

Propofol and Remifentanyl Differentially Modulate Frontal Electroencephalographic Activity

David T. J. Liley, M.B., Ch.B., Ph.D.,*

Nicholas C. Sinclair, B.Sc. (Medical Biophysics), B.E. (Electrical and Electronic) (Hons),†

Tarmo Lipping, Dr.Tech.,‡ Bjorn Heyse, M.D.,§ Hugo E. M. Vereecke, M.D., Ph.D.,||

Michel M. R. F. Struys, M.D., Ph.D.#

ABSTRACT

Background: The purpose of this study was to evaluate a new, physiologically inspired method for the analysis of the electroencephalogram during propofol–remifentanyl anesthesia. Based on fixed-order autoregressive moving-average modeling, this method was hypothesized to be capable of dissociating the effects that hypnotic and analgesic agents have on brain electrical activity.

Methods: Raw electroencephalographic waves from a previously published study were reanalyzed. In this study, 45 American Society of Anesthesiologists status I patients were randomly allocated to one of three groups according to a specific target effect-site remifentanyl concentration (0, 2, and 4 ng/ml). All patients received stepwise-increased targeted effect-site concentrations of propofol (C_{EPROP}). At

each step change in target C_{EPROP} , the Observer's Assessment of Alertness/Sedation score was evaluated. Raw electroencephalograph was continuously acquired from frontal electrodes. Electroencephalography traces were analyzed using a fixed-order autoregressive moving average model to give derived measures of Cortical State and Cortical Input. Response surfaces were visualized and modeled using Hierarchical Linear Modeling.

Results: Cortical State (a measure of cortical responsiveness) and Cortical Input (a measure of the magnitude of cortical input) were shown to respond differently to C_{EPROP} and effect-site remifentanyl concentration. Cortical Input decreased significantly with increasing effect-site remifentanyl concentration, whereas Cortical State remained unchanged with increasing effect-site remifentanyl concentration but decreased with increasing C_{EPROP} .

Conclusion: Because Cortical State responds principally to variations in C_{EPROP} , it is a potential measure of hypnosis, whereas the dependence of Cortical Input on effect-site remifentanyl concentration suggests that it may be useful as a measure of analgesic efficacy and the nociceptive–antinociceptive balance.

* Associate Professor, Brain Sciences Institute, Swinburne University of Technology, Hawthorn, Victoria, Australia, and Chief Technology Officer, Cortical Dynamics Ltd., North Perth, Western Australia, Australia. † Biomedical Research Engineer, Cortical Dynamics Ltd. ‡ Professor, Department of Information Technology, Tampere University of Technology, Pori, Finland. § Resident in Anesthesia, || Staff Anesthesiologist, Department of Anesthesia, Ghent University, Ghent, Belgium. # Professor and Chair, Department of Anesthesiology, University Medical Center Groningen, University of Groningen, Groningen, The Netherlands, and Professor, Department of Anesthesia, Ghent University, Ghent, Belgium.

Received from the Brain Dynamics Research Group, Brain Sciences Institute, Swinburne University of Technology, Hawthorn, Victoria, Australia; Department of Anesthesia, Ghent University Hospital, Ghent, Belgium; University Medical Center Groningen, University of Groningen, Groningen, The Netherlands; and Department of Information Technology, Tampere University of Technology, Pori, Finland. Submitted for publication December 10, 2009. Accepted for publication April 5, 2010. Supported by institutional sources of Cortical Dynamics Ltd., North Perth, Western Australia, Australia, and the Brain Sciences Institute, Swinburne University of Technology, Hawthorn, Victoria, Australia, and by grant No. 7225 from Estonian Science Foundation, Tallinn, Estonia. Dr. Liley holds an unvalued equity stake in Cortical Dynamics Ltd.

Address correspondence to Dr. Liley: Brain Sciences Institute, Swinburne University of Technology, P.O. Box 218, Hawthorn, Victoria 3122, Australia. dliley@swin.edu.au. Information on purchasing reprints may be found at www.anesthesiology.org or on the masthead page at the beginning of this issue. ANESTHESIOLOGY's articles are made freely accessible to all readers, for personal use only, 6 months from the cover date of the issue.

What We Already Know about This Topic

- ❖ Assessing depth of anesthesia by spontaneous electroencephalographic activity is limited
- ❖ Neurophysiology-based processed electroencephalographic monitoring in response to an arbitrary stimulus might improve performance

What This Article Tells Us That Is New

- ❖ In 45 patients undergoing surgery, fixed-order time-series modeling of electroencephalographic activity differentiated effects of the hypnotic propofol from those of the analgesic remifentanyl
- ❖ This approach might enable independent monitoring of hypnotic and analgesic drug actions

◆ This article is accompanied by an Editorial View. Please see: Sleigh J: Disentangling Hypnos from his poppies. ANESTHESIOLOGY 2010; 113:271–2.

TO date depth of anesthesia monitoring has relied on a range of heuristic measures to objectively assess depth of anesthesia. The most successful existing methods are arguably those derived from the analysis of spontaneous or time-locked electroencephalographic activity.¹ In particular, the Bispectral Index[®] (BIS[®]; Aspect Medical Systems, Norwood, MA) has achieved a substantial level of routine clinical use because of its reported efficacy in defining optimal levels of hypnosis such that intraoperative awareness is minimized.² Although reportedly enabling anesthesia to be more optimally administered, it does so in the context of a number of well-documented limitations: not all hypnotic agents are reliably detected or monitored (nitrous oxide^{3–6} and the short-acting synthetic opioids^{7–9} being quintessential examples), and the index admits of no clear physiologic interpretation because it has been constructed to act as a quantitative surrogate for an ostensibly subjective state. Although a range of other processed electroencephalographic monitoring approaches have been developed in an attempt to circumvent such limitations or to improve on the predictive ability of the BIS in quantifying anesthesia, none has shown any clear advantage.¹ Such approaches include those based on spontaneous electroencephalographic activity, such as the Narcotrend index (Narcotrend[®]; Schiller AG, Baar, Switzerland) and the State Entropy and Response Entropy indices (M-entropy[®] module; GE Healthcare Finland Oy, Helsinki, Finland), and those based on analyzing the morphology of the middle latency auditory-evoked potential such as the A-Line ARX index (AAI[®]; formerly Danmeter A/S, Odense, Denmark, no longer trading). These indices, and a range of other empirical measures that are based on assumed changes in the complexity of the electroencephalogram signal with increasing depth of anesthesia, are all heuristic constructs. Because these measures are not derived from an understanding of the mechanisms responsible for the genesis of dynamical activity in the electroencephalogram, any anesthetic-induced electroencephalographic changes detected using such measures must necessarily be of suboptimal sensitivity and specificity and consequently will be of limited physiologic relevance. Therefore, the development of physiologically more specifically motivated processed electroencephalographic approaches would be expected to result in improved performance compared with existing methods. We outline one such approach and show that it is able to differentiate the effects of propofol and remifentanyl on frontally recorded electroencephalograms. This has the potential to pave the way for monitoring the hypnotic effect of propofol independent of the analgesic effect of remifentanyl, a feature absent in all existing processed clinical electroencephalogram-based monitoring approaches.¹⁰

The approach we will consider is based on a detailed theory of mammalian cortical electrorhythmogenesis.^{11–13} In brief, it speculates that the rhythmic activity observed in the electroencephalogram arises from the reverberant activity of spatially distributed networks of excitatory and inhibitory cortical neurons. This theory is able to account for a number

of electroencephalographic phenomena that are of relevance to better understand and monitor anesthesia—the benzodiazepine-induced “ β buzz,”¹³ the proconvulsant effects of the volatile general anesthetic agent enflurane,¹⁴ and the biphasic surge in total electroencephalographic power that typically accompanies anesthetic induction and emergence.^{11,15} Although the full theory is mathematically elaborate, it does suggest, to first approximation, that resting electroencephalography may be regarded as a filtered pseudorandom linear process. In particular, it posits that the electroencephalogram can be regarded as arising from cortex linearly filtering subcortical (thalamic) input. The direct empirical consequence is that the electroencephalogram can be modeled as a fixed-order autoregressive moving average (ARMA) process.¹³ In this manner, the estimated ARMA coefficients characterize the properties of the “cortical” filter, whereas the estimated amplitude of the white noise driving corresponds to the assumed magnitude of the subcortical (thalamic) input. In subsequent analyses, a single scalar measure of the filter characteristics is referred to as Cortical State (CS), whereas the amplitude of the innovating noise is defined as the Cortical Input (CI). From a functional point of view, CS can be understood as characterizing the response of cortex to an arbitrary stimulus or input. Because of this increase in physiologic specificity, it was speculated that this fixed-order ARMA analysis would be able to detect the effects of agents not readily detected using other methods. Initial application of this method to sevoflurane in the presence of varying levels of adjuvant nitrous oxide¹⁶ revealed that nitrous oxide, consistent with its antinociceptive properties, reduced CI but left CS unaffected.

To further investigate the relevance of fixed-order ARMA modeling for monitoring depth of anesthesia, we sought to determine whether the ultra-short-acting synthetic opioid remifentanyl, like nitrous oxide, exerted its principle cortical effect by reducing CI. Even in the absence of specific noxious stimuli, we would expect there to be a “background” of subcortical input arising from ambient sensory stimulation that will be ablated by opioid action. In the study reported here, it is found that during propofol–remifentanyl anesthesia CS responds principally to variations in propofol effect-site concentration ($C_{E\text{PROP}}$) and is therefore a likely measure of hypnotic state, whereas CI responds dominantly to changes in remifentanyl effect-site concentrations ($C_{E\text{REM}}$) and therefore might represent a measure of analgesic state (nociceptive–antinociceptive balance).

