know about This Topic

- MicroRNAs are 18- to 22-nucleotide sequences that suppress protein expression through degradation or suppression of messenger RNA
- MicroRNAs have previously been linked to the cardioprotective effects of ischemic, hypoxic, and heat shock preconditioning
- This study investigated whether isoflurane protects cardiomyocytes against oxidative stress through upregulation of miR-21

What This Article Tells Us That Is New

- Isoflurane directly affects microRNA expression profiles in cardiomyocytes and that miR-21 is acting to protect cardiomyocytes after isoflurane exposure, most likely through its actions on programmed cell death protein 4

microRNAs are playing a role in isoflurane-mediated protection of cardiomyocytes.

MicroRNAs are 18- to 22-nucleotide sequences that suppress protein expression through degradation or suppression of messenger RNA (mRNA). MicroRNAs have previously

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been linked to the cardioprotective effects of ischemic, hypoxic, and heat shock preconditioning.^{9–11} In 2009, the Zhang laboratory showed that overexpression of miR-21, similar to the up-regulation observed during ischemic preconditioning, significantly decreased infarct size after acute myocardial infarction.⁹ The Abdellatif laboratory has shown that miR-199a is a key regulator of hypoxia-triggered pathways, through actions on hypoxia-inducible factor-1 α and Sirtuin 1.¹⁰ MicroRNAs -378 and -711 are suppressed during both ischemic and heat shock preconditioning, resulting in increased the expression of heat shock protein 70 and increased cell viability.¹² So far, no studies have been done on microRNAs regarding anesthetic protection of cardiomyocytes.

Recent studies have also shown that isoflurane, sevoflurane, and other anesthetics can influence microRNA expression profiles in the liver and in neurons,^{13–15} suggesting that isoflurane exposure may influence microRNA in cardiomyocytes as well. Thus, in this study, we used *in vivo* and *in vitro* rat models to investigate, for the first time, the role of miR-21 in isoflurane-mediated preconditioning. We hypothesized that isoflurane protects cardiomyocytes against oxidative stress through up-regulation of miR-21.

Materials and Methods

Housing and Care of Rats and Mice

All experimental procedures and protocols were reviewed and approved by the Institutional Animal Care and Use Committee of the Medical College of Wisconsin (Milwaukee, Wisconsin; approval number 2602; 12-19-2011), and the investigation conformed to the Guide for the Care and Use of Laboratory Animals published by the United States National Institutes of Health (National Institutes of Health Publication No. 85-23, revised 1996). All animals were housed with free access to food and water. Male Wistar rats (Charles River Laboratories International, Inc., Wilmington, MA) weighing between 300 and 400 g were used for *in vivo* microRNA screening. For *in vitro* microRNA screening, time-pregnant Sprague-Dawley female rats (Jackson Laboratory, Bar Harbor, ME) were housed in separate cages until gestation was complete. Male B6129SF1/J wild-type (WT) mice (Jackson Laboratory) and miR-21 knockout mice (Jackson Laboratory) weighing between 25 and 29 g were used for *in vivo* functional studies.

Neonatal Rat Cardiomyocyte Cell Culture

Neonatal cardiomyocytes were isolated as described our in previous studies.^{8,16} Left ventricles were harvested from litters of 1- to 2-day-old Sprague-Dawley rat pups. Two litters were pooled per experiment to decrease variation within

species, as well as to ensure that enough tissue was isolated for Western blots and time course studies. Cells were isolated using serial digestion with collagenase and pancreatin (Sigma-Aldrich, St. Louis, MO) and plated on gelatin-coated dishes at 1.0×10^6 cells per 60 mm dish or 2.7×10^5 cells per well in a 12-well plate.* Cells were cultured under these conditions for 1 week before isoflurane exposure.

Exposure of Neonatal Rat Cardiomyocyte Culture to Isoflurane

Each dish of neonatal rat cardiomyocytes was randomly assigned to the isoflurane-exposed or control group. Isoflurane (Baxter Healthcare Corporation, Deerfield, IL) exposure was performed in a sealed chamber placed in a 37°C incubator for 30 min at a concentration of 0.5 mM. The same protocol has been established in our previous studies and mirrors a clinically relevant dosage.^{17,18} Control cells remained in the same incubator throughout the experiment. Isoflurane-exposed cells were removed from the sealed chamber and placed with control cells for 15 min to allow for washout of the volatile anesthetic before RNA isolation.

Exposure of Wistar Rats to Isoflurane

Each rat was randomly assigned to the isoflurane-exposed or control group. Rats were exposed to 30 min of 0.5 mM of isoflurane in a sealed plexiglass unit. Isoflurane was delivered using a VetEquip funnel-fill vaporizer (VetEquip Inc., Pleasanton, CA). A 32% O₂ environment was monitored using the Criticare Systems Poet IQ anesthesia gas monitor (SOMA Technology Inc., Bloomfield, CT). Control animals were exposed to 32% O₂ for 30 min. Left ventricles were harvested after a 15-min washout period in the original housing units. Left ventricles were pooled in groups of three for each experiment to decrease variation between groups of animals on the 12 arrays used for microRNA screening.

Total RNA Isolation and Complementary DNA (cDNA) Synthesis

Total RNA was isolated using the miRNeasy Mini Kit (Qiagen Inc., Valencia, CA). Cells were homogenized in QIAzol lysis reagent. Afterward, chloroform was added and samples were centrifuged to separate aqueous and organic phases. The aqueous phase was extracted and combined with ethanol to facilitate RNA binding to miRNeasy Mini spin columns. Samples were washed twice in buffer, and total RNA was eluted using RNase-free water. cDNA was synthesized by adding RNA to a mixture of miScript Reverse Transcriptase Mix, 10 \times miScript Nucleic Mix, and 5 \times miScript HiFlex Buffer (Qiagen Inc.). The mixture was incubated at 37°C for 60 min and then at 95°C for 5 min to convert RNA to cDNA.

MicroRNA Analysis by Quantitative Reverse Transcriptase-Polymerase Chain Reaction (qRT-PCR)

MicroRNA profiling was conducted using the miFinder miScript microRNA Polymerase Chain Reaction (PCR) Array

* Abcam neonatal rat cardiomyocyte harvest protocol. Available at: http://www.abcam.com/ps/pdf/protocols/neonatal_rat_cardiomyocyte_harvest.pdf. Accessed January 3, 2014.

(Qiagen Inc.) according to the manufacturer's instructions. These arrays contain 84 lyophilized microRNA primers distributed in single wells along with four housekeeping genes (RNAs Rnu-6, U87, 4.5 S-V1, and Y1). Samples without reverse transcriptase were run to confirm that no genomic DNA was present in the sample. The microRNAs present in this assay represent 84 of the most abundantly expressed microRNAs across all mammalian tissues. A total of 12 microRNA arrays were run for *in vivo* rat hearts and 12 for *in vitro* cardiomyocytes: six control plates and six exposed to isoflurane, as designed in previous microRNA screening studies.¹⁹ Differentially expressed microRNAs were validated in technical triplicate using miScript Primer Assays for miR-21 (Qiagen Inc., Lot Nos. 117241738 and 117259379). qRT-PCR was conducted using the Bio-Rad iCycler Real-Time PCR Detection System (Bio-Rad Laboratories Inc., Hercules, CA). cDNA was diluted and combined with QuantiTect SYBR Green PCR Master Mix and miScript Universal Primer using low-adhesion pipette tips (VWR, Radnor, PA). Data collected from these experiments defined a threshold cycle number (Ct) of detection for the microRNAs present in each sample. MicroRNAs were excluded from the study if Ct values were greater than or equal to 35. Primer specificity was tested by running qRT-PCR analysis with miScript Primer Assays specific for miR-21 in a solution containing only synthetic miR-21 mimic (Qiagen Inc.), as well as by size fractionation of stem-loop RT-PCR products on 2% agarose gel. Melt curve denaturation levels also occurred in the predicted 70° to 80°C range (see fig., Supplemental Digital Content 1, <http://links.lww.com/ALN/B128>).