Materials and Methods

Patient Recruitment and Study Design

Raw electroencephalographic waves from a previously published study were reanalyzed.¹⁷ The original study was approved by the institutional ethics committee (Ghent University Hospital, Ghent, Belgium) and written informed consent was obtained from 45 patients of American Society of Anesthesiologists status I, aged 18–60 yr, and scheduled

Table 1. Responsiveness Scores of the Modified Observer's Assessment of Alertness/Sedation Scale

Score	Responsiveness
5	Responds readily to name spoken in normal tone
4	Lethargic response to name spoken in normal tone
3	Responds only after name is called loudly or repeatedly
2	Responds only after mild prodding or shaking
1	Responds only after painful trapezius squeeze
0	No response after painful trapezius squeeze

to undergo orthopedic surgery. Exclusion criteria were as follows: weight less than 70% or more than 130%, of ideal body weight (per table of Desirable Weights, Metropolitan Life Insurance, 1983), neurologic disorder, recent use of psychoactive medication or alcohol. Per study by Ferenets *et al.*¹⁷, patients were randomly allocated to one of three groups: remi0, in which no remifentanyl was given, and groups remi2 and remi4, in which effect compartment-controlled infusions of remifentanyl were targeted at 2 and 4 ng/ml, respectively. Four minutes after the start of the remifentanyl infusion, a "stair-case" computer-controlled infusion of propofol was commenced and initially targeted to an effect-site concentration of 0.75 $\mu\text{g/ml}$, which was subsequently increased every 4 min in steps of 0.25–0.3 $\mu\text{g/ml}$ until the loss of response to all clinically relevant measurements of alertness and sedation was observed. Ten seconds before each increase in target propofol concentration, clinical assessment of the level of alertness and sedation was made using the Modified Observer's Assessment of Alertness/Sedation (OAA/S) (table 1). This scale is assessed by applying progressively more intense stimulation ranging from a moderate speaking voice to physical shaking or moderate noxious stimulus (trapezius squeeze) until a response is observed. Patients were considered responsive to vocal stimulus at OAA/S levels 5, 4, or 3 and scored as unresponsive to vocal stimulus at OAA/S levels 2, 1, or 0.

Propofol and remifentanyl were administered *via* a computer-assisted continuous infusion device to a target effect-site concentration (RUGLOOP II; Demed, Temse, Belgium) using a three-compartment model enlarged with an effect-site compartment. For propofol, the pharmacokinetic-dynamic model previously published by Schnider *et al.*^{18,19} was used. For remifentanyl, the corresponding model used was that previously published by Minto *et al.*^{20,21} Predicted effect-site propofol concentration ($C_{E\text{PROP}}$) was computed to yield a time to peak effect of 1.6 min after bolus injection (also as published by Minto *et al.*²²) and pharmacokinetically confirmed in a clinical population by Struys *et al.*²³ For remifentanyl, an age-dependent k_{e0} (effect-site elimination rate constant) value of $0.595 - 0.007 \times (\text{age} - 40) \text{ min}^{-1}$ was applied as described by Minto *et al.*^{20,21} Propofol and remifentanyl infusions were administered using a Fresenius Modular DPS Infusion Pump connected to a Fresenius Base (Fresenius Vial Infusion Systems, Bresin, France). RUG-

LOOP II controls the pump at infusion rates between 0 and 1,200 ml/h *via* an RS232 interface. This infusion technique enables titration to a steady state defined as the equilibration between the calculated plasma and effect-site concentrations of the drug. To minimize the prediction error of the steady-state drug concentration at the time of clinical observation, an equilibration time of 4 min was allowed after every change of drug concentration before response to stimuli was tested. Remifentanyl and propofol were infused *via* a large left forearm vein. Each patient received approximately 200 ml of crystalloid fluid during the study period. No fluid load was given before induction. None of the patients received any preanesthetic medication, and no other drugs were given. During the study period, all patients maintained spontaneous ventilation *via* a facemask delivering 6 l/min O_2 .

Data Acquisition

Heart rate, noninvasive blood pressure, oxygen saturation measured by pulse oximetry, and capnography were monitored continuously using an S/5 Anesthesia Monitor (GE Healthcare, Helsinki, Finland) and recorded electronically using RUGLOOP II data management software. The raw electroencephalogram was recorded with the M-Entropy module of the S/5 Anesthesia Monitor and was sampled at 400 Hz, and written to disk, by using the S5-collect software. The standard entropy sensor was used with a slightly modified positioning: the two recording electrodes of the sensor were located bilaterally on the forehead approximately 5 cm above the eyebrows and 4 cm from the midline in either direction. The ground electrode was located between the two recording electrodes. This alternative montage was chosen to minimize electromyographic activity that normally contributes to the calculation of the State Entropy and Response Entropy measures, but for our purposes it is considered artifactual. This bifrontal montage gives rise to approximately the same mean electroencephalographic amplitudes as a unilaterally placed sensor.

Offline Signal Processing and Artifact Rejection

Both sampled raw and resampled raw electroencephalograms were used in subsequent analyses. Time series models (see Eqs. 1 and 2 below) were fitted to resampled (from 400 to 80 Hz) raw electroencephalogram as per Liley *et al.*¹⁶ As discussed therein, this was performed to avoid spurious fitting to 50-Hz spectral peaks or any low-pass filter band edges. Resampling was performed in MATLAB (Mathworks, Natick, MA) using a process of antialiasing filtering and downsampling. The antialias filter used was a finite impulse response filter with sharp cutoff at 40 Hz with the transition band made sufficiently sharp to minimize any aliasing.

Both the original and resampled electroencephalogram time series were segmented into 2-s 50% overlapping epochs and aligned with the respective measurements of estimated steady-state propofol concentration and OAA/S. For the original electroencephalogram time series, the electromyograph (defined as the total power between 70 and 110 Hz

excluding a notch at 98–102 Hz due to 50 Hz electric power harmonic at 100 Hz) was calculated. The root mean square (RMS) amplitude was calculated from the resampled electroencephalogram time series. Subsequently, an automated artifact rejection method was used to classify all epochs based on the original and resampled electroencephalogram time series. Epochs were excluded from further analysis if any of the following occurred: total electromyographic power greater than approximately $400 \mu\text{V}^2$ or less than approximately $0.004 \mu\text{V}^2$, RMS amplitude less than $5 \mu\text{V}$ or greater than $150 \mu\text{V}$, amplitude distributions were not normal (based on Lilliefors²⁴ test at $P = 0.01$) or epochs to either side, of the epoch in question, were rejected. For each event (targeted propofol concentration or OAA/S observation), average CS, CI, RMS, and electromyogram were calculated for the 30 s preceding the event. If more than 50% of the corresponding epochs were corrupted then this event was not subsequently used.

CS and CI were calculated using the resampled electroencephalogram as described previously by Liley *et al.*¹⁶ We now briefly summarize the salient details of this method. Based on significant experimental evidence that electroencephalogram recorded in the presence and absence of anesthesia can be modeled as a random linear process,^{13,25–30} a linearized version of a fully nonlinear theory of electrorhythmogenesis was used to motivate fixed-order (ARMA) time series modeling. Specifically, the sampled electroencephalogram signal $s[n]$ was modeled using an (8,5) ARMA model

$$s[n] = -\sum_{k=1}^{k=8} a_k s[n-k] + \sum_{k=0}^{k=5} b_k u[n-k] \quad (1)$$

or

$$A(z)S(z) = B(z)U(z) \quad (2)$$

where $u[n]$ represents a stationary sequence of uncorrelated random variables of variance σ_u^2 , a_k and b_k are the respective estimated autoregressive and moving average parameters. $S[z]$ and $U[z]$ are the respective Z-transforms of $s[n]$ and $u[n]$ (*i.e.*, $S[z] = Z\{s[n]\}$, $U[z] = Z\{u[n]\}$), $A(z) = 1 + a_1 z^{-1} + \dots + a_8 z^{-8}$ and $B(z) = 1 + b_1 z^{-1} + \dots + a_5 z^{-5}$. $\frac{B(z)}{A(z)}$ represents the electrocortical filter and describes how subcortical input (assumed to be so complicated as to be indistinguishable from an uncorrelated random process) is filtered to give rise to the surface recordable electroencephalogram. The theoretically derived autoregressive and moving average orders of 8 and 5 accord well with empirical determinations of optimal autoregressive (range, 3–14) and moving average (range, 2–5) orders obtained from resting awake eyes closed electroencephalogram using a range of information theoretic criteria.^{27,30} The poles and zeros of the electrocortical filter are the respective solutions to $A(z) = 0$ and $B(z) = 0$. The poles and zeros of the estimated electrocortical filter are predicted to be of physiologic significance. For example, weakly damped poles will be seen as dominant oscillatory processes in the electroencephalogram (for example, the 8–13 Hz α

rhythm). Therefore, tracking how the poles and zeros of the electrocortical filter change would seem to provide the best means of characterizing variations in the state of the electrocortical filter. One easily calculated scalar measure of the state of the electrocortical filter is the mean pole location. Therefore, for each resampled epoch $s[n]$, CS was calculated as the scaled mean pole location a_1 . CI was calculated as the square root of the variance of $Z^{-1} \left\{ \frac{A(z)S(z)}{B(z)} \right\}$ (*i.e.*, the variance of $s[n]$ divided by the power gain of the derived filter). Thus, CI represents the RMS amplitude of the noise innovating the electrocortical filter. The (8,5) ARMA model parameters were robustly determined with well-established methods,³¹ using the ARMASA MATLAB Toolbox.³² In brief, ARMASA removes the mean of the epoch then estimates an invertible and stationary ARMA model using a variant of Durbin methods with optimal intermediate autoregressive order.

Statistical Analysis

Normally distributed data were summarized as mean \pm SD, and skewed data were given as median (range) and counts as number (%). Omnibus tests were performed using analysis of variance or the Kruskal–Wallis test appropriately based on the results of the Levene test for homogeneity of variance. *Post hoc* multiple comparisons were made using Tukey Honestly Significant Difference or the Mann–Whitney U test with Bonferroni correction wherever appropriate. All statistical analyses, except for the hierarchical linear modeling (see Eqs. 3 and 4 below), were performed using SPSS for Windows (version 16; SPSS Inc., Chicago, IL). A value of p less than 0.05 was considered statistically significant.

To assess the ability of CS and CI to indicate the subjects level of sedation, both prediction probability (P_k) and Spearman ρ were calculated. P_k is an asymmetric measure of ordinal association and is a rescaled version of the more familiar statistics Somers' d_{XY} and Kim's d_{YX} .^{33,34} In particular, $P_k \equiv \frac{d_{XY} + 1}{2} = \frac{d_{YX} + 1}{2}$, where X is the dependent variable (OAA/S level) and Y is the independent regressor variable (CI or CS). We chose to calculate P_k using Somers' D statistic in SPSS, which also provides an estimate of the Goodman and Kruskal approximate SE,³⁴ $\sigma_{\text{SOMERS } D}$. As a consequence, we define the SE of P_k , σ_{PK} , to be $\frac{\sigma_{\text{SOMERS } D}}{2}$.

The P_k , and its SE, calculated in this way is reported to be associated with no significant bias compared with the corresponding jackknife estimates calculated using the PKMACRO of Smith *et al.*³⁴ P_k has a value of 1 when the indicator variable (CI or CS) predicts observed anesthetic depth perfectly and a value of 0.5 when the indicator predicts no better than a 50:50 chance. Because it is often reported, we also chose to calculate the Spearman rank correlation coefficient with correction for tied ranks. Although it has the advantage of avoiding distributional assumptions of other correlational measures, it has the disadvantage of lacking an

Table 2. Patient Demographics for No Remifentanyl (Remi0), 2 ng/ml Remifentanyl (Remi2), and 4 ng/ml Remifentanyl (Remi4) Treatment Groups

Group	Age, yr (SD)	Height, cm (SD)	Weight, kg (SD)	M/F
Remi0	36 (10)	171 (10)	70 (8)	6/8
Remi2	33 (5)	168 (13)	67 (15)	5/9
Remi4	39 (8)	172 (9)	71 (16)	7/7

intrinsic meaning, in that its units depend very much on the ordinal scales used. This makes subsequent comparisons with other depth of anesthesia measures difficult. For this reason, P_k is typically preferred.

The relationship between C_{ePROP} and C_{eREMI} and the derived electroencephalographic measures of CI and CS was analyzed using hierarchical linear modeling (also known as multilevel analysis). This multilevel analysis is a more advanced form of simple multivariate linear regression.³⁵ This regression strategy was preferred because (1) we had no *a priori* reason to believe that CI and CS would follow a bivariate sigmoidal E_{max} model and (2) the data were nested, such that each participant will have had CI and CS measured at one target remifentanyl concentration but multiple target propofol concentrations; that is, data are first grouped with respect to C_{eREMI} and with respect to C_{ePROP} . Specifically, the following two-level mixed effects model was posed

$$y = \sum_{n=0}^N \beta_n(R) P^n + \epsilon \quad (3)$$

$$\beta_n(R) = \sum_{m=0}^{M_n} \gamma_{nm} R^m + u_n \quad (4)$$

where y is either CI or CS, P and R are C_{ePROP} and C_{eREMI} , respectively, and ϵ and u_n are error terms (assumed to be normally distributed). Default regressor orders were set to cubic (*i.e.*, $N = 3$, $M_n = 3$) for initial exploratory analyses. Fitting was performed using HLM 6.08 (Scientific Software International, Lincoln, IL). Optimal regressor orders were subsequently determined based on the residual variance, the structural simplicity of the model, the homogeneity of the level 1 residuals of regression, and the collinearity of the level 2 Mahalanobis distance (test of normality/outliers) and chi-square measures. A linear relationship between the Mahalanobis distance and chi-square supports the assumption of normality in the data and ensures that no outliers have biased any of the estimated regression coefficients. All possible residual covariance terms were used for the level 2 modeling.

Results

All changes in hemodynamic and capnography were within clinical limits (data not presented). The demographic data for all patients are given in table 2.

Relationship between Electroencephalographic Measures and Clinical Assessments of Patient State

Approximately 23% of all 2-s electroencephalogram epochs were rejected because of artifact. This resulted in elimination of 9% of all OAA/S measurements as a result of the absence of sufficient artifact-free electroencephalogram (see Materials and Methods for further details). Figure 1 shows box and whisker plots for CS, CI, RMS, and electromyogram activity *versus* OAA/S levels for each remifentanyl treatment group. CS and electromyogram clearly decrease with decreases

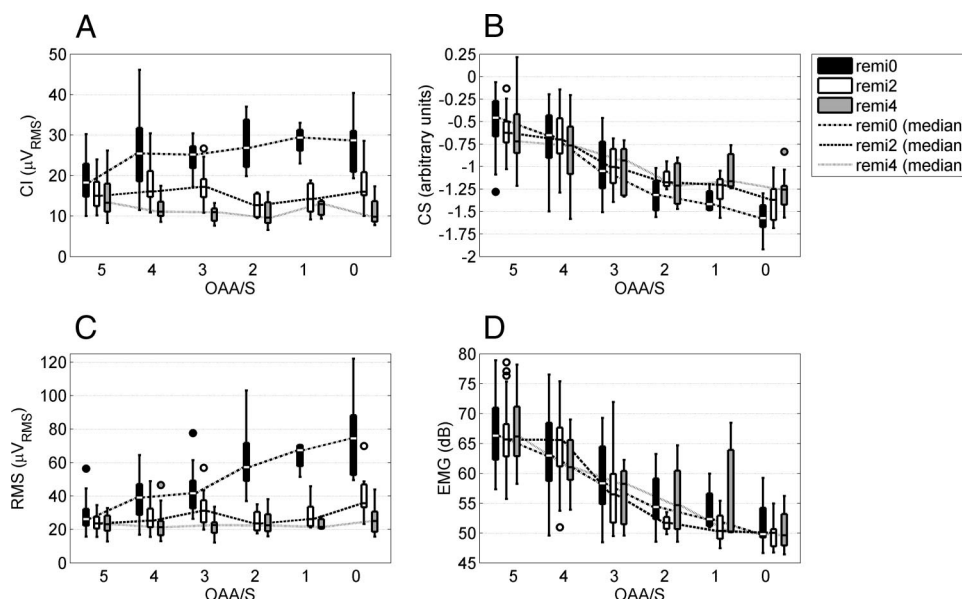


Fig. 1. Box-and-whisker plots for derived electroencephalographic measures as a function of the Observer's Assessment of Alertness/Sedation (OAA/S) level for no remifentanyl (remi0), 2 ng/ml remifentanyl (remi2), and 4 ng/ml remifentanyl (remi4) treatment groups. Boxes represent interquartile ranges, lines enclosed within boxes (and connected lines) median values, whiskers represent the largest (smallest) nonoutlier, and circles represent outliers (defined as values extending further than 1.5 times the interquartile range—equivalent to approximately 3 SDs for normally distributed data). (A) Cortical Input (CI), (B) Cortical State (CS), (C) root mean square (RMS) electroencephalogram amplitude, and (D) electromyogram (EMG).

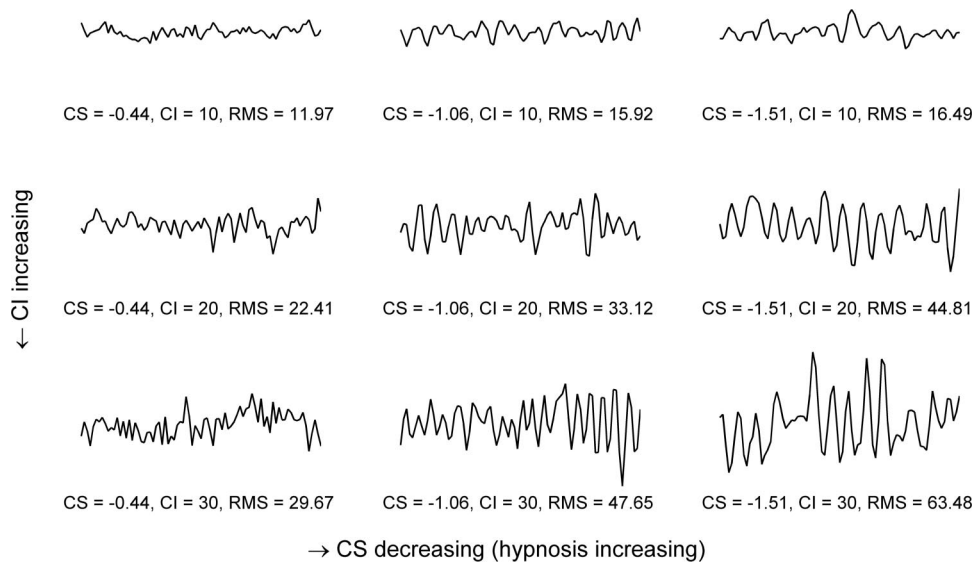


Fig. 2. Examples of synthetic 2-s epochs of electroencephalogram data illustrating the independence of Cortical Input (CI) and root mean square (RMS) electroencephalogram amplitude. Each column of this figure represents a realization of a fixed autoregressive moving average process, estimated from an artifact-free 2-s epoch of electroencephalogram to which a Gaussian white-noise innovation of differing amplitudes was applied. The 2-s electroencephalogram epochs were chosen at random from the recorded electroencephalogram of a patient in the treatment group that received no remifentanyl (remi0) at various levels of estimated hypnosis (Cortical State [CS]). Note that for a fixed CS (column-wise), CI and RMS covary, whereas for varying CS (row-wise), CI remains fixed while RMS varies.