Knockdown of miR-21 Expression

The neonatal rat cardiomyocytes were transfected with doses ranging from 1 to 25 nM of locked nucleic acid (LNA)-modified anti-miR-21 (Exiqon, Skelstedet, Denmark) using the Lipofectamine 2000 transfection system (Life Technologies, Carlsbad, CA). The LNAs comprise a single ribose ring connected by a methylene bridge at the 2'-O and 4'-C atoms, allowing for the nucleotides to bind in an optimal conformation. To transfect the cardiomyocytes, LNAs were incubated briefly with the Lipofectamine reagent to allow for anti-miR incorporation into lipid bubbles. The mixture was then exposed to cells for 20 h. Scrambled LNA anti-miRs (Exiqon) were used as a control. After an exposure to anti-miR-21, cells were lysed and tested for miR-21 expression using qRT-PCR analysis as described under "MicroRNA analysis by quantitative reverse transcriptase PCR" in this Materials and Methods section.

Lactate Dehydrogenase Release Assay and Propidium Iodide Staining

Cardiomyocytes were exposed to 0.5 mM of isoflurane in an anesthetic chamber, as described under "Exposure of neonatal rat cardiomyocyte culture to isoflurane" in this Materials and Methods section, while the control plates were kept in a 37°C

incubator. After a 15-min washout period, 50 μ M of hydrogen peroxide was used to cause oxidative stress to the cardiomyocytes. After 6 h of exposure to hydrogen peroxide, lactate dehydrogenase (LDH) release was measured by absorbance of indicator dyes as calculated by the Epoch Spectrophotometer (Biotek, Winooski, VT). After media samples were taken for LDH analysis, cells were stained with PI (Life Technologies). Nuclei were stained with Hoechst 33342 (Life Technologies) at a concentration of 1:800 diluted in cell culture media, and 1.0 mg/ml of PI solution was diluted in media at a concentration of 1:500. Hoechst-positive nuclei were counted as a control using fluorescence microscopy, and PI-positive cells were taken as a percentage of the control.

Short Interfering RNA Knockdown of PDCD4

Cardiomyocytes were transfected with 10 nM of programmed cell death protein 4 (PDCD4) Stealth RNAi short interfering RNA (siRNA) Select RNAi (Life Technologies, Oligo ID: RSS329752) using the Lipofectamine 2000 (Life Technologies) transfection system for 24 h. PDCD4 knockdown of mRNA expression was confirmed by qRT-PCR analysis, as described under "MicroRNA analysis by quantitative reverse transcriptase PCR" in this Materials and Methods section. Knockdown of PDCD4 protein expression was confirmed by Western blot. Negative control cells were transfected with Stealth RNAi siRNA Negative Control Med GC (Life Technologies).

mRNA Analysis of miR-21 Targets by qRT-PCR

miR-21 mRNA targets PDCD4 and phosphatase and tensin homolog (PTEN) were investigated using rat RT² PCR Assays (SABiosciences, Valencia, CA) according to the manufacturer's instructions. Total RNA was extracted using the Qiagen RNeasy kit as described under "Total RNA isolation and cDNA synthesis" in this Methods, and cDNA was prepared using the RT² First Strand Kit (Qiagen Inc.). RNA was combined with genomic DNA elimination buffer and incubated at 42°C for 5 min to ensure cDNA purity. Samples were chilled on ice for 1 min and then mixed with 5 \times RT buffer 3, primer and external control mix, RT enzyme mix 3, and water. cDNA was diluted and combined with RT² SYBR Green/Fluorescein Master Mix and distributed evenly across the wells of a 96-well plate in technical triplicates. qRT-PCR was conducted using the Bio-Rad iCycler Real-Time PCR Detection System. Samples were exposed to an initial 95°C hot-start activation step, followed by 40 cycles of 94°C denaturation and 60°C annealing/extension phases. Data collected from these experiments defined Ct values of the mRNAs present in each sample.

Expression levels of the housekeeping genes β -actin and β -2 microglobulin were used as controls to normalize samples. Samples without reverse transcriptase were run to confirm that no genomic DNA was present in the sample. mRNAs with Ct values greater than or equal to 35 were excluded from the study.

Western Blot Analysis

Cells were lysed and sonicated in RIPA lysis buffer (Cell Signaling, Danvers, MA) containing phosphatase inhibitor cocktail (Roche Diagnostics, Indianapolis, IN). Lysates were centrifuged at 10,000g for 10 min at 4°C. Pellets were discarded, and the total protein concentration of the supernatants was determined by DC Protein Assay Reagents Package kit (Bio-Rad, Hercules, CA). The samples were boiled for 5 min at 97°C. Twenty-five micrograms of protein was loaded per lane for sodium dodecyl sulfate–polyacrylamide gel electrophoresis gel separation and then transferred to nitrocellulose membrane. Membranes were blocked with blocking buffer (Thermo Fisher Scientific, Waltham, MA) and then incubated overnight at 4°C with primary antibodies against PTEN, actin (Cell Signaling), or PDCD4 (Rockland Immunochemicals, Inc., Limerick, PA). After the incubation of membranes with secondary antibodies conjugated to horseradish peroxidase (Cell Signaling) for 1 h at room temperature. Labeled proteins were detected with chemiluminescence detection reagent (Cell Signaling). The membrane was transferred to x-ray film and exposed. The intensity of the protein bands in films was quantified using ImageJ software,[†] and the data were reported as percentage of control.

Myocardial Ischemia/Reperfusion Injury In Vivo

Myocardial I/R injury *in vivo* was produced by occluding the left coronary artery, as previously described.^{20,21} WT and miR-21 knockout mice were randomly divided into isoflurane-exposed or control groups. After instrumentation was completed, all mice were stabilized for 30 min and subjected to 30 min of coronary artery occlusion followed by 2 h of reperfusion. Isoflurane was administered at 1.0 minimum alveolar concentration for 30 min *via* an isoflurane-specific vaporizer (Ohio Medical Instruments, Cincinnati, OH) followed by a 15-min washout period before coronary artery occlusion. Control mice were not exposed to isoflurane. The infarct area was delineated by perfusing the coronary arteries with 2,3,5-triphenyltetrazolium chloride *via* the aortic root, and the area at risk was delineated by perfusing phthalo blue dye (Heucotech Ltd., Fairless Hills, PA) into the aortic root after tying the coronary artery at the site of the previous occlusion.

Statistical Analysis

All rats, mice, and neonatal rat cardiomyocyte cell culture dishes were randomly assigned to isoflurane or control groups, as well as scramble or transfected groups. Western blot analysis was conducted blindly, with samples divided into randomly numbered groups. Values are expressed as means \pm SEM. *t* Test confirmed statistically significant differences between microRNAs in single groups. We used an analysis of variance test followed by a Tukey test to compare microRNAs in multiple groups. *P* values less than 0.05 (two-tailed) were considered to be significant. MicroRNA Arrays were analyzed using the miScript microRNA PCR

Array Data Analysis Web Portal (SABiosciences), and the Benjamini and Hochberg procedure was applied to control the false discovery rate in our samples.²² Three microRNAs (miRs-137, -142, and -196b) had a single data point greater than 2 SDs from the mean and were considered to be outliers. After these data points were discarded, the isoflurane-exposed groups were no longer significantly different from the control. All other statistical analysis was performed using SigmaPlot 12.0 (Systat Software, Inc., San Jose, CA).