ing levels of consciousness, whereas CI and RMS are seen to be largely independent of the clinically assessed patient state. However, as confirmed subsequently by hierarchical linear modeling (see Relationship between Electroencephalo-

graphic Measures and Effect-site Remifentanyl and Propofol Concentrations below), significant differences in these latter measures were observed as a function of predicted effect-site remifentanyl concentration and were increasingly marked at

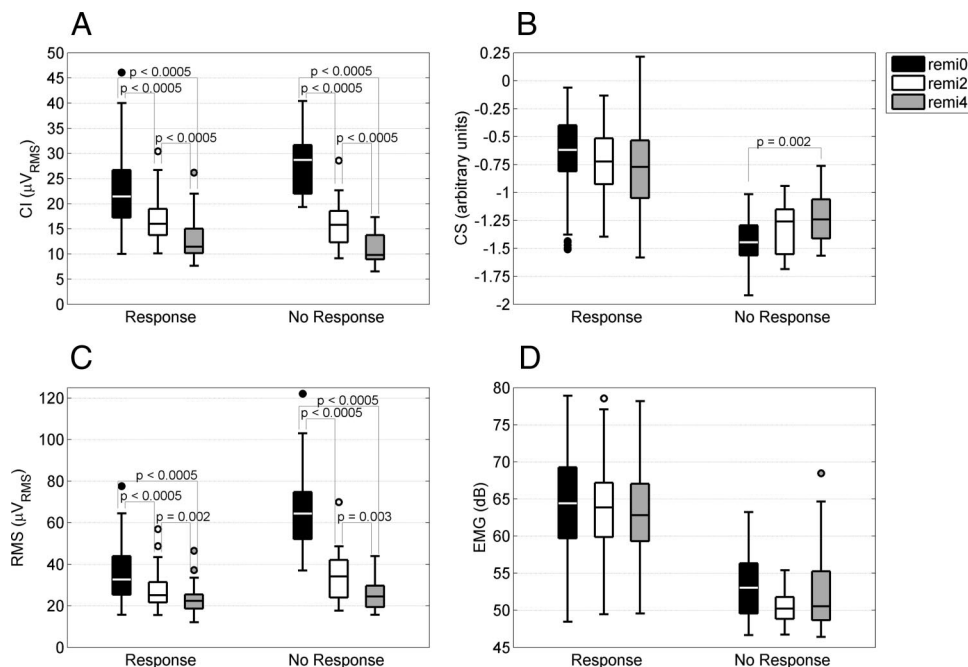


Fig. 3. Box-and-whisker plots for derived electroencephalographic measures for loss of vocal response for no remifentanyl (remi0), 2 ng/ml remifentanyl (remi2), and 4 ng/ml remifentanyl (remi4) treatment groups. Boxes represent interquartile ranges, lines enclosed within boxes (and connected lines) median values, whiskers represent the largest (smallest) nonoutlier, and circles represent outliers (defined as values extending an additional 1.5 times the interquartile range—approximately equivalent to 3 SDs for normally distributed data). (A) Cortical Input (CI), (B) Cortical State (CS), (C) root mean square (RMS) electroencephalogram amplitude, and (D) electromyogram (EMG).

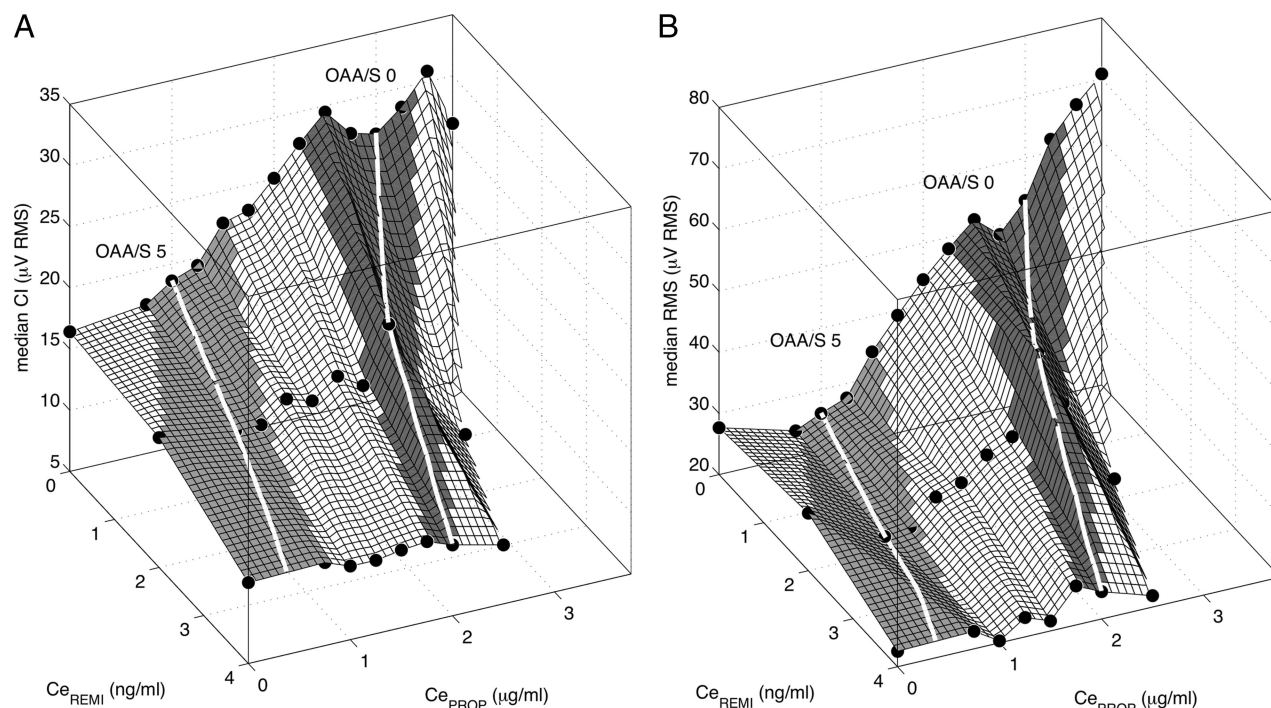


Fig. 4. (A) Median Cortical Input (CI) versus target remifentanyl (C_{e_REMI}) and propofol (C_{e_PROP}) concentrations. Mesh represents the linearly interpolated surface through data (filled circles). Interquartile (25th–75th percentile) ranges of target propofol concentration for a given Observer's Assessment of Alertness/Sedation (OAA/S) level and remifentanyl level are shown as shaded (OAA/S 5 = light gray, OAA/S 0 = dark gray) patches on the interpolated surface. Median values correspond to the respective solid white line. This surface together with the shaded interquartile ranges indicates that for a fixed OAA/S level and target propofol concentration, CI decreases with increasing C_{e_REMI} . (B) Median root mean square (RMS) electroencephalogram amplitude versus C_{e_REMI} and C_{e_PROP} . All other details as for A.

deeper levels of clinically assessed sedation. In particular, at OAA/S level 0 (unresponsive to painful stimulus), CI displayed significant reductions with increasing predicted effect-site remifentanyl concentration. The similarity between the changes in CI and RMS as a function of OAA/S levels is a reflection of the fact that the former measure depends on the latter for its calculation. Nevertheless, as illustrated in figure 2 CI can remain fixed whereas RMS changes depending on variations in CS. Therefore, despite its simpler calculation, RMS cannot be used as a proxy for CI. In contrast, the similarity between CS and the estimated electromyogram cannot be a consequence of the method of their calculation because the latter is calculated only on recorded scalp electrical activity between 70 and 110 Hz, whereas the former is calculated on the range of 0–40 Hz. On this basis, we can reasonably speculate that CS and the estimated electromyogram are related at a deeper physiologic level.

Relationship between Electroencephalographic Measures and Loss of Response to Vocal Stimulus

Because CI and CS are being assessed with respect to their ability to characterize the level of sedation in the presence of remifentanyl, it is important to determine their ability to predict loss of response. On this basis, the above OAA/S data were dichotomously aggregated into either the presence or absence of response to vocal stimulus. Loss of response to a

vocal stimulus corresponds to the transition from OAA/S level 3 (responds only after name is called loudly or repeatedly) to OAA/S level 2 (responds only after mild prodding or shaking). Therefore, OAA/S levels 5–3 are treated as responsive to verbal command, whereas OAA/S levels 2–0 are defined as unresponsive to verbal command. Figure 3 shows the box and whisker plots for this aggregated data. As expected from figure 1, CS and the electromyogram are particularly sensitive to the loss of response to vocal stimulus.