Results

Isoflurane Exposure Causes Changes in Expression Levels of MicroRNAs

Array-based analyses investigated the effect of isoflurane on expression levels of 84 microRNAs in 36 *in vivo* rat hearts pooled into groups of three and *in vitro* cultured neonatal rat cardiomyocytes from 24 litters of Sprague-Dawley pups pooled into two litters per group (for a total of *n* = 6 per group). Eleven microRNAs were found to be differentially expressed between isoflurane and control groups. All 11 microRNAs were up-regulated with a Benjamini–Hochberg-adjusted *P* value to control the false discovery rate at 0.05 level (fig. 1A).²² Bioinformatics analysis (Ingenuity Pathway Analysis Software, Redwood City, CA) was used to nominate a candidate for further functional studies. MicroRNAs were investigated that directly targeted proteins in cell signaling pathways previously reported to mediate anesthetic preconditioning. These microRNAs were compared with the *in vivo* and *in vitro* array data, as well as against known literature.

MicroRNA-21 emerged as the strongest candidate for further functional studies, as it was linked to multiple cardioprotective cell signaling pathways, exerting effects on PDCD4, Akt, and PTEN.^{9,23–25} In addition, miR-21 amplified at the lowest Ct values of all candidate microRNAs identified, indicating a higher relative abundance, and potentially, a more robust functional effect.

miR-21 Is Transiently Up-regulated in Cardiomyocytes

Samples were taken after 30 min of isoflurane exposure, as well as after 3-, 6-, and 24-h time points to determine miR-21 expression changes over time. At 30 min after isoflurane exposure, miR-21 was significantly up-regulated ($219.2 \pm 17.1\%$). Three hours after the initial up-regulation, miR-21 expression returned to baseline values (fig. 1B). qRT-PCR sample purity was confirmed by melt curve analysis and the presence of a single PCR product after size fractionation on a 4% agarose gel (see fig., Supplemental Digital Content 1, <http://links.lww.com/ALN/B128>).

Dose-dependent Knockdown of miR-21 Expression with Lipofectamine

Neonatal rat cardiomyocytes were exposed to 20 h of multiple doses of LNA-modified anti-miR-21 or scramble antagomirs. qRT-PCR analysis revealed a dose-dependent

[†] Available at: <http://imagej.nih.gov/ij/>. Accessed December 18, 2014.

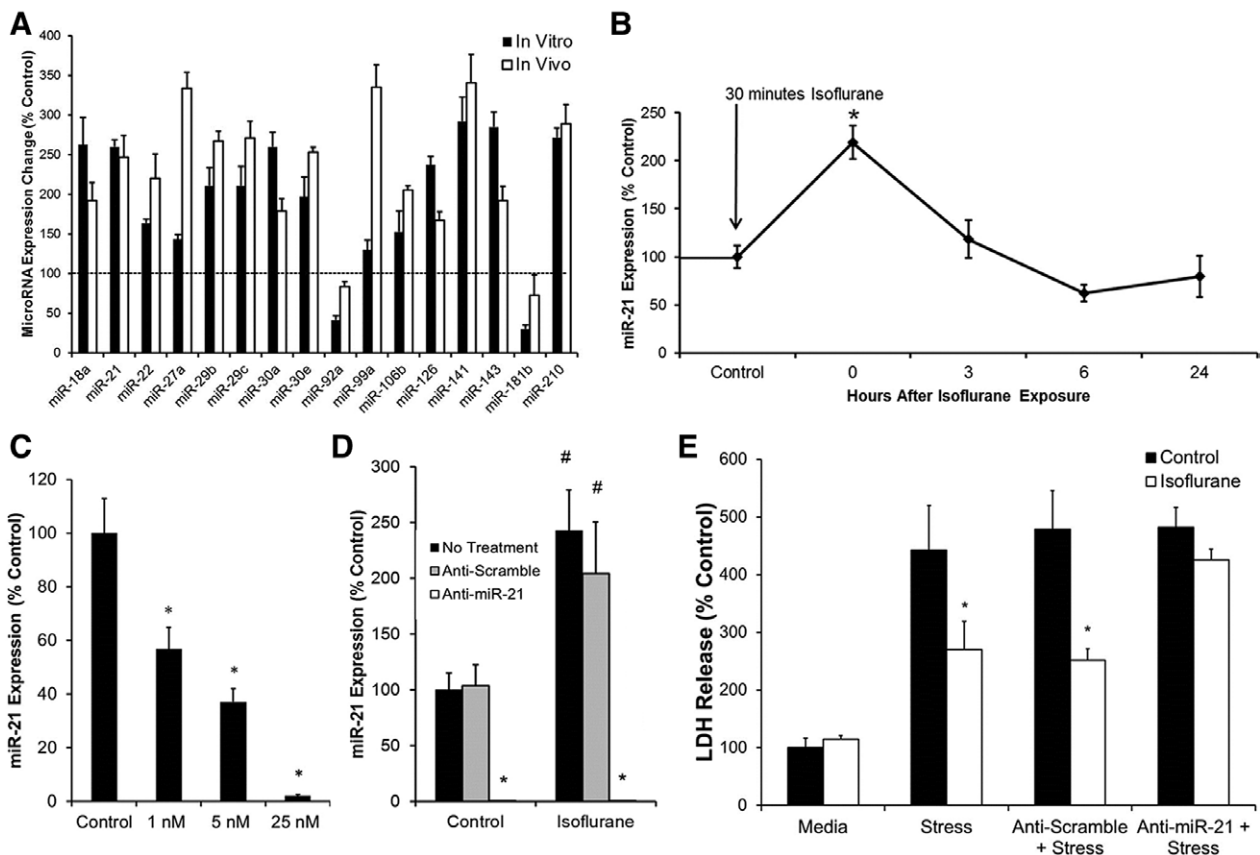


Fig. 1. MicroRNAs play an important role in isoflurane-mediated protection of cardiomyocytes. (A) Isoflurane exposure causes changes in expression levels of microRNAs. MicroRNA expression changes were measured by quantitative real-time polymerase chain reaction in the *in vitro* neonatal rat cardiomyocyte model (black bars) and in the *in vivo* rat model (white bars) after exposure to 30 min of isoflurane followed by a 15-min washout period. $n = 6$; all data = $P < 0.05$ compared with control without isoflurane exposure. (B) miR-21 is transiently up-regulated by isoflurane in cardiomyocytes. miR-21 increased significantly after a 30-min exposure to isoflurane, followed by a 15-min washout period. miR-21 expression was measured 3, 6, and 24 h after exposure. Expression decreased over time and returned to baseline after 3 h. $n = 6$, $*P < 0.05$ compared with non-isoflurane-exposed control. (C) Anti-miR-21 transfection dose-dependently knocks down the expression of miR-21. Neonatal rat cardiomyocytes were treated with 1, 5, and 25 nM of transfection reagent for 20 h. miR-21 expression was decreased in a dose-dependent manner and significantly knocked down in the 25 nM transfection group. $n = 6$, $*P < 0.05$ compared with control. (D) Anti-miR-21 transfection knocks down the expression of miR-21 after isoflurane exposure. Cultured neonatal rat cardiomyocytes were treated with media, 25 nM of microRNA scramble inhibitor, or 25 nM of anti-miR-21 for 20 h. miR-21 expression was increased in nontreated and scramble-transfected groups after isoflurane exposure. miR-21 expression was significantly knocked down in both control and isoflurane-exposed groups. $n = 6$, $*P < 0.05$ compared with nontreated and scramble-transfected controls. # $P < 0.05$ compared with non-isoflurane-exposed controls. (E) Anti-miR-21 abolishes the protective preconditioning effect of isoflurane. miR-21-transfected cells showed no significant difference in lactate dehydrogenase (LDH) release between isoflurane and control groups. Cells were exposed to 50 μ M of hydrogen peroxide for 6 h. Nontreated and scramble-transfected controls showed significantly decreased LDH release when exposed to 30 min of isoflurane before hydrogen peroxide exposure. $n = 6$, $*P < 0.05$ compared with controls.

response of miR-21 knockdown at 1 nM ($56.7 \pm 8.1\%$), 5 nM ($37.0 \pm 4.9\%$), and 25 nM ($2.1 \pm 0.6\%$) of transfection reagent (fig. 1C).

Anti-miR-21 Transfection with Lipofectamine Knocks Down the Expression of miR-21 after Isoflurane Exposure

Neonatal rat cardiomyocytes were exposed to 20 h of 25 nM anti-miR-21 or scramble antagonists. qRT-PCR analysis revealed similar baseline levels of miR-21 between nontreated and scramble-transfected groups. Nontreated and scramble-transfected groups also exhibited up-regulation of

miR-21 after isoflurane exposure similar to previous experiments. In anti-miR-21-transfected cells, both the isoflurane-exposed (0.01 ± 0.002) and control cells ($0.10 \pm 0.06\%$) exhibited significantly decreased the expression of miR-21 compared with nontransfected controls (fig. 1D), confirming a complete knockdown.

Knockdown of miR-21 Abolishes the Preconditioning Effect of Isoflurane

Measurements of cell viability after 6 h of oxidative stress with 50 μ M hydrogen peroxide revealed an increased LDH release

in nontransfected ($442.3 \pm 77.0\%$), scramble-transfected ($478 \pm 66.9\%$), and miR-21-transfected ($481.9 \pm 34.9\%$) cells compared with nonstressed control cells. Exposure to isoflurane decreased LDH release in nontransfected ($269.8 \pm 49.2\%$) and scramble-transfected ($251 \pm 20.5\%$) cells but not anti-miR-21-transfected ($425 \pm 18.1\%$) cells (fig. 1E).

Knockdown of miR-21 Abolishes the Isoflurane-induced Down-regulation of PDCD4, but Not PTEN

The mRNA expression of miR-21 targets PDCD4, and PTEN were evaluated *via* qRT-PCR. PDCD4 mRNA expression was significantly decreased after isoflurane exposure. Western blot analysis also revealed that cells exposed to isoflurane exhibited significantly decreased the expression of PDCD4 protein when compared with controls. Transfection with anti-miR-21 increased the endogenous levels of PDCD4 and abolished the isoflurane-induced down-regulation of PDCD4 (fig. 2A).

Both mRNA and Western blot analysis revealed no change in PTEN expression after isoflurane exposure. Anti-miR-21 transfection also had no effect on endogenous PTEN expression (fig. 2B).

PDCD4 siRNA Transfection with Lipofectamine Knocks Down the Expression of PDCD4

Neonatal rat cardiomyocytes were exposed to 10 nM of PDCD4 siRNA or medium guanine/cytosine content negative control siRNA. qRT-PCR analysis revealed that PDCD4 mRNA expression after siRNA knockdown was reduced to $25.7 \pm 3.0\%$ of control values (fig. 3A). Western blot analysis showed that PDCD4 protein expression was reduced to $8.8 \pm 1.3\%$ of control values (fig. 3B).

PDCD4 Inhibition Mimics the Preconditioning Effect of Isoflurane in Nontreated Cells

Measurements of cell viability after 6 h of oxidative stress with 50 μ M hydrogen peroxide revealed that anti-PDCD4 transfection caused a protective effect in cardiomyocytes that mimicked the protective effect of isoflurane. Oligo-transfected cardiomyocytes were preconditioned with isoflurane ($11.8 \pm 1.9\%$ PI-positive cells compared with $21.3 \pm 3.6\%$ of cells not exposed to isoflurane), while there was no significant difference in the protective effect of isoflurane in PDCD4-transfected cells ($7.5 \pm 2.3\%$ control cells compared with $8.6 \pm 2.1\%$ isoflurane-exposed cells). As a comparison, anti-miR-21-transfected cells were also stained with PI. Scramble-transfected cells were successfully preconditioned with isoflurane ($10.6 \pm 2.2\%$ compared with $20.0 \pm 1.3\%$ non-isoflurane-exposed cells). Transfection with 25 nM of anti-miR-21 abolished the protective effect of isoflurane ($21 \pm 2.7\%$ compared with $17.5 \pm 2.2\%$) (fig. 3C). Representative PI staining images show decreased cell viability after miR-21 knockdown and increased cell viability after PDCD4 knockdown (fig. 3D).

MiR-21 Knockout Abolishes Isoflurane-induced Decreases in Infarct Size

There were no significant differences in area at risk between WT and miR-21 knockout mice subjected to I/R injury with or without isoflurane exposure (fig. 4A). Coronary occlusion followed by reperfusion resulted in an infarct size of $59.1 \pm 4.2\%$ and $62.3 \pm 4.6\%$ in WT and miR-21 knockout mice, respectively. Pretreatment of WT mice with isoflurane significantly decreased infarct size ($44.1 \pm 2.9\%$). However, disruption of miR-21 abolished isoflurane-induced decreases in infarct size ($56.2 \pm 3.2\%$) (fig. 4B).

Discussion

In this study, we showed for the first time that isoflurane induces up-regulation of miR-21 and that this up-regulation is linked to protection of cardiomyocytes. Lipofectamine-mediated knockdown of miR-21 completely abolished the protective effect of isoflurane and resulted in increased the expression of PDCD4, a well-established target of miR-21.²³ Inhibition of PDCD4 resulted in a protective effect on control cells that mimicked the protective effect on isoflurane-treated cardiomyocytes. However, not all miR-21 targets that we investigated were suppressed. PTEN, a protein suppressed by miR-21 during ischemic postconditioning,²⁴ showed no difference in expression levels after isoflurane exposure or miR-21 knockdown during our study. From this, we conclude that anesthetic preconditioning acts through a miR-21/PDCD4-mediated pathway. This protective effect was correlated *in vivo*, with miR-21 knockout mice failing to show significant decreases in infarct size after exposure to isoflurane when compared with control mice.

Our array results showed that preconditioning with isoflurane for 30 min altered the profiles of microRNAs in cardiomyocytes (fig. 1). We used a Benjamini–Hochberg-adjusted *P* value for false discovery rate and found that 16 microRNAs were differentially expressed. When a more conservative Bonferroni correction is applied, one third of the microRNAs remain significant. In particular, isoflurane up-regulated members of the miR-30 family, as well as miR-210 and miR-21. These microRNAs have been previously linked to cardioprotective pathways.^{26–28} The miR-30 family (miR-30a, b, c, d, and e) has recently been shown to be down-regulated in response to hydrogen peroxide-induced stress in both neonatal rat ventricular cells and rat aorta vascular smooth muscle cells.²⁸ miR-210 has been shown to be up-regulated in cardiomyocytes under hypoxic conditions and confers cardioprotection.^{27,28} Especially, acute up-regulation of miR-21 was involved in ischemic and hypoxic protective signaling mechanisms in multiple cell types including cardiomyocytes.^{15,24,29} In the heart, acute up-regulation of miR-21 has been shown to decrease myocardial infarct size, increase cell viability, and indirectly up-regulate the expression of numerous cardioprotective proteins.^{25,29} Up-regulation of miR-21 has been linked to the protective effects of ischemic preconditioning in both an *in vivo* rat model and