Relationship between Electroencephalographic Measures and Effect-site Remifentanyl and Propofol Concentrations

Figures 4 and 5 show interpolated surface plots of median CI, CS, RMS, and electromyogram as a function of predicted effect-site remifentanyl and propofol concentrations. In general, it is observed that median CI and RMS (fig. 4) vary with propofol and remifentanyl concentration, whereas CS and the electromyogram (fig. 5) principally depend on variations in target propofol levels. The results of the hierarchical linear modeling (tables 3 and 4) confirm that significant reductions in CI occur with increasing effect-site remifentanyl concentration as for fixed propofol levels CI is negatively correlated (γ_{11} is less than 0) with remifentanyl level. It is notable that none of the level 2 random effects for CI are significant, implying that individual level differences were not important contributors to CI

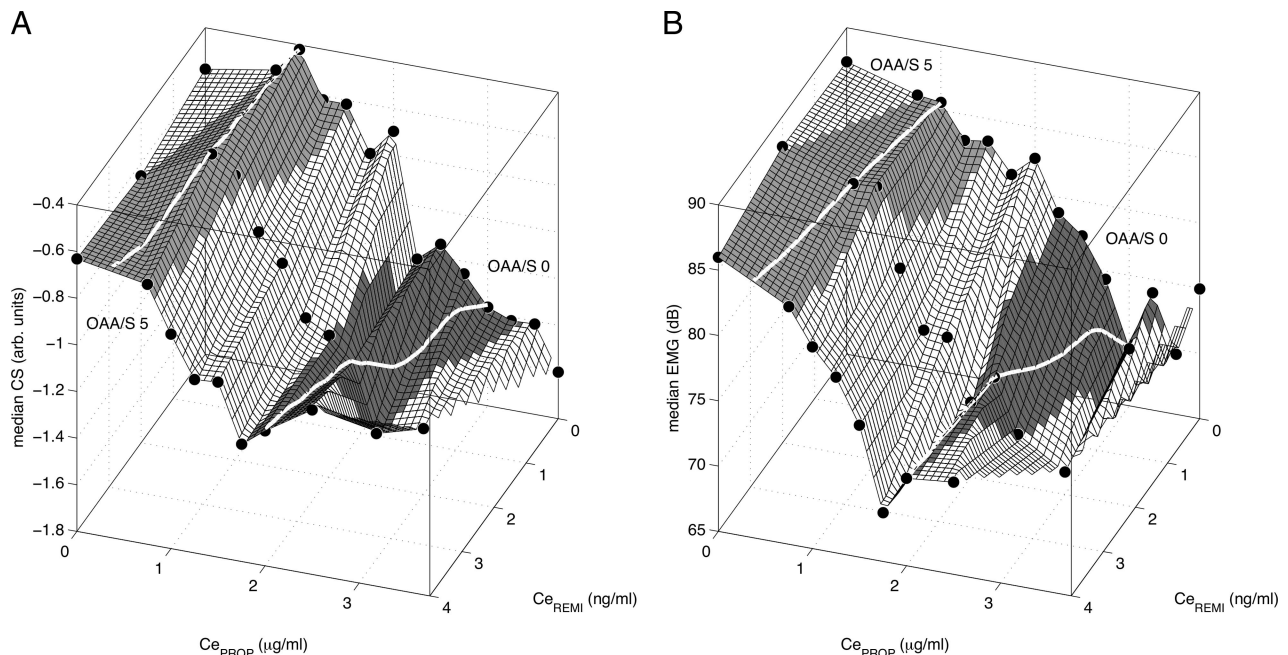


Fig. 5. (A) Median Cortical State (CS) versus target remifentanyl ($C_{e,REMI}$) and propofol ($C_{e,PROP}$) effect-site concentrations. Mesh represents the linearly interpolated surface through data (filled circles). Interquartile (25th–75th percentile) ranges of target propofol concentration for a given Observer's Assessment of Alertness/Sedation (OAA/S) level and remifentanyl level are shown as shaded (OAA/S 5 = light gray, OAA/S 0 = dark gray) patches on the interpolated surface. Median values correspond to the respective solid white line. This surface together with the shaded interquartile ranges indicates that for a fixed OAA/S level and target propofol concentration, CS remains unchanged with increasing $C_{e,REMI}$. (B) Median electromyographic (EMG) activity versus $C_{e,REMI}$ and $C_{e,PROP}$. All other details as for A.

variability. In contrast, CS is found to be independent of remifentanyl level, because the optimum hierarchical linear model does not depend on target remifentanyl concentration. In further contrast to CI, some of the level 2 random effects for CS were significant, allowing us to infer that individual level differences were making some contribution to CS variability.

Prediction Probability (P_k) and Spearman ρ for Clinically Assessed Levels of Sedation

Prediction probability (P_k) and Spearman ρ are measures of ordinal association and provide information regarding how well quantitative measures of sedative state correlate with clinically relevant endpoints. Tables 5–7 show P_k and ρ .

Table 3. Results of Hierarchical Linear Modeling of the Effects of Target Remifentanyl and Propofol Effect-site Concentration on Cortical Input (CI)

Fixed Effect	Coefficient	SE (Robust)	<i>t</i> Ratio	<i>P</i> Value
β_0				
γ_{00}	162.569	8.065	20.079	<0.0005
β_1				
γ_{11}	−17.500	2.080	−9.641	<0.0005
β_2				
γ_{20}	40.091	5.673	6.981	<0.0005
β_3				
γ_{30}	−8.941	1.588	−5.758	<0.0005
Random Effect	Variance Component	<i>df</i>	Chi-square	
u_0	2461.937	35	141.125	<0.0005
u_1	1902.514	35	40.351	0.245
u_2	965.518	35	37.101	0.372
u_3	23.420	35	38.072	0.331
ϵ	548.473			

Estimated effects of target remifentanyl ($C_{e,REMI}$) and propofol ($C_{e,PROP}$) effect-site concentrations on CI for the model $CI = \gamma_{00} + \gamma_{11}R \times P + \gamma_{20}P^2 + \gamma_{30}P^3 + u_0 + u_1P + u_2P^2 + u_3P^3 + \epsilon$, where P and R are $C_{e,PROP}$ and $C_{e,REMI}$, respectively, and ϵ and u_n are normally distributed error terms. Because $\gamma_{11} < 0$ and the associated $p < 0.0005$, we conclude that CI is significantly negatively correlated with the product of $C_{e,REMI}$ and $C_{e,PROP}$. All possible residual covariance terms were used for the level 2 modeling (data not shown).

df = degrees of freedom.

Table 4. Results of Hierarchical Linear Modeling of the Effects of Target Remifentanyl and Propofol Effect-site Concentration on Cortical State (CS)

Fixed Effect	Coefficient	SE (Robust)	t Ratio	P Value
β_0				
γ_{00}	-0.588	0.050	-11.754	<0.0005
β_1				
γ_{10}	0.381	0.102	3.737	0.001
β_2				
γ_{20}	-0.559	0.078	-7.162	<0.0005
β_3				
γ_{30}	0.106	0.016	6.648	<0.0005
Random Effect	Variance Component	df	Chi-square	
u_0	0.241	35	105.646	<0.0005
u_1	0.445	35	48.715	0.061
u_2	0.363	35	62.983	0.003
u_3	0.078	35	73.039	<0.0005
ϵ	0.026			

Estimated effects of target remifentanyl ($C_{e,REMI}$) and propofol ($C_{e,PROP}$) effect-site concentrations on CS for the model $CS = \gamma_{00} + \gamma_{10}P + \gamma_{20}P^2 + \gamma_{30}P^3 + u_0 + u_1P + u_2P^2 + u_3P^3 + \epsilon$, where P and R are $C_{e,PROP}$ and $C_{e,REMI}$, respectively, and ϵ and u_n are normally distributed error terms. This optimal model allows us to conclude that CS depends significantly on $C_{e,PROP}$ but is independent of $C_{e,REMI}$. All possible residual covariance terms were used for the level 2 modeling (data not shown).

df = degrees of freedom.

These tables show measures of ordinal association calculated at all OAA/S levels, OAA/S levels 0 and 5, and dichotomized levels, respectively. These tables reveal that CI, and to a lesser extent RMS, are not predictive of the level of sedation, whereas CS and the electromyogram are highly predictive of sedative state. Therefore, we can conclude that CS represents a meaningful measure of the hypnotic state, whereas CI is essentially uncorrelated with the level of sedation. It is worth noting that the P_k values obtained for CS and the electromyogram are in the same range as those obtained in previous studies using other indices of depth of anesthesia, such as the BIS and State Entropy and Response Entropy indices.