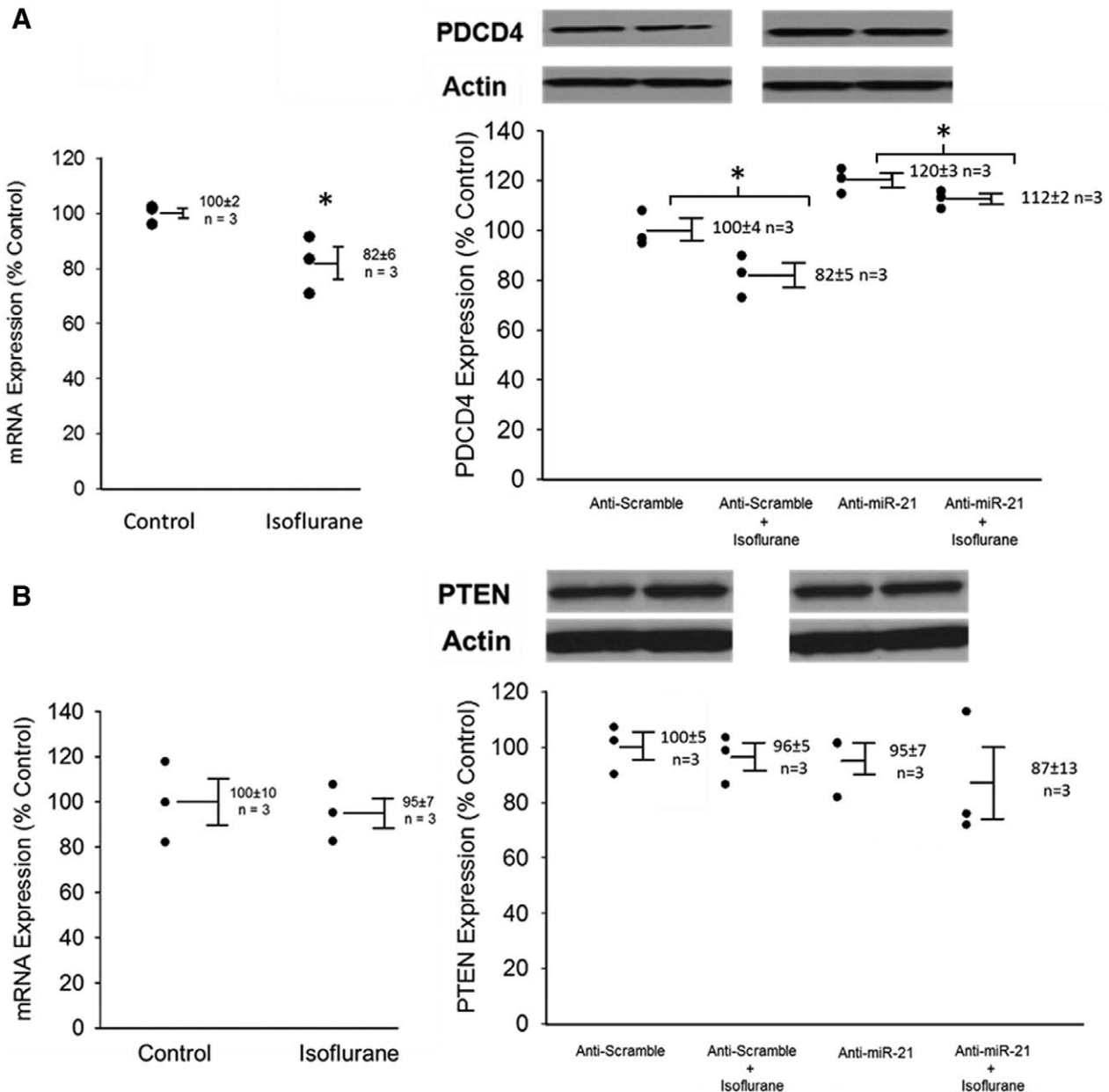


Fig. 2. Anti-miR-21 abolishes the isoflurane-induced down-regulation of programmed cell death protein 4 (PDCD4) but not phosphatase and tensin homolog (PTEN). (A) PDCD4 expression. Cells were exposed to 30 min of isoflurane followed by a 15-min washout period. Messenger RNA (mRNA) expression changes as measured by quantitative real-time polymerase chain reaction in cardiomyocytes and protein expression levels as determined by Western blot. In anti-scramble-transfected cells, miR-21 expression was significantly decreased. In anti-miR-21-transfected cells, endogenous levels of PDCD4 were increased, and the isoflurane-induced down-regulation of PDCD4 was abolished. $n = 3$, $*P < 0.05$ compared with nonisoflurane controls. (B) PTEN expression. In anti-scramble and anti-miR-21-transfected cells, isoflurane did not influence PTEN expression at either mRNA or protein levels. In addition, miR-21 knocking down did not change endogenous PTEN expression. $n = 3$.

the cultured cardiomyocytes.^{9,23} In other organs, miR-21 has been linked to protection as well. In 2013, Jia *et al.*¹⁵ showed that up-regulation of miR-21 contributed to the protective effect of xenon preconditioning before I/R injury in the mouse kidney model. These studies, as well as the link to multiple cell signaling pathways involved in anesthetic preconditioning, led us to focus our study on the dissection of the important role of miR-21 in isoflurane-mediated cardiomyocyte protection.

After isoflurane exposure, miR-21 initially increased and then decreased back to baseline levels over a 24-h period (fig. 1B). Knocking down miR-21 attenuated the isoflurane-induced decrease of LDH in cardiomyocytes exposed to hydrogen peroxide (fig. 1E), suggesting that acute up-regulation of miR-21 after isoflurane exposure is sufficient to contribute to protection of cardiomyocytes. Several studies conflict on the long-term effects of miR-21

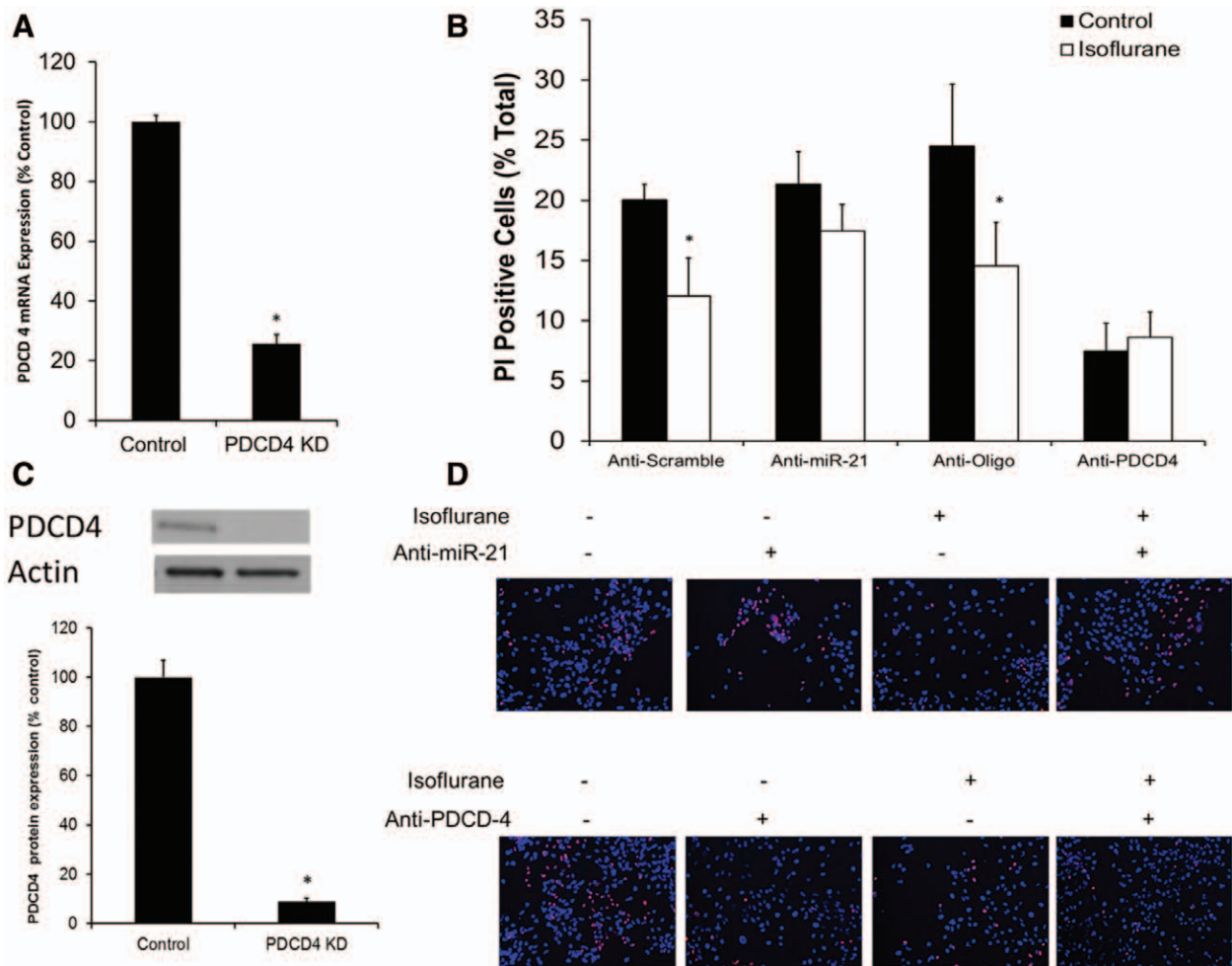


Fig. 3. Programmed cell death protein 4 (PDCD4) small interfering RNA (siRNA) transfection knocks down the expression of PDCD4. Neonatal rat cardiomyocytes were transfected with 10 nM of PDCD4 siRNA or the negative control. Twenty-four hours of transfection significantly decreased (A) siRNA knockdown of PDCD4 messenger RNA (mRNA) expression. $n = 3$, $*P < 0.05$ compared with negative control-transfected cells. (B) Anti-miR-21 knockdown and PDCD4 knockdown impair isoflurane preconditioning. All cells were exposed to 6 h of 50 μ M hydrogen peroxide. Anti-miR-21 abolished the cardioprotective effect of isoflurane, while PDCD4 knockdown mimicked the effect of isoflurane in nonexposed cells. $n = 6$, $*P < 0.05$. (C) siRNA knockdown of PDCD4 protein expression. $n = 3$, $*P < 0.05$ compared with negative control-transfected cells. (D) Representative images from propidium iodide (PI) staining.

up-regulation, but an acute up-regulation is well established as protective.^{30,31}

Our results revealed that exposure to isoflurane decreased the expression of both PDCD4 mRNA and protein levels compared with nonexposed controls. This decrease was abolished when miR-21 was knocked down. Endogenous levels of PDCD4 protein were raised, and the decrease after isoflurane exposure was not significant compared with nonexposed controls (fig. 2A). In addition, inhibition of PDCD4 expression led to protection in non-isoflurane-exposed cardiomyocytes that was similar to the preconditioning effect of isoflurane (fig. 3). It should be noted that the magnitude of PDCD4 suppression was greater than the down-regulation seen after isoflurane exposure, so it is not surprising that the amount of protection conferred is greater. This suggests that PDCD4 is the downstream target mediated by miR-21

up-regulation. PDCD4 is widely studied as a tumor suppressor³²; however, it has recently emerged as a target of miR-21 with powerful downstream effects. In renal I/R injury, miR-21 knockdown increased renal I/R injury accompanied by up-regulation of PDCD4.³³ In a cardioprotective response to chronic oxidative stress, neonatal cardiomyocytes were shown to up-regulate the expression of miR-21. Inhibition of miR-21 was linked to an increase in the expression of PDCD4 and decreased cell viability.²³

Interestingly, not all reported miR-21 targets previously implicated in preconditioning pathways were influenced by the presence of miR-21 or isoflurane. PTEN has been widely shown to be a direct target of miR-21 that is involved in cardioprotection and was initially hypothesized to be a main contributor to isoflurane-induced cardioprotection. Ischemic postconditioning showed a significant up-regulation

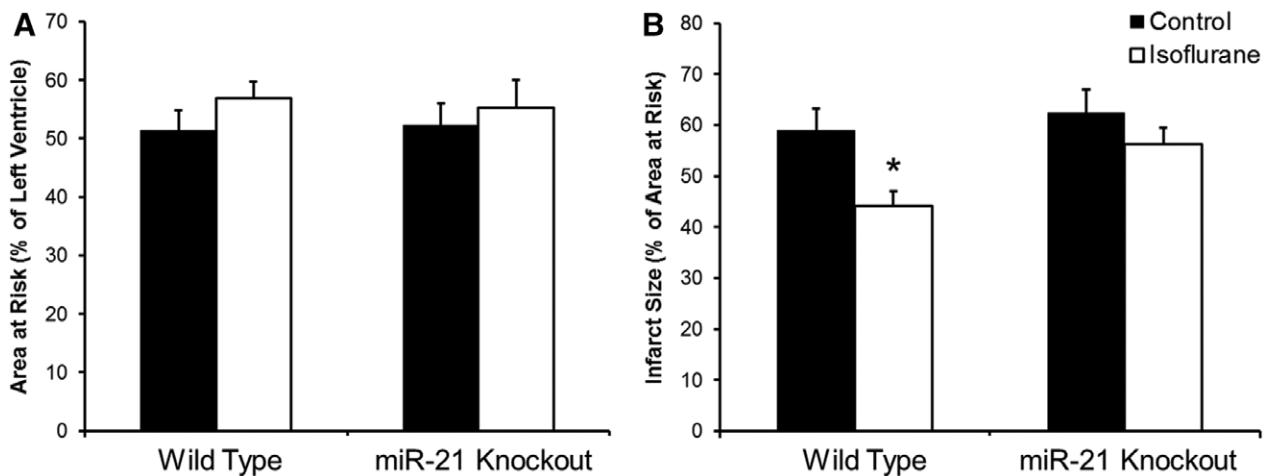


Fig. 4. Isoflurane preconditioning is abolished in miR-21 knockout mice. Wild-type (WT) and miR-21 knockout mice were exposed to 30 min of coronary artery occlusion followed by 2 h of reperfusion with and without isoflurane exposure. (A) Area at risk measurements. There were no significant differences between area at risk in control or isoflurane-exposed WT and miR-21 knockout mice. $n = 8/\text{group}$. (B) Isoflurane preconditioning is abolished in miR-21 knockout mice. WT mice were successfully preconditioned with isoflurane, while miR-21 knockout mice showed no significant difference between control and isoflurane-exposed groups. $n = 8/\text{group}$, $*P < 0.05$.

miR-21 in mouse hearts, which correlated with a significant up-regulation in the PTEN/Akt signaling pathway.²⁴ Extensive investigation of PTEN was conducted in our model, at multiple time points up to 6 h after exposure to isoflurane. However, all time points failed to show a significant change in PTEN expression after isoflurane exposure or miR-21 knockdown. Specifically, 30 min of isoflurane exposure failed to decrease PTEN mRNA or protein levels compared with control values. miR-21 knockdown also failed to influence both endogenous and isoflurane-exposed levels of PTEN compared with control values (fig. 2B).

To ensure that the results seen *in vitro* correlated to *in vivo* conditions, we tested whether the miR-21 knockout mouse could be successfully preconditioned with isoflurane. miR-21 knockout mice are created using a targeting vector, which replaces the 93-bp precursor to miR-21 with a pGK-gb2 *loxP/FRT*-flanked neomycin resistance cassette.³⁴ These mice are healthy and viable and are capable of cardiac remodeling in response to stress.³⁵ However, when exposed to isoflurane, these mice displayed no decrease in infarct size after I/R injury when compared with non-isoflurane-exposed controls. Both the *in vivo* and *in vitro* data provide direct evidence that miR-21 is playing a critical role in the cardioprotection against I/R injury mediated by isoflurane.