Discussion

The electroencephalographic monitoring of anesthetic depth has well and truly become a part of standard anesthetic practice; it may become an important component of standard-of-care patient monitoring during surgery in a manner similar to that of pulse oximetry.³⁶ However, unlike pulse oximetry, the physiologic underpinnings of electroencephalographic monitoring remain somewhat obscure. For example, doubts

remain about whether processed electroencephalographic measures are characterizing the response of brain electrical activity to anesthetic effect or are merely measuring the effects of these agents in ameliorating tonic electromyographic activity.³⁷ This is arguably due in large part to the fact that the physiologic mechanisms responsible for the generation of rhythmic activity in the electroencephalogram remain unresolved. As a consequence, all processed electroencephalographic measures of depth of anesthesia have had to depend on the application of a range of heuristic, and thus physiologically arbitrary, criteria. This physiologically nonspecific “black-box” analysis can be argued to underlie the current inability of the BIS and other processed measures to detect, and thus monitor, a range of anesthetic agents that include the opioids and nitrous oxide. Therefore, the development of better physiologically motivated methods for the analysis and characterization of electroencephalographic activity can be reasonably speculated to result in more sensitive and specific methods for monitoring brain state during anesthesia. In this article, we have evaluated this proposition using a physiologically motivated linear time series analysis method^{13,16} and

Table 5. Prediction Probability (P_k) and Spearman ρ , plus the Respective SE, Calculated over All OAA/S Levels

	CI		CS		RMS		EMG	
	P_k (SE)	ρ (SE)	P_k (SE)	ρ (SE)	P_k (SE)	ρ (SE)	P_k (SE)	ρ (SE)
Remi0	0.709 (0.029)	-0.489 (0.060)	0.838 (0.022)	0.724 (0.045)	0.835 (0.022)	-0.726 (0.044)	0.785 (0.025)	0.636 (0.052)
Remi2	0.533 (0.045)	-0.077 (0.108)	0.826 (0.024)	0.744 (0.045)	0.686 (0.038)	-0.441 (0.086)	0.836 (0.026)	0.758 (0.050)
Remi4	0.649 (0.048)	0.356 (0.109)	0.763 (0.030)	0.625 (0.061)	0.518 (0.050)	-0.041 (0.119)	0.850 (0.031)	0.772 (0.056)
All	0.527 (0.025)	-0.063 (0.059)	0.814 (0.014)	0.707 (0.028)	0.651 (0.023)	-0.351 (0.053)	0.819 (0.015)	0.712 (0.031)

CI = Cortical Input; CS = Cortical State; EMG = electromyogram; OAA/S = Observer's Assessment of Alertness/Sedation; Remi0 = patient group receiving no remifentanyl; Remi2 = patient group receiving 2 ng/ml remifentanyl; Remi4 = patient group receiving 4 ng/ml remifentanyl; RMS = root mean square electroencephalogram amplitude.

Table 6. Prediction Probability (P_k) and Spearman ρ , plus the Respective SE, Calculated over OAA/S Levels 5 (Responds Readily) and 0 (No Response to Painful Stimulus)

	CI		CS		RMS		EMG	
	P_k (SE)	ρ (SE)	P_k (SE)	ρ (SE)	P_k (SE)	ρ (SE)	P_k (SE)	ρ (SE)
Remi0	0.858 (0.053)	-0.517 (0.088)	1.000 (0.000)	0.722 (0.063)	0.992 (0.009)	-0.710 (0.063)	0.995 (0.006)	0.715 (0.063)
Remi2	0.642 (0.097)	-0.232 (0.158)	0.997 (0.004)	0.812 (0.046)	0.914 (0.054)	-0.677 (0.096)	1.000 (0.000)	0.817 (0.046)
Remi4	0.765 (0.085)	0.439 (0.142)	0.962 (0.029)	0.765 (0.063)	0.542 (0.110)	-0.069 (0.182)	1.000 (0.000)	0.828 (0.044)
All	0.550 (0.063)	-0.078 (0.099)	0.991 (0.006)	0.771 (0.032)	0.787 (0.053)	-0.451 (0.085)	0.998 (0.002)	0.782 (0.032)

CI = Cortical Input; CS = Cortical State; EMG = electromyogram; OAA/S = Observer's Assessment of Alertness/Sedation; Remi0 = patient group receiving no remifentanyl; Remi2 = patient group receiving 2 ng/ml remifentanyl; Remi4 = patient group receiving 4 ng/ml remifentanyl; RMS = root mean square electroencephalogram amplitude.

have shown, in contrast to existing processed measures, that the effect of remifentanyl on frontally recorded spontaneous electroencephalograms can be dissociated from that of propofol. The existing literature paints a complex picture of the effects of opioids on the electroencephalogram. For example, the sole administration of remifentanyl is often reported to cause a dose-dependent slowing of the electroencephalogram, but typically only for levels much higher than those used in the current study.^{20,38–40} In contrast, remifentanyl, when administered with propofol, is generally reported to have no effect on derived electroencephalographic parameters such as the BIS^{41–48} but is occasionally found to result in an increase^{49,50} or a decrease^{51–56} in such derived measures of hypnosis. The contention that opioids such as remifentanyl produce a predictable dose-dependent slowing of the electroencephalogram is not borne out by our own results, because CS remains unchanged to variations in the level of remifentanyl. Although CS was unaffected by remifentanyl, it nevertheless remains a possibility that CI and RMS were affected indirectly due to increased arterial carbon dioxide levels. In experimentally induced hypercapnia in animals, increased arterial carbon dioxide levels are correlated with reductions in resting amplitude of the electroencephalogram.^{57,58} Although end-tidal carbon dioxide levels were within clinical limits in our study, future studies involving CS and CI should have these levels percutaneously measured to ensure that increased carbon dioxide levels are not acting as a confounding influence.

In contrast to the BIS⁵⁹ and State Entropy and Response Entropy⁶⁰ indices, our method does not depend on quantifying the changes in either the nonlinearity or complexity of

brain activity that are hypothesized to attend anesthetic action. We have found, somewhat surprisingly, that putative measures of hypnosis and analgesic drug action can be defined based on a relatively standard but physiologically constrained linear signal analysis technique. The constrained use of this linear technique has emerged from a detailed theory for the rhythmogenesis of the electroencephalogram¹² that has been successfully applied to modeling the effects of anesthetics on brain electrical activity.^{11,14} Therefore, the possibility exists that estimated ARMA parameters (see Eq. 1) may be theoretically more specifically linked to the central modes and sites of drug action, thus suggesting additional methods by which anesthetic action may be better monitored. Because the computational demands of the fixed-order ARMA method are relatively modest, it can easily be calculated in real time using dedicated hardware similar to that used in BIS[®] monitoring.

The derived electroencephalogram measures of CS (a measure of the responsiveness of cortex) and CI (a measure of the magnitude of cortical input) were shown to respond differentially to target propofol and remifentanyl effect-site concentrations. In particular, it was found, on the basis of hierarchical linear modeling, that CI decreased significantly with increasing target remifentanyl concentration, whereas CS was found to be statistically independent of variations in effect-site remifentanyl level. Both CS and CI responded to effect-site propofol levels: CS monotonically decreased, whereas CI responded nonuniformly depending on remifentanyl level; propofol was agonistic at low remifentanyl levels but antagonistic at high remifentanyl levels. The reduction in CI is consistent with the reported effects of remifentanyl in atten-

Table 7. Prediction Probability (P_k) and Spearman ρ , plus the Respective SE, Calculated over Aggregated OAA/S Levels 5–3 (Responsive) and Levels 2–0 (Nonresponsive)

	CI		CS		RMS		EMG	
	P_k (SE)	ρ (SE)	P_k (SE)	ρ (SE)	P_k (SE)	ρ (SE)	P_k (SE)	ρ (SE)
Remi0	0.725 (0.047)	-0.320 (0.069)	0.951 (0.017)	0.641 (0.045)	0.939 (0.021)	-0.625 (0.048)	0.911 (0.027)	0.585 (0.051)
Remi2	0.550 (0.073)	0.075 (0.111)	0.928 (0.026)	0.654 (0.054)	0.691 (0.069)	-0.292 (0.106)	0.960 (0.018)	0.703 (0.049)
Remi4	0.653 (0.073)	0.246 (0.117)	0.836 (0.046)	0.540 (0.078)	0.578 (0.074)	-0.125 (0.118)	0.883 (0.049)	0.616 (0.084)
All	0.508 (0.043)	0.012 (0.065)	0.916 (0.016)	0.628 (0.031)	0.676 (0.040)	-0.265 (0.061)	0.919 (0.019)	0.632 (0.035)

CI = Cortical Input; CS = Cortical State; EMG = electromyogram; OAA/S = Observer's Assessment of Alertness/Sedation; Remi0 = patient group receiving no remifentanyl; Remi2 = patient group receiving 2 ng/ml remifentanyl; Remi4 = patient group receiving 4 ng/ml remifentanyl; RMS = root mean square electroencephalogram amplitude.

uating a range of somatosensory and auditory-evoked potentials,^{10,61} thus providing further weight to the notion that this derived measure is indeed quantifying some aspect of input to cortex. At present, we do not know why propofol alone increases CI, but we can speculate that it is due to its differential effects on a range of subcortical structures that contribute to cortical input. Because propofol enhances inhibitory activity through the potentiation of γ -aminobutyric acid receptor subtype A activity, it can result in the inhibition or disinhibition of activity depending on how it differentially modulates inhibitory activity terminating on excitatory and inhibitory neurons. Indeed, there is good evidence to suggest that such differential binding is responsible for the characteristic increase in β (13–30 Hz) band electroencephalogram activity seen with most sedatives and anesthetics.¹³

We chose to model drug effect using hierarchical linear modeling,³⁵ rather than the more familiar bivariate sigmoidal E_{\max} models,^{51,62–64} principally because a bivariate sigmoidal E_{\max} model contains insufficient degrees of freedom to account for the dependency of CI and CS on target drug concentrations. Although a great deal of pharmacodynamic effects and interactions are plausibly based on the paradigm of molecular mass action, there is no *a priori* reason to believe that our processed measures of CS and CI will adhere to such a principle. CS and CI characterize the collective activity of many thousands of neurons, interacting over many temporal and spatial scales, and thus the steps between the microscopic details of drug binding and the consequent macroscopic physiologic effect are simply too complicated to be accounted for by a uni- or bivariate monotonic dose–response relationship. Indeed, even in the study of much simpler pharmacologic processes, sigmoid dose–response relationships, although common, are not universal—linear, linear–quadratic, log–linear, and exponential best–fit relationships are also found.⁶³ Although our use of hierarchical linear modeling is not fully general, it is nevertheless more flexible in that it is able to statistically account for the nonuniform dose–response relationship (propofol agonistic at low remifentanyl levels but antagonistic at high remifentanyl levels; see fig. 4A) that we have observed between CI and remifentanyl and propofol concentrations. A further reason for choosing hierarchical linear modeling over sigmoid-based curve-fitting strategies is the issue of the nesting of patient data. The nesting of these data arises because variance and covariance cannot be expected to be distributed uniformly across repeated observations and patients. As far as we are aware, such heteroscedasticity cannot be dealt with by NONMEM.

At this point in time, the optimal response surface models relating CI and CS to drug levels admit of no obvious physiologic interpretation. Nevertheless, they provide clear statistical evidence for the contention that increases in remifentanyl levels are associated with a significant reduction in CI, whereas CS remains unaffected. The significance of our find-

ing of a correlation of CI with remifentanyl level is underscored by the development of the surgical stress index.** The surgical stress index, developed to provide a measure of analgesic efficacy during surgery, is based on a sum of the normalized pulse beat interval and the pulse wave amplitude time series of the photoplethysmogram. A number of studies have shown it to correlate well with target-controlled remifentanyl levels⁶⁵ and the probability of response to a noxious stimulus.⁶⁶ Therefore, given its clear dependence on target remifentanyl level, CI should be compared prospectively with the surgical stress index, under conditions involving noxious surgical stimuli, as a potential additional measure of the nociception–antinociception balance. CI may have a number of specific advantages in that it may reflect both the central and autonomic responses to noxious stimuli. Being able to differentiate the effects of a hypnotic agent and an analgesic agent, as is done here, is a necessary first step toward the development of such an index of analgesic state.

On the basis of our results, we speculate that CS and CI provide “orthogonal” measures of hypnosis and analgesia. In particular, as per our hypothesis, we found CI had a P_k of approximately 0.5, meaning it was no better than chance in predicting sedative state. In contrast, CS had a much higher P_k of approximately 0.8, meaning that it was a meaningfully predictive method of the level of hypnosis as quantified by OAA/S assessment. The values of P_k obtained for CS are in the same range as those seen in similar studies involving other quantitative depth of anesthesia measures such as BIS and State Entropy and Response Entropy indices. Although CI and RMS seemed to be correlated, the P_k of the latter (~ 0.65) was intermediate between that of CI and CS and thus would be neither a good measure of sedative state nor a potential measure of analgesia.^{48,50,55} Because of the lack of any significant correlation between the measures of CS and CI, they may subsequently be found to have utility in guiding clinical decision support during the control and administration of balanced anesthesia.⁶⁷

Denny Myer, Ph.D., Senior Lecturer in Biostatistics, Faculty of Life and Social Sciences, Swinburne University of Technology, Hawthorn, Victoria, Australia, advised on the use of hierarchical linear modeling.

References

1. Bruhn J, Myles PS, Sneyd R, Struys MM: Depth of anaesthesia monitoring: What's available, what's validated and what's next? *Br J Anaesth* 2006; 97:85–94
2. Myles PS, Leslie K, McNeil J, Forbes A, Chan MT: Bispectral index monitoring to prevent awareness during anaesthesia: The B-Aware randomised controlled trial. *Lancet* 2004; 363:1757–63
3. Karalapillai D, Leslie K, Umraniar A, Bjorksten AR: Nitrous oxide and anesthetic requirement for loss of response to command during propofol anesthesia. *Anesth Analg* 2006; 102:1088–93
4. Rampil IJ, Kim JS, Lenhardt R, Negishi C, Sessler DI: Bispectral EEG index during nitrous oxide administration. *ANESTHESIOLOGY* 1998; 89:671–7
5. Soto RG, Smith RA, Zaccaria AL, Miguel RV: The effect of addition of nitrous oxide to a sevoflurane anesthetic on

** Expected to be renamed the Surgical Pleth Index (SPI) by the manufacturer (GE Healthcare, Helsinki, Finland).⁶⁴

- BIS, PSI, and entropy. *J Clin Monit Comput* 2006; 20: 145-50
6. Wong CA, Fragen RJ, Fitzgerald P, McCarthy RJ: A comparison of the SNAP II and BIS XP indices during sevoflurane and nitrous oxide anaesthesia at 1 and 1.5 MAC and at awakening. *Br J Anaesth* 2006; 97:181-6
 7. Glass PS, Bloom M, Kears L, Rosow C, Sebel P, Manberg P: Bispectral analysis measures sedation and memory effects of propofol, midazolam, isoflurane, and alfentanil in healthy volunteers. *ANESTHESIOLOGY* 1997; 86:836-47
 8. Iselin-Chaves IA, El Moalem HE, Gan TJ, Ginsberg B, Glass PS: Changes in the auditory evoked potentials and the bispectral index following propofol or propofol and alfentanil. *ANESTHESIOLOGY* 2000; 92:1300-10
 9. Schmidt GN, Bischoff P, Standl T, Jensen K, Voigt M, Schulte Am Esch J: Narcotrend and bispectral index monitor are superior to classic electroencephalographic parameters for the assessment of anesthetic states during propofol-remifentanil anesthesia. *ANESTHESIOLOGY* 2003; 99:1072-7
 10. Schmidt GN, Scharein E, Siegel M, Muller J, Debener S, Nitzschke R, Engel A, Bischoff P: Identification of sensory blockade by somatosensory and pain-induced evoked potentials. *ANESTHESIOLOGY* 2007; 106:707-14
 11. Bojak I, Liley DT: Modeling the effects of anesthesia on the electroencephalogram. *Phys Rev E Stat Nonlin Soft Matter Phys* 2005; 71:041902
 12. Liley DT, Cadusch PJ, Dafilis MP: A spatially continuous mean field theory of electrocortical activity. *Network* 2002; 13:67-113
 13. Liley DT, Cadusch PJ, Gray M, Nathan PJ: Drug-induced modification of the system properties associated with spontaneous human electroencephalographic activity. *Phys Rev E Stat Nonlin Soft Matter Phys* 2003; 68:051906
 14. Liley DT, Bojak I: Understanding the transition to seizure by modeling the epileptiform activity of general anesthetic agents. *J Clin Neurophysiol* 2005; 22:300-13
 15. Steyn-Ross ML, Steyn-Ross DA, Sleight JW, Liley DT: Theoretical electroencephalogram stationary spectrum for a white-noise-driven cortex: Evidence for a general anesthetic-induced phase transition. *Phys Rev E Stat Phys Plasmas Fluids Relat Interdiscip Topics* 1999; 60:7299-311
 16. Liley DT, Leslie K, Sinclair NC, Feckie M: Dissociating the effects of nitrous oxide on brain electrical activity using fixed order time series modeling. *Comput Biol Med* 2008; 38:1121-30
 17. Ferenets R, Vanluchene A, Lipping T, Heyse B, Struys MM: Behavior of entropy/complexity measures of the electroencephalogram during propofol-induced sedation: Dose-dependent effects of remifentanil. *ANESTHESIOLOGY* 2007; 106:696-706
 18. Schnider TW, Minto CF, Gambus PL, Andresen C, Goodale DB, Shafer SL, Youngs EJ: The influence of method of administration and covariates on the pharmacokinetics of propofol in adult volunteers. *ANESTHESIOLOGY* 1998; 88: 1170-82
 19. Schnider TW, Minto CF, Shafer SL, Gambus PL, Andresen C, Goodale DB, Youngs EJ: The influence of age on propofol pharmacodynamics. *ANESTHESIOLOGY* 1999; 90:1502-16
 20. Minto CF, Schnider TW, Egan TD, Youngs E, Lemmens HJ, Gambus PL, Billard V, Hoke JF, Moore KH, Hermann DJ, Muir KT, Mandema JW, Shafer SL: Influence of age and gender on the pharmacokinetics and pharmacodynamics of remifentanil: I. Model development. *ANESTHESIOLOGY* 1997; 86:10-23
 21. Minto CF, Schnider TW, Shafer SL: Pharmacokinetics and pharmacodynamics of remifentanil: II. Model application. *ANESTHESIOLOGY* 1997; 86:24-33
 22. Minto CF, Schnider TW, Gregg KM, Henthorn TK, Shafer SL: Using the time of maximum effect site concentration to combine pharmacokinetics and pharmacodynamics. *ANESTHESIOLOGY* 2003; 99:324-33
 23. Struys MM, De Smet T, Depoorter B, Versichelen LF, Mortier EP, Dumortier FJ, Shafer SL, Rolly G: Comparison of plasma compartment *versus* two methods for effect compartment-controlled target-controlled infusion for propofol. *ANESTHESIOLOGY* 2000; 92:399-406
 24. Lilliefors H: On the Kolmogorov-Smirnov test for normality with mean and variance unknown. *J Am Stat Assoc* 1967; 62:399-402
 25. Jeleazcov C, Fechner J, Schwilden H: Electroencephalogram monitoring during anesthesia with propofol and alfentanil: The impact of second order spectral analysis. *Anesth Analg* 2005; 100:1365-9
 26. Hagihira S, Takashina M, Mori T, Mashimo T: EEG during anesthesia is not a linear random process. *Anesth Analg* 2006; 102:966; author reply 966-7
 27. Schack B, Krause W: Dynamic power and coherence analysis of ultra short-term cognitive processes—A methodical study. *Brain Topogr* 1995; 8:127-36
 28. Schwilden H, Jeleazcov C: Does the EEG during isoflurane/alfentanil anesthesia differ from linear random data? *J Clin Monit Comput* 2002; 17:449-57
 29. Stam CJ, Pijn JP, Suffczynski P, Lopes da Silva FH: Dynamics of the human alpha rhythm: Evidence for non-linearity? *Clin Neurophysiol* 1999; 110:1801-13
 30. Tseng S-Y, Chen R-C, Chong F-K, Kuo T-S: Evaluation of parametric methods in EEG signal analysis. *Med Eng Phys* 1995; 17:71-8
 31. Broersen PMT: Automatic Autocorrelation and Spectral Analysis. London, Springer, 2006; pp xii, 293
 32. Broersen P: Automatic spectral analysis with time series models. *IEEE Trans Instrum Meas* 2002; 51:211-6
 33. Smith WD, Dutton RC, Smith NT: A measure of association for assessing prediction accuracy that is a generalization of non-parametric ROC area. *Stat Med* 1996; 15:1199-215
 34. Smith WD, Dutton RC, Smith NT: Measuring the performance of anesthetic depth indicators. *ANESTHESIOLOGY* 1996; 84:38-51
 35. Raudenbush SW, Bryk AS: Hierarchical Linear Models: Applications and Data Analysis Methods, 2nd edition. Thousand Oaks, CA, Sage, 2002
 36. Bowdle TA: Depth of anesthesia monitoring. *Anesthesiol Clin* 2006; 24:793-822
 37. Messner M, Beese U, Romstock J, Dinkel M, Tschaikowsky K: The bispectral index declines during neuromuscular block in fully awake persons. *Anesth Analg* 2003; 97:488-91.
 38. Egan TD, Minto CF, Hermann DJ, Barr J, Muir KT, Shafer SL: Remifentanil *versus* alfentanil: Comparative pharmacokinetics and pharmacodynamics in healthy adult male volunteers. *ANESTHESIOLOGY* 1996; 84:821-33
 39. Kang SH, Poynton MR, Kim KM, Lee H, Kim DH, Lee SH, Bae KS, Linares O, Kern SE, Noh GJ: Population pharmacokinetic and pharmacodynamic models of remifentanil in healthy volunteers using artificial neural network analysis. *Br J Clin Pharmacol* 2007; 64:3-13
 40. Noh GJ, Kim KM, Jeong YB, Jeong SW, Yoon HS, Jeong SM, Kang SH, Linares O, Kern SE: Electroencephalographic approximate entropy changes in healthy volunteers during remifentanil infusion. *ANESTHESIOLOGY* 2006; 104:921-32
 41. Barvais L, Engelman E, Eba JM, Coussaert E, Cantraine F, Kenny GN: Effect site concentrations of remifentanil and pupil response to noxious stimulation. *Br J Anaesth* 2003; 91:347-52
 42. Guignard B, Menigaux C, Dupont X, Fletcher D, Chauvin M: The effect of remifentanil on the bispectral index change and hemodynamic responses after orotracheal intubation. *Anesth Analg* 2000; 90:161-7