One limitation of this study is the use of neonatal rat cardiomyocytes and hydrogen peroxide-induced stress to dissect the important roles of miR-21 in isoflurane-induced protection of cardiomyocytes. This *in vitro* model may not be able to completely recapitulate the *in vivo* environments. However, the results found in neonatal rat cardiomyocytes parallel our work in the miR-21 knockout mouse, as well as previous work in our laboratory,

studying anesthetic preconditioning in whole animal models, isolated heart preparations, and human-induced pluripotent stem cell-derived cardiomyocytes.^{36–38} In addition, neonatal rat cardiomyocytes allow for a broad range of studies and an in-depth view of microRNA signaling pathways in a cardiomyocyte-specific model. When compared with isolated adult rat cardiomyocytes, neonatal myocytes have been shown to maintain a phenotype and contractile profile comparable with *in situ* hearts during I/R injury.³⁹

The use of hydrogen peroxide to mediate reactive oxygen species-induced injury has also been used in previous studies to investigate the role of microRNAs in ischemic preconditioning and in anesthetic preconditioning studies done previously in our laboratory.^{16,23,40} Reactive oxygen species-induced stress plays an important role not only in I/R injury but also in myocardial infarction, heart failure, and hypertrophy.^{41–44} Understanding the changes in expression levels of genes after reactive oxygen species-induced injury can provide important insight into the pathogenesis of these diseases.

In conclusion, the results obtained in this study reveal for the first time that isoflurane directly affects microRNA expression profiles in cardiomyocytes and that miR-21 is acting to protect cardiomyocytes after isoflurane exposure, most likely through its actions on PDCD4. This shows that while isoflurane-induced cardioprotection shares many pathways with ischemic, hypoxic, and heat shock preconditioning, the mechanisms are not identical. Investigating unique, isoflurane-mediated cardioprotective pathways may lead to mechanisms by which infarct size after I/R injury may be decreased, leading to improved patient outcomes after cardiac events.

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Competing Interests

The authors declare no competing interests.

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References

- Najjar SS, Rao SV, Melloni C, Raman SV, Povsic TJ, Melton L, Barsness GW, Prather K, Heitner JF, Kilari R, Gruber L, Hasselblad V, Greenbaum AB, Patel M, Kim RJ, Talan M, Ferrucci L, Longo DL, Lakatta EG, Harrington RA; REVEAL Investigators: Intravenous erythropoietin in patients with ST-segment elevation myocardial infarction: REVEAL: A randomized controlled trial. *JAMA* 2011; 305:1863–72
- Lønborg J, Vejstrup N, Kelbæk H, Holmvang L, Jørgensen E, Helqvist S, Saunamäki K, Ahtarvski KA, Bøtker HE, Kim WY, Clemmensen P, Engstrøm T: Final infarct size measured by cardiovascular magnetic resonance in patients with ST elevation myocardial infarction predicts long-term clinical outcome: An observational study. *Eur Heart J Cardiovasc Imaging* 2013; 14:387–95
- Amour J, Brzezinska AK, Weihrauch D, Billstrom AR, Zielonka J, Krolkowski JG, Bienengraeber MW, Warltier DC, Pratt PF Jr, Kersten JR: Role of heat shock protein 90 and endothelial nitric oxide synthase during early anesthetic and ischemic preconditioning. *ANESTHESIOLOGY* 2009; 110:317–25
- Sedlic F, Sepac A, Pravdic D, Camara AK, Bienengraeber M, Brzezinska AK, Wakatsuki T, Bosnjak ZJ: Mitochondrial depolarization underlies delay in permeability transition by preconditioning with isoflurane: Roles of ROS and Ca²⁺. *Am J Physiol Cell Physiol* 2010; 299:C506–15
- Stadnicka A, Marinovic J, Bienengraeber M, Bosnjak ZJ: Impact of *in vivo* preconditioning by isoflurane on adenosine triphosphate-sensitive potassium channels in the rat heart: Lasting modulation of nucleotide sensitivity during early memory period. *ANESTHESIOLOGY* 2006; 104:503–10
- Ljubkovic M, Mio Y, Marinovic J, Stadnicka A, Warltier DC, Bosnjak ZJ, Bienengraeber M: Isoflurane preconditioning uncouples mitochondria and protects against hypoxia-reoxygenation. *Am J Physiol Cell Physiol* 2007; 292:C1583–90
- Kersten JR, Schmeling TJ, Pagel PS, Gross GJ, Warltier DC: Isoflurane mimics ischemic preconditioning *via* activation of K(ATP) channels: Reduction of myocardial infarct size with an acute memory phase. *ANESTHESIOLOGY* 1997; 87:361–70
- Leucker TM, Bienengraeber M, Muravyeva M, Baotic I, Weihrauch D, Brzezinska AK, Warltier DC, Kersten JR, Pratt PF Jr: Endothelial-cardiomyocyte crosstalk enhances pharmacological cardioprotection. *J Mol Cell Cardiol* 2011; 51:803–11
- Dong S, Cheng Y, Yang J, Li J, Liu X, Wang X, Wang D, Krall TJ, Delphin ES, Zhang C: MicroRNA expression signature and the role of microRNA-21 in the early phase of acute myocardial infarction. *J Biol Chem* 2009; 284:29514–25
- Rane S, He M, Sayed D, Vashistha H, Malhotra A, Sadoshima J, Vatner DE, Vatner SF, Abdellatif M: Downregulation of miR-199a derepresses hypoxia-inducible factor-1alpha and Sirtuin 1 and recapitulates hypoxia preconditioning in cardiac myocytes. *Circ Res* 2009; 104:879–86
- Lishmanov IuB, Maslov LN, Khaliulin IG, Zhang Y, Pei J: [Role of heat shock proteins, aldose reductase, Bcl-2 protein and microRNA in the mechanism of delayed preconditioning of heart]. *Russ Fiziol Zh Im I M Sechenova* 2010; 96:472–88
- Yin C, Salloum FN, Kukreja RC: A novel role of microRNA in late preconditioning: Upregulation of endothelial nitric oxide synthase and heat shock protein 70. *Circ Res* 2009; 104:572–5
- Ishikawa M, Tanaka S, Arai M, Genda Y, Sakamoto A: Differences in microRNA changes of healthy rat liver between sevoflurane and propofol anesthesia. *ANESTHESIOLOGY* 2012; 117:1245–52
- Cao L, Feng C, Li L, Zuo Z: Contribution of microRNA-203 to the isoflurane preconditioning-induced neuroprotection. *Brain Res Bull* 2012; 88:525–8
- Jia P, Teng J, Zou J, Fang Y, Zhang X, Bosnjak ZJ, Liang M, Ding X: miR-21 contributes to xenon-conferred amelioration of renal ischemia-reperfusion injury in mice. *ANESTHESIOLOGY* 2013; 119:621–30
- Jamnicki-Abegg M, Weihrauch D, Pagel PS, Kersten JR, Bosnjak ZJ, Warltier DC, Bienengraeber MW: Isoflurane inhibits cardiac myocyte apoptosis during oxidative and inflammatory stress by activating Akt and enhancing Bcl-2 expression. *ANESTHESIOLOGY* 2005; 103:1006–14
- Hirata N, Shim YH, Pravdic D, Lohr NL, Pratt PF Jr, Weihrauch D, Kersten JR, Warltier DC, Bosnjak ZJ, Bienengraeber M: Isoflurane differentially modulates mitochondrial reactive oxygen species production *via* forward *versus* reverse electron transport flow: Implications for preconditioning. *ANESTHESIOLOGY* 2011; 115:531–40
- Bienengraeber M, Pellitteri-Hahn M, Hirata N, Baye TM, Bosnjak ZJ, Olivier M: Quantitative characterization of changes in the cardiac mitochondrial proteome during anesthetic preconditioning and ischemia. *Physiol Genomics* 2013; 45:163–70
- Funari VA, Winkler M, Brown J, Dimitrijevic SD, Ljubimov AV, Saghizadeh M: Differentially expressed wound healing-related microRNAs in the human diabetic cornea. *PLoS One* 2013; 8:e84425
- Ge ZD, Ionova IA, Vladic N, Pravdic D, Hirata N, Vásquez-Vivar J, Pratt PF Jr, Warltier DC, Pieper GM, Kersten JR: Cardiac-specific overexpression of GTP cyclohydrolase 1 restores ischemic preconditioning during hyperglycaemia. *Cardiovasc Res* 2011; 91:340–9
- Ge ZD, van der Hoeven D, Maas JE, Wan TC, Auchampach JA: A(3) adenosine receptor activation during reperfusion reduces infarct size through actions on bone marrow-derived cells. *J Mol Cell Cardiol* 2010; 49:280–6
- Green GH, Diggle PJ: On the operational characteristics of the Benjamini and Hochberg false discovery rate procedure. *Stat Appl Genet Mol Biol* 2007; 6:Article27
- Cheng Y, Liu X, Zhang S, Lin Y, Yang J, Zhang C: MicroRNA-21 protects against the H₂O₂-induced injury on cardiac myocytes *via* its target gene PDCD4. *J Mol Cell Cardiol* 2009; 47:5–14
- Tu Y, Wan L, Fan Y, Wang K, Bu L, Huang T, Cheng Z, Shen B: Ischemic postconditioning-mediated miRNA-21 protects

- against cardiac ischemia/reperfusion injury *via* PTEN/Akt pathway. *PLoS One* 2013; 8:e75872
25. Roy S, Khanna S, Hussain SR, Biswas S, Azad A, Rink C, Gnyawali S, Shilo S, Nuovo GJ, Sen CK: MicroRNA expression in response to murine myocardial infarction: miR-21 regulates fibroblast metalloprotease-2 *via* phosphatase and tensin homologue. *Cardiovasc Res* 2009; 82:21–9
 26. Hinkel R, Penzkofer D, Zühlke S, Fischer A, Husada W, Xu QF, Baloch E, van Rooij E, Zeiher AM, Kupatt C, Dimmeler S: Inhibition of microRNA-92a protects against ischemia/reperfusion injury in a large-animal model. *Circulation* 2013; 128:1066–75
 27. Mutharasan RK, Nagpal V, Ichikawa Y, Ardehali H: microRNA-210 is upregulated in hypoxic cardiomyocytes through Akt- and p53-dependent pathways and exerts cytoprotective effects. *Am J Physiol Heart Circ Physiol* 2011; 301:H1519–30
 28. Chen M, Ma G, Yue Y, Wei Y, Li Q, Tong Z, Zhang L, Miao G, Zhang J: Downregulation of the miR-30 family microRNAs contributes to endoplasmic reticulum stress in cardiac muscle and vascular smooth muscle cells. *Int J Cardiol* 2014; 173:65–73
 29. Haider KH, Idris NM, Kim HW, Ahmed RP, Shujia J, Ashraf M: MicroRNA-21 is a key determinant in IL-11/Stat3 antiapoptotic signalling pathway in preconditioning of skeletal myoblasts. *Cardiovasc Res* 2010; 88:168–78
 30. Kukreja RC, Yin C, Salloum FN: MicroRNAs: New players in cardiac injury and protection. *Mol Pharmacol* 2011; 80:558–64
 31. Thum T, Gross C, Fiedler J, Fischer T, Kissler S, Bussen M, Galuppo P, Just S, Rottbauer W, Frantz S, Castoldi M, Soutschek J, Kotliarsky V, Rosenwald A, Basson MA, Licht JD, Pena JT, Rouhanifard SH, Muckenthaler MU, Tuschl T, Martin GR, Bauersachs J, Engelhardt S: MicroRNA-21 contributes to myocardial disease by stimulating MAP kinase signalling in fibroblasts. *Nature* 2008; 456:980–4
 32. Vikhreva PN, Shepelev MV, Korobko EV, Korobko IV: Pdcd4 tumor suppressor: Properties, functions, and possible applications in oncology. *Mol Gen Microbiol Virusol* 2010; 2:3–11
 33. Xu X, Kriegel AJ, Liu Y, Usa K, Mladinov D, Liu H, Fang Y, Ding X, Liang M: Delayed ischemic preconditioning contributes to renal protection by upregulation of miR-21. *Kidney Int* 2012; 82:1167–75
 34. Ma X, Kumar M, Choudhury SN, Becker Buscaglia LE, Barker JR, Kanakamedala K, Liu MF, Li Y: Loss of the miR-21 allele elevates the expression of its target genes and reduces tumorigenesis. *Proc Natl Acad Sci U S A* 2011; 108:10144–9
 35. Patrick DM, Montgomery RL, Qi X, Obad S, Kauppinen S, Hill JA, van Rooij E, Olson EN: Stress-dependent cardiac remodeling occurs in the absence of microRNA-21 in mice. *J Clin Invest* 2010; 120:3912–6
 36. Tampo A, Hogan CS, Sedlic F, Bosnjak ZJ, Kwok WM: Accelerated inactivation of cardiac L-type calcium channels triggered by anaesthetic-induced preconditioning. *Br J Pharmacol* 2009; 156:432–43
 37. An J, Rhodes SS, Jiang MT, Bosnjak ZJ, Tian M, Stowe DF: Anesthetic preconditioning enhances Ca²⁺ handling and mechanical and metabolic function elicited by Na⁺-Ca²⁺ exchange inhibition in isolated hearts. *ANESTHESIOLOGY* 2006; 105:541–9
 38. Canfield SG, Sepac A, Sedlic F, Muravyeva MY, Bai X, Bosnjak ZJ: Marked hyperglycemia attenuates anesthetic preconditioning in human-induced pluripotent stem cell-derived cardiomyocytes. *ANESTHESIOLOGY* 2012; 117:735–44
 39. Yamashita N, Nishida M, Hoshida S, Kuzuya T, Hori M, Taniguchi N, Kamada T, Tada M: Induction of manganese superoxide dismutase in rat cardiac myocytes increases tolerance to hypoxia 24 hours after preconditioning. *J Clin Invest* 1994; 94:2193–9
 40. Ogbi M, Johnson JA: Protein kinase Cepsilon interacts with cytochrome c oxidase subunit IV and enhances cytochrome c oxidase activity in neonatal cardiac myocyte preconditioning. *Biochem J* 2006; 393:191–9
 41. Lefer DJ, Granger DN: Oxidative stress and cardiac disease. *Am J Med* 2000; 109:315–23
 42. Seddon M, Looi YH, Shah AM: Oxidative stress and redox signalling in cardiac hypertrophy and heart failure. *Heart* 2007; 93:903–7
 43. Takano H, Zou Y, Hasegawa H, Akazawa H, Nagai T, Komuro I: Oxidative stress-induced signal transduction pathways in cardiac myocytes: Involvement of ROS in heart diseases. *Antioxid Redox Signal* 2003; 5:789–94
 44. Berg K, Jynge P, Bjerve K, Skarra S, Basu S, Wiseth R: Oxidative stress and inflammatory response during and following coronary interventions for acute myocardial infarction. *Free Radic Res* 2005; 39:629–36