43. Hoymork SC, Raeder J, Grimsø B, Steen PA: Bispectral index, predicted and measured drug levels of target-controlled infusions of remifentanyl and propofol during laparoscopic cholecystectomy and emergence. *Acta Anaesthesiol Scand* 2000; 44:1138-44
44. Kortelainen J, Koskinen M, Mustola S, Seppanen T: Remifentanyl modifies the relation of electroencephalographic spectral changes and clinical endpoints in propofol anesthesia. *ANESTHESIOLOGY* 2008; 109:198-205
45. Schmidt GN, Bischoff P, Standl T, Lankenau G, Hilbert M, Schulte Am Esch J: Comparative evaluation of Narcotrend, bispectral index, and classical electroencephalographic variables during induction, maintenance, and emergence of a propofol/remifentanyl anesthesia. *Anesth Analg* 2004; 98:1346-53
46. Schraag S, Flaschar J, Schleyer M, Georgieff M, Kenny GN: The contribution of remifentanyl to middle latency auditory evoked potentials during induction of propofol anesthesia. *Anesth Analg* 2006; 103:902-7
47. Struys MM, Vereecke H, Moerman A, Jensen EW, Verhaeghen D, De Neve N, Dumortier FJ, Mortier EP: Ability of the bispectral index, autoregressive modelling with exogenous input-derived auditory evoked potentials, and predicted propofol concentrations to measure patient responsiveness during anesthesia with propofol and remifentanyl. *ANESTHESIOLOGY* 2003; 99:802-12
48. Vanluchene AL, Struys MM, Heyse BE, Mortier EP: Spectral entropy measurement of patient responsiveness during propofol and remifentanyl: A comparison with the bispectral index. *Br J Anaesth* 2004; 93:645-54
49. Lysakowski C, Dumont L, Pellegrini M, Clergue F, Tassonyi E: Effects of fentanyl, alfentanil, remifentanyl and sufentanil on loss of consciousness and bispectral index during propofol induction of anaesthesia. *Br J Anaesth* 2001; 86:523-7
50. Manyam SC, Gupta DK, Johnson KB, White JL, Pace NL, Westenskow DR, Egan TD: When is a bispectral index of 60 too low? Rational processed electroencephalographic targets are dependent on the sedative-opioid ratio. *ANESTHESIOLOGY* 2007; 106:472-83
51. Bouillon TW, Bruhn J, Radulescu L, Andresen C, Shafer TJ, Cohane C, Shafer SL: Pharmacodynamic interaction between propofol and remifentanyl regarding hypnosis, tolerance of laryngoscopy, bispectral index, and electroencephalographic approximate entropy. *ANESTHESIOLOGY* 2004; 100:1353-72
52. Ferreira DA, Nunes CS, Antunes LM, Santos IA, Lobo F, Casal M, Ferreira L, Amorim P: The effect of a remifentanyl bolus on the bispectral index of the EEG (BIS) in anaesthetized patients independently from intubation and surgical stimuli. *Eur J Anaesthesiol* 2006; 23:305-10
53. Koitabashi T, Johansen JW, Sebel PS: Remifentanyl dose/electroencephalogram bispectral response during combined propofol/regional anesthesia. *Anesth Analg* 2002; 94:1530-3
54. Ropcke H, Konen-Bergmann M, Cuhls M, Bouillon T, Hoeft A: Propofol and remifentanyl pharmacodynamic interaction during orthopedic surgical procedures as measured by effects on bispectral index. *J Clin Anesth* 2001; 13:198-207
55. Schmidt GN, Bischoff P, Standl T, Hellstern A, Teuber O, Schulte Esch J: Comparative evaluation of the Datex-Ohmeda S/5 Entropy Module and the Bispectral Index monitor during propofol-remifentanyl anesthesia. *ANESTHESIOLOGY* 2004; 101:1283-90
56. Strachan AN, Edwards ND: Randomized placebo-controlled trial to assess the effect of remifentanyl and propofol on bispectral index and sedation. *Br J Anaesth* 2000; 84:489-90
57. Speckman EJ, Caspers H: The effect of O₂ and CO₂ tensions in the nervous tissue on neuronal activity and DC potentials, *Handbook of Electroencephalography and Clinical Neurophysiology*. Edited by Remond A. Amsterdam, Elsevier, 1974; pp 71-89
58. Speckman EJ, Caspers H: Origin of Cerebral Field Potentials. Stuttgart, Thieme, 1979
59. Rampil IJ: A primer for EEG signal processing in anesthesia. *ANESTHESIOLOGY* 1998; 89:980-1002
60. Viertio-Oja H, Maja V, Sarkela M, Talja P, Tenkanen N, Tolvanen-Laakso H, Paloheimo M, Vakkuri A, Yli-Hankala A, Merilainen P: Description of the Entropy algorithm as applied in the Datex-Ohmeda S/5 Entropy Module. *Acta Anaesthesiol Scand* 2004; 48:154-61
61. Crabb I, Thornton C, Konieczko KM, Chan A, Aquilina R, Frazer N, Dore CJ, Newton DE: Remifentanyl reduces auditory and somatosensory evoked responses during isoflurane anaesthesia in a dose-dependent manner. *Br J Anaesth* 1996; 76:795-801
62. Berenbaum MC: What is synergy? *Pharmacol Rev* 1989; 41:93-141
63. Berenbaum MC: Re: W. R. Greco et al., Application of a new approach for the quantitation of drug synergism to the combination of *cis*-diamminedichloroplatinum and 1-beta-D-arabinofuranosylcytosine. *Cancer Res* 1990; 50:5318-27. *Cancer Res* 1992; 52:4558-60; author reply 4561-5
64. Greco WR, Bravo G, Parsons JC: The search for synergy: A critical review from a response surface perspective. *Pharmacol Rev* 1995; 47:331-85
65. Struys MM, Vanpeteghem C, Huiku M, Uutela K, Blyart NB, Mortier EP: Changes in a surgical stress index in response to standardized pain stimuli during propofol-remifentanyl infusion. *Br J Anaesth* 2007; 99:359-67
66. Gruenewald M, Meybohm P, Ilies C, Hocker J, Hanss R, Scholz J, Bein B: Influence of different remifentanyl concentrations on the performance of the surgical stress index to detect a standardized painful stimulus during sevoflurane anaesthesia. *Br J Anaesth* 2009; 103:586-93
67. Huiku M, Uutela K, van Gils M, Korhonen I, Kymäläinen M, Meriläinen P, Paloheimo M, Rantanen M, Takala P, Viertio-Oja H, Yli-Hankala A: Assessment of surgical stress during general anaesthesia. *Br J Anaesth* 2007; 98:447-